

Flow cytometry data-Analysis and Visualization



ChUG
Cytometry



Challenges in analyzing high-dimensional data

- Historically
 - ↗ manual gating
 - ↘ histograms and biaxial plots
- Manual gating pitfalls-subjective, time consuming, will not work for high-dimensional data)
- Histograms and biaxial plots are almost impossible for high parameter flow cytometry.
- Supervised vs semi-supervised vs unsupervised



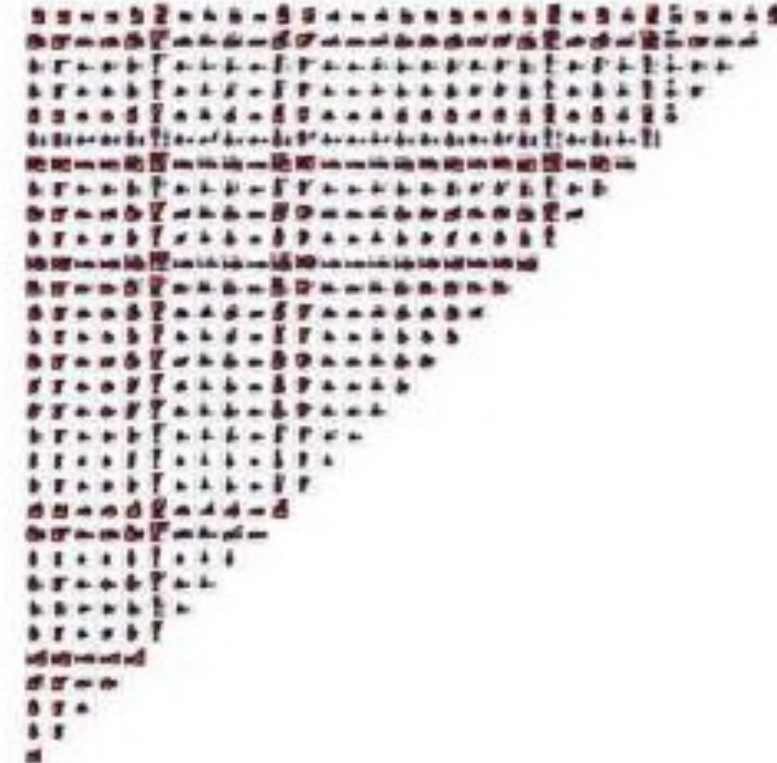
9 parameters

36 plots



32 parameters

496 plots



Bendall et al., Trends Immunol. 2012 Jul; 33(7): 323–332.



Supervised vs unsupervised vs semisupervised

Supervised, useful for hypothesis testing

- manual gating
- visual QC with 2 and 3D plots
- Abundance, MFI, statistics etc.

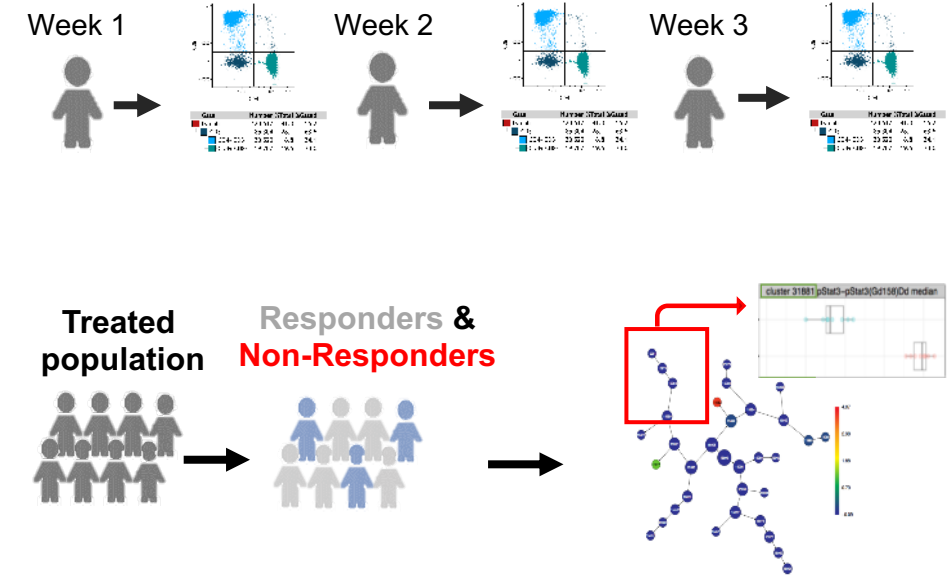
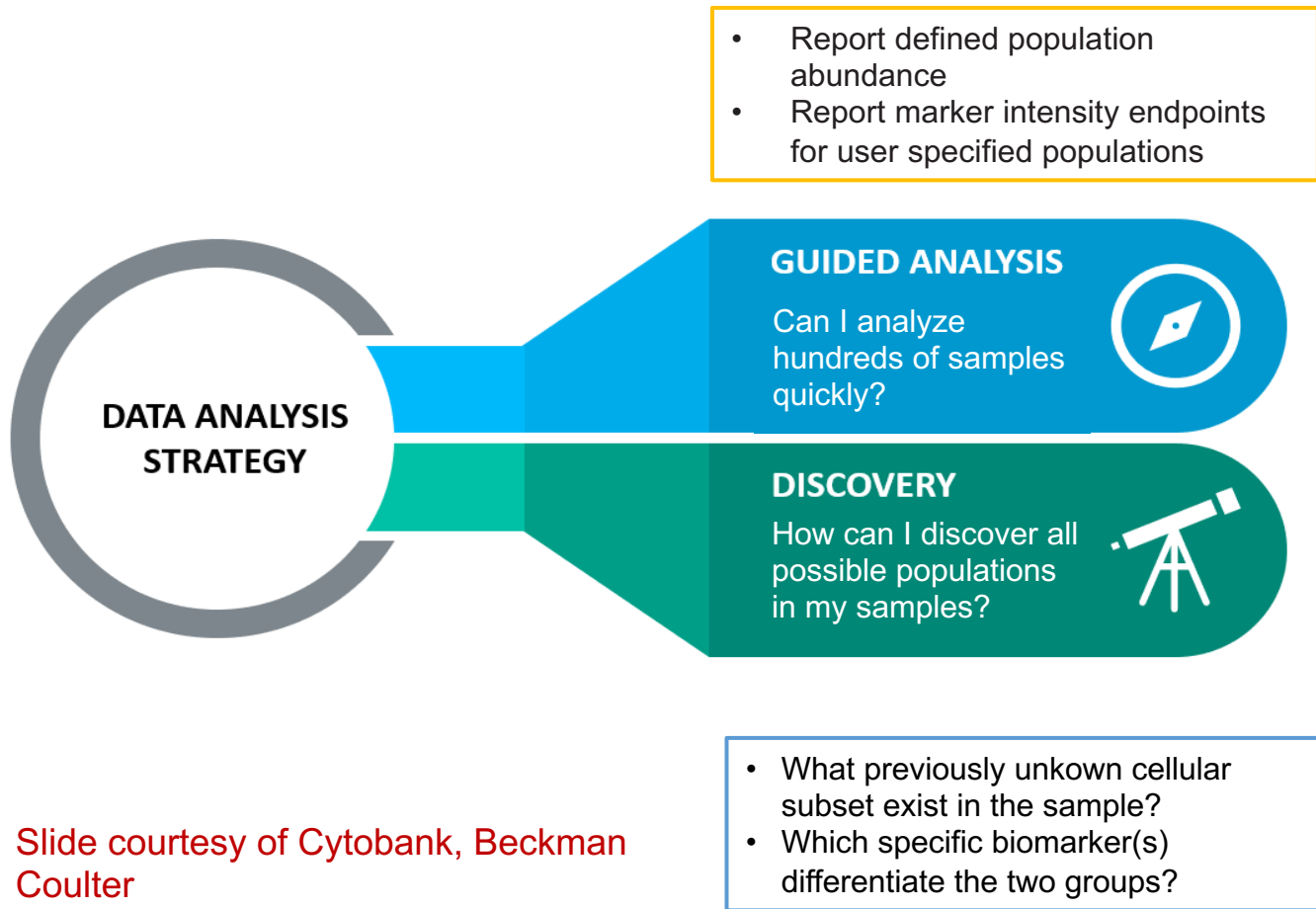
Unsupervised, approach is better suited for exploratory analysis and hypothesis-generating experiments

- Dimensionality reduction
- Clustering
- HeatMaps

With well designed multi-parameter panel of antibodies this two approaches can be combined in one experiment. We postulate that most experiments would benefit from dual approach and will be demonstrated today.



The Best Data Analysis Approach Depends on Your Experiment



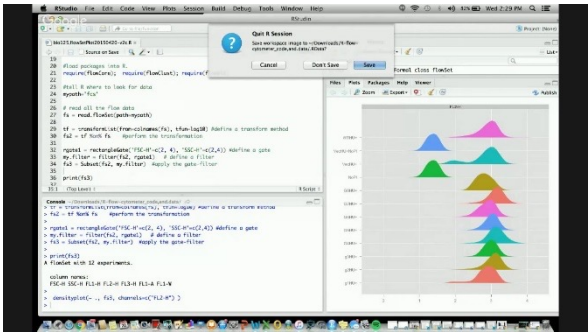
Slide courtesy of Cytobank, Beckman Coulter



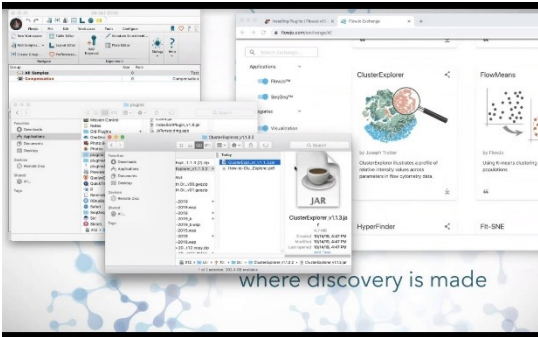
Some of the common platforms

Desktop

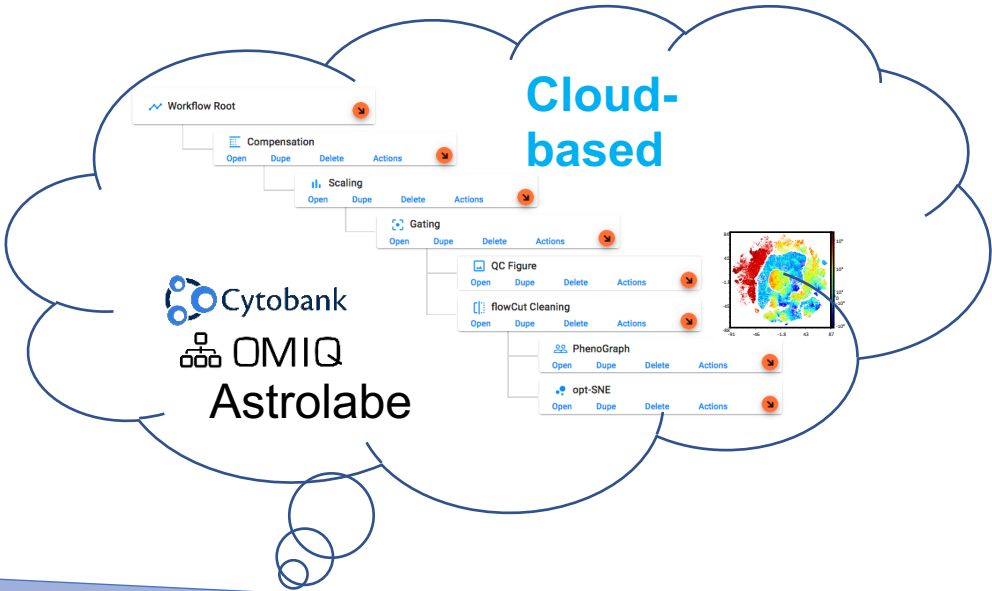
R



FlowJo
FCS Express
Kaluza



Cloud-based



Learning curve

Computational
resources

Flexibility

\$\$\$ Cost



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Cloud based analysis vs Desktop based

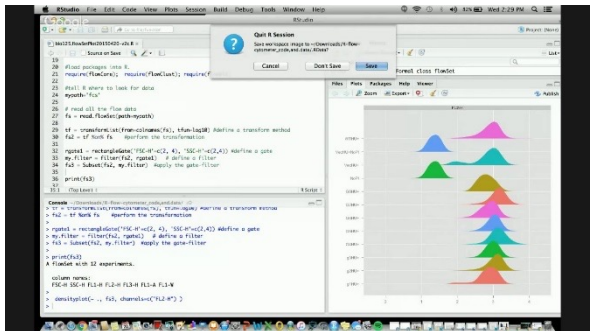
- Easier management of large data
- Easier sharing of data and analysis
- Data is secure
- Cloud computing
- Desktop- Comfort Zone
- Need high-efficiency computer
- Most likely resort to the cloud for data sharing



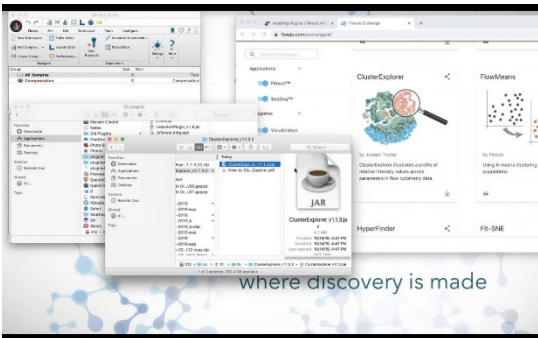
Some of the common platforms

R

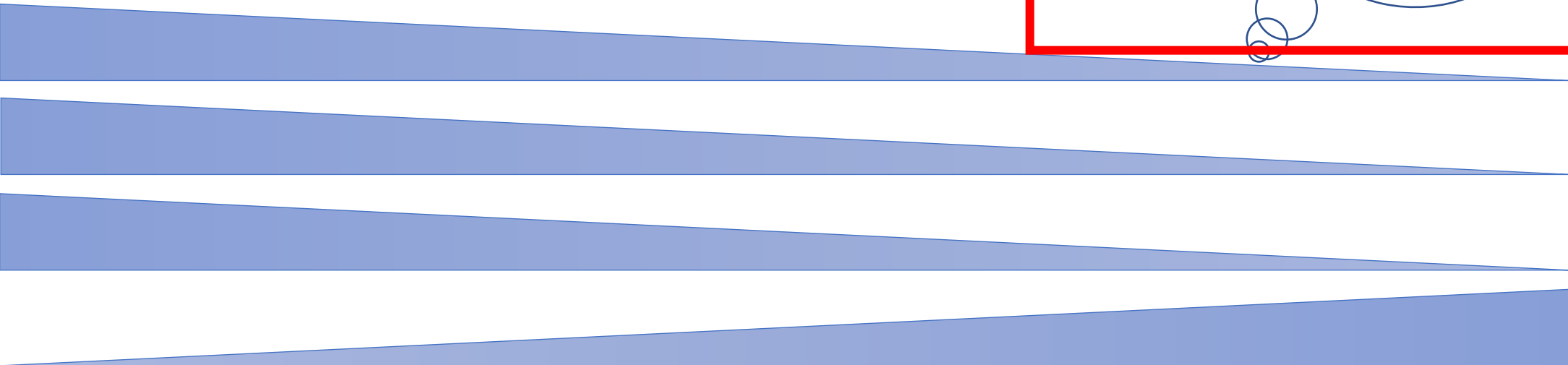
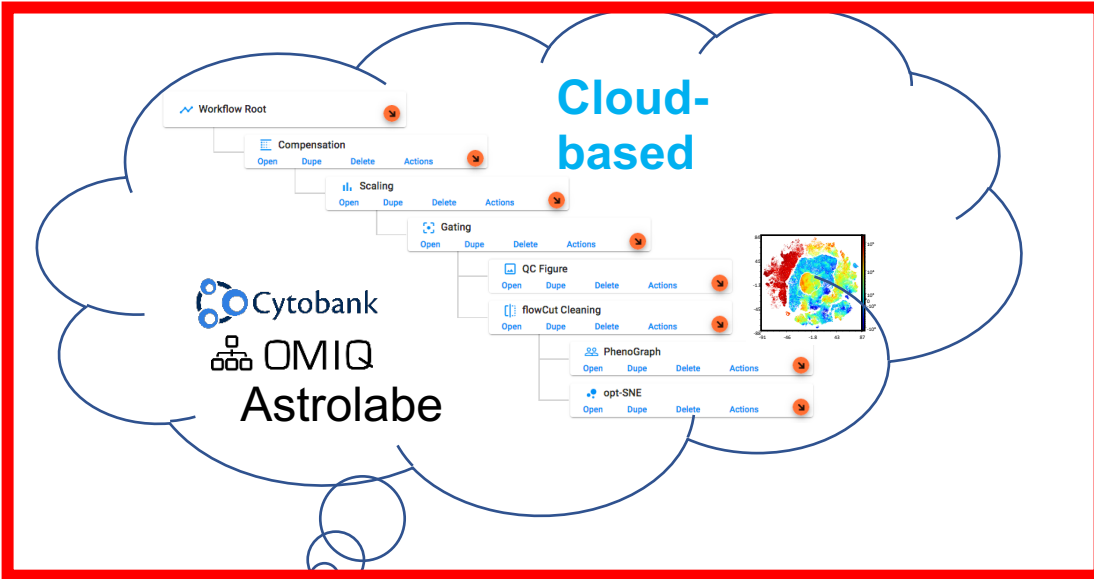
Desktop



FlowJo
FCS Express
Kaluza



Cloud-based



Learning curve

Computational resources

Flexibility

\$\$\$ Cost



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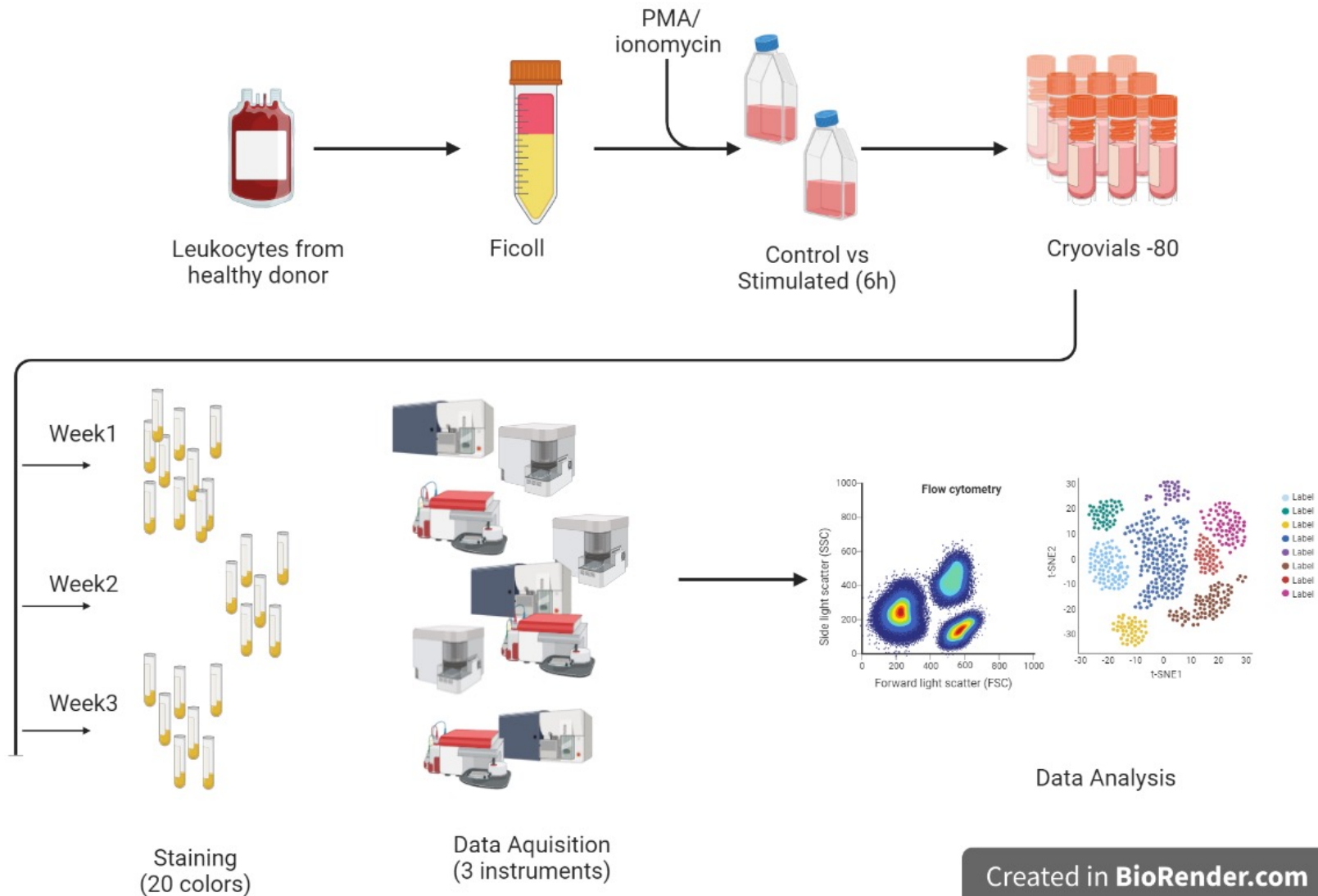


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Experiment outline



Created in **BioRender.com** **bio**



Antibodies donated by:



BD Biosciences

ThermoFisher
S C I E N T I F I C



<https://www.chugcytometry.com>



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Antibodies donated by:



BD Biosciences

I would be happy to offer a one-time 50% promo valid for 2 weeks (excluding OptiBuild) to help academic researchers They would just need to reach out to me directly for a quote.



Shari Sanders

Research Reagent Specialist

Single Cell Multiomics (scM)

BD Biosciences

2350 Qume Drive

San Jose, California, 95131-1807

mobile: 331-302-6716



<https://www.chugcytometry.com>



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Antibodies donated by:



Receive 50% off one order from Oct. 24th-Nov.4th

Attendees of the Advanced Flow Cytometry Workshop can save 50% off one order during Oct. 24th - Nov. 4th when you contact your local FAS, Kim Cardenas, at kcardenas@biolegend.com and mention "ChUG 2022".

GolnVivo™, TotalSeq™ Panels, Equipment and Any Custom products are excluded

*Recipients must be from Illinois and have registered and attended the workshop

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[biolegend.com](https://www.biolegend.com)
07-0162-00



<https://www.chugcytometry.com>



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Antibodies donated by:

ThermoFisher
S C I E N T I F I C

I am happy to let you know I was able to create a promotion that will give all ChUG attendees **50% off all Flow Antibodies and Reagents** (comp beads, Live/Dead Dyes, Molecular Probe Dyes, etc...) and this promotion will extend until December 15th, 2022

Emily Hennessy

Technical Sales Specialist

Flow Cytometry Reagents

MN, WI, IA, IL, MI, IN

Thermo Fisher Scientific

Mobile +1 978-766-9552

Emily.Hennessy@Thermofisher.com



<https://www.chugcytometry.com>



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Antibodies donated by:

The Bio-Rad logo consists of the words "BIO-RAD" in a bold, white, sans-serif font. The letters are slightly italicized. The text is set against a green background that is shaped like a rounded rectangle with a slight gradient.

Yes! 50% off the first three StarBright antibodies per customer until the end of this year, and I will do 50% off any other flow antibody for the rest of the year (up to three per customer). They are welcome to reach out to me directly and you are welcome to provide them with my contact information.

Don Boyer

Senior Account Executive | Cell Biology
Southeast and Midwest Regions

Bio-Rad, Inc.

Email: Donald_Boyer@Bio-Rad.com

Cell: (510) 806-5779



<https://www.chugcytometry.com>



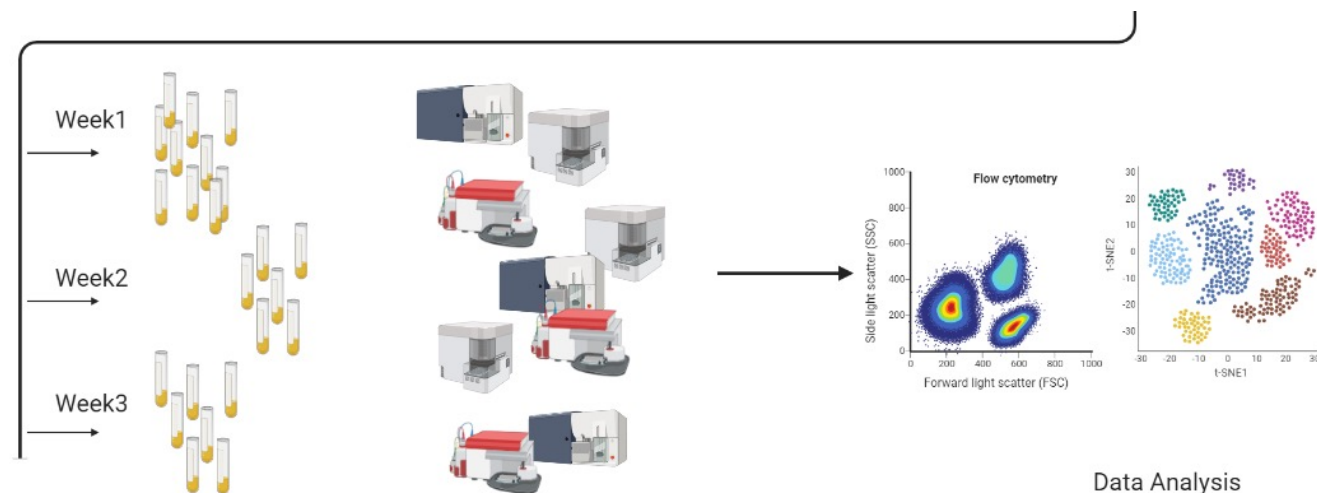
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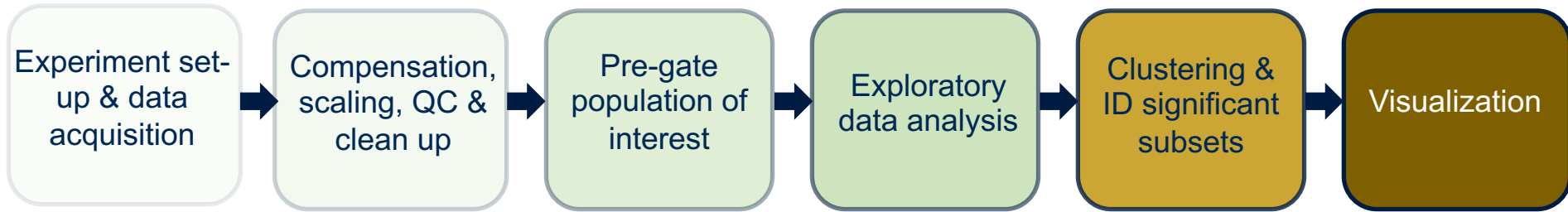
ChUG Cytometry

Using this data set we may be able to:

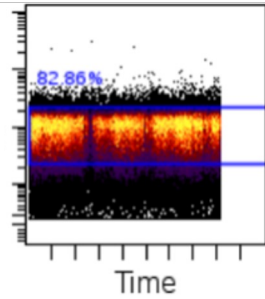
- Evaluate different analysis methods
- Observe, quantify and correct batch effect
- Take a closer look at spectral unmixing methods
- Compare data from different instruments



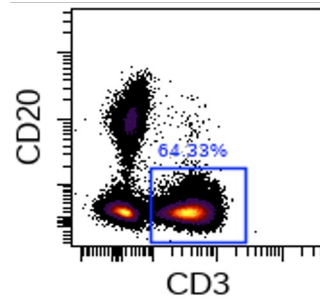
Computational Cytometry Data Analysis Workflow for Cytobank



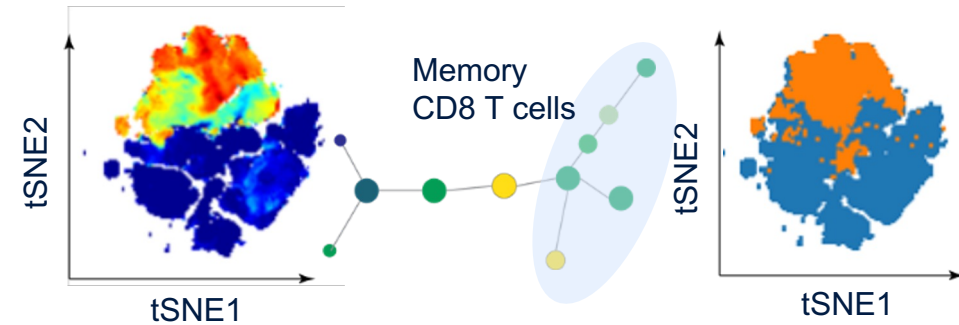
Data Acquisition



QC and clean up



Pre-gate (optional)



viSNE,
tSNE-CUDA,
opt-SNE,
UMAP

SPADE,
FlowSOM,
CITRUS

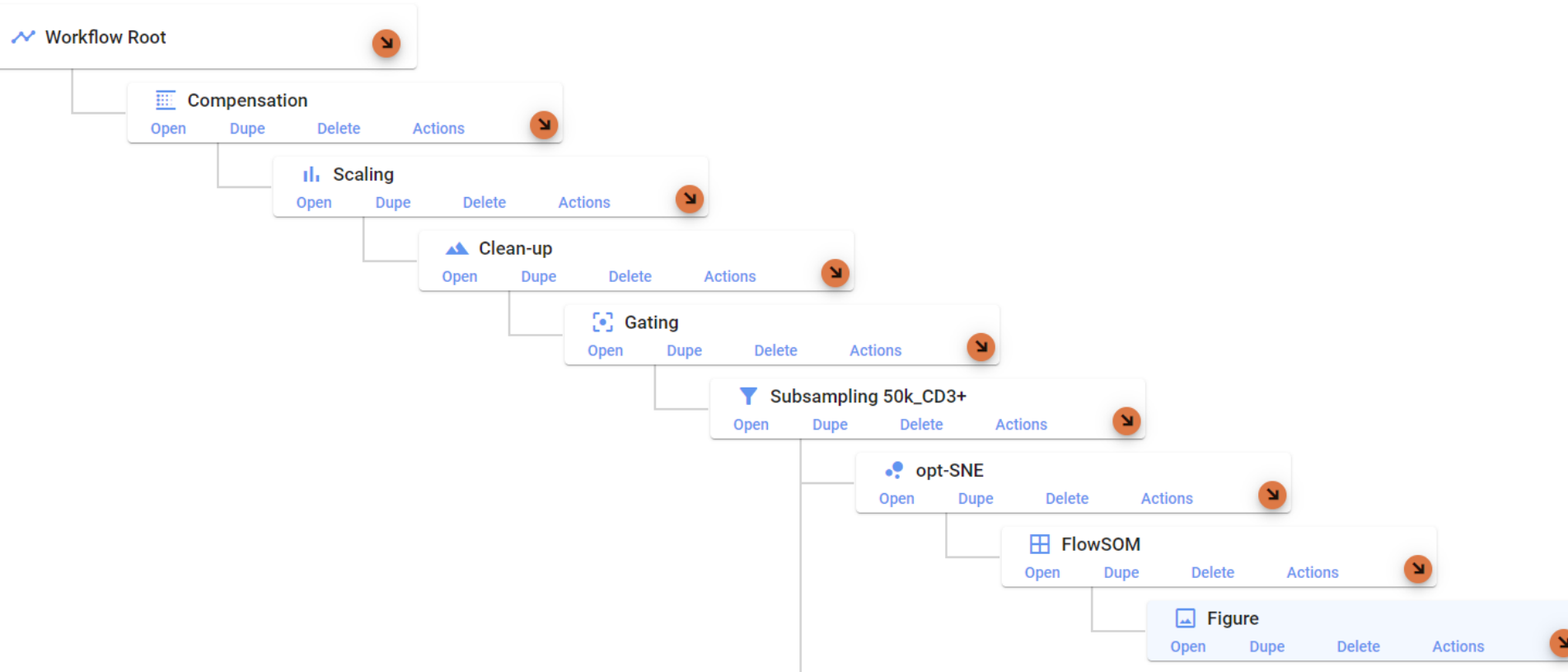
viSNE Overlay



Preprocessing

supervised

unsupervised



Dimensionality reduction

Clustering



<https://www.chugcytometry.com>

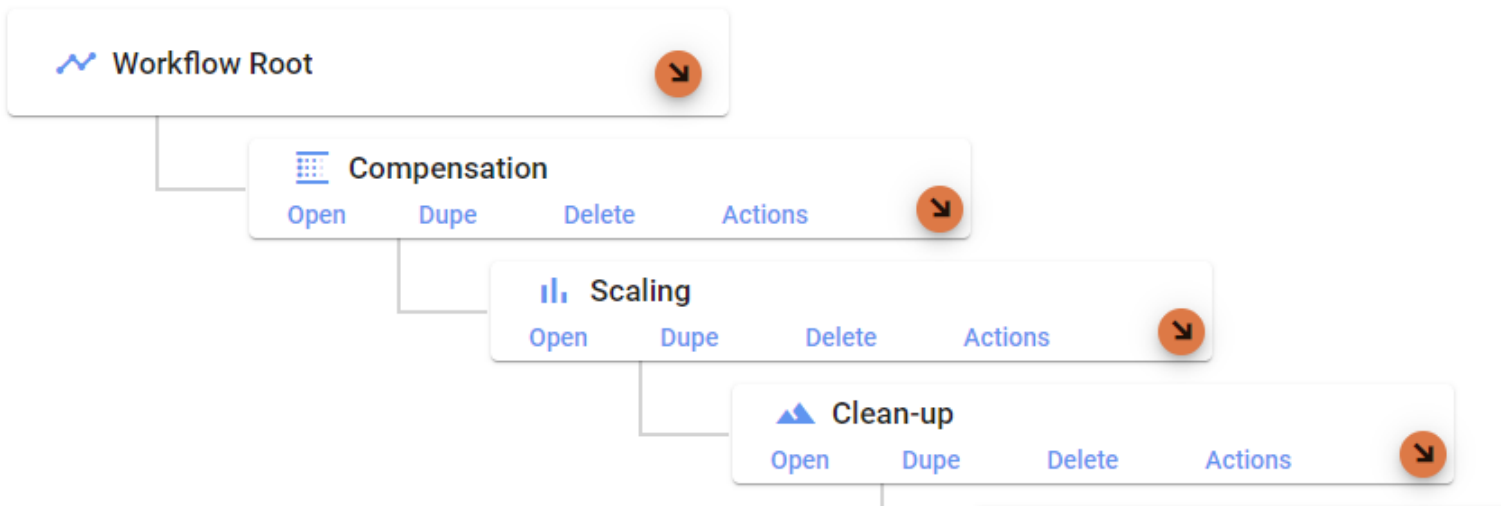


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Preprocessing



Compensation

Select a Compensation Group

Comp Group 1 ▾

[+ Create New Comp](#)

[🔍 Find File](#)

[⋮ More](#)

Files in Group (10)

File Name

PBMC Unstim_exp3_3.fcs
PBMC Unstim EXP 2.fcs
PBMC Unstim.fcs
PBMC Unstim_exp3_2.fcs
PBMC Stim_exp3_1.fcs
PBMC Stim.fcs
PBMC Unstim_exp3_1.fcs
PBMC Stim_exp3_3.fcs
PBMC Stim EXP 2.fcs
PBMC Stim_exp3_2.fcs

Comp Matrix

⋮ Actions

Features	Copy			BV805-A	BV421-A	LIVE DEAD Aqua-A	StarBright570-A	BV605-A	StarBright670-A	BV711-A	BV785-A	BB515-A	NovaBlue 610-A	NovaBlue 660-A	PE-A	PE-CF594-A	PE-Cy7-A	APC-A	APC-R700-A	APC-Fire 750-A	AF-A
				0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
BUV395-A	Paste			0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
BUV496-A	Highlight Feature Pair			0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
BUV737-A				0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
BUV805-A				100	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
BV421-A	Add Files to Group			0	100	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
LIVE DEAD Aqua-A	Remove Files from Group			0	0	100	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
StarBright570-A				0	0	0	100	0	0	0	0	0	0	0	0	0	0	0	0	0	
BV605-A				0	0	0	0	100	0	0	0	0	0	0	0	0	0	0	0	0	
StarBright670-A	Copy Features Only			0	0	0	0	0	100	0	0	0	0	0	0	0	0	0	0	0	
BV711-A	Revert to Original			0	0	0	0	0	0	100	0	0	0	0	0	0	0	0	0	0	
BV785-A				0	0	0	0	0	0	100	0	0	0	0	0	0	0	0	0	0	
BB515-A	Delete Group			0	0	0	0	0	0	0	0	100	0	0	0	0	0	0	0	0	
NovaBlue 610-A				0	0	0	0	0	0	0	0	0	100	0	0	0	0	0	0	0	
NovaBlue 660-A	0	0	0	0	0	0	0	0	0	0	0	0	0	100	0	0	0	0	0	0	
PE-A	0	0	0	0	0	0	0	0	0	0	0	0	0	0	100	0	0	0	0	0	
PE-CF594-A	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	100	0	0	0	0	
PE-Cy7-A	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	100	0	0	0	
APC-A	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	-2.7	0	100	0	0	
APC-R700-A	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	100	0	0	
APC-Fire 750-A	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	100	0	
AF-A	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	100	



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Scaling - aka transformation

Filter Features

Feature Name	Scaling Type	Cofactor	Min	Max
Time	None (linear)		0	6023025
SSC-H	None (linear)		0	4194304
SSC-A	None (linear)		0	4194304
FSC-H	None (linear)		0	4194304
FSC-A	None (linear)		0	4194304
SSC-B-H	None (linear)		0	4194304
SSC-B-A	None (linear)		0	4194304
BUV395-A CD11b	Arcsinh	600	-13000	4194304
BUV496-A CD16	Arcsinh	2000	-13000	4194304
BUV737-A IgD	Arcsinh	6000	-13000	4194304
BUV805-A CD14	Arcsinh	6000	-13000	4194304
BV421-A CD66b	Arcsinh	6000	-13000	4194304
LIVE DEAD Aqua-A Live/Dead	Arcsinh	3000	-20000	4194304
StarBright570-A CD27	Arcsinh	6000	-13000	4194304
BV605-A IgM	Arcsinh	6000	-13000	4194304
StarBright670-A CD19	Arcsinh	1000	-20000	4194304
BV711-A CD20	Arcsinh	6000	-13000	4194304
BV785-A CD8a	Arcsinh	1000	-13000	4194304
BB515-A CD35	Arcsinh	6000	-13000	4194304
NovaBlue 610-A CD45	Arcsinh	400	-13000	4194304
NovaBlue 660-A CD3	Arcsinh	1500	-13000	4194304
PE-A TCRab	Arcsinh	1000	-13000	4194304

Select File

PBMC Unstim.fcs

Select Plot Type

Scatterplot (Density)

Select Filter

Y Min

-13000

Y Max

4194304

X Min

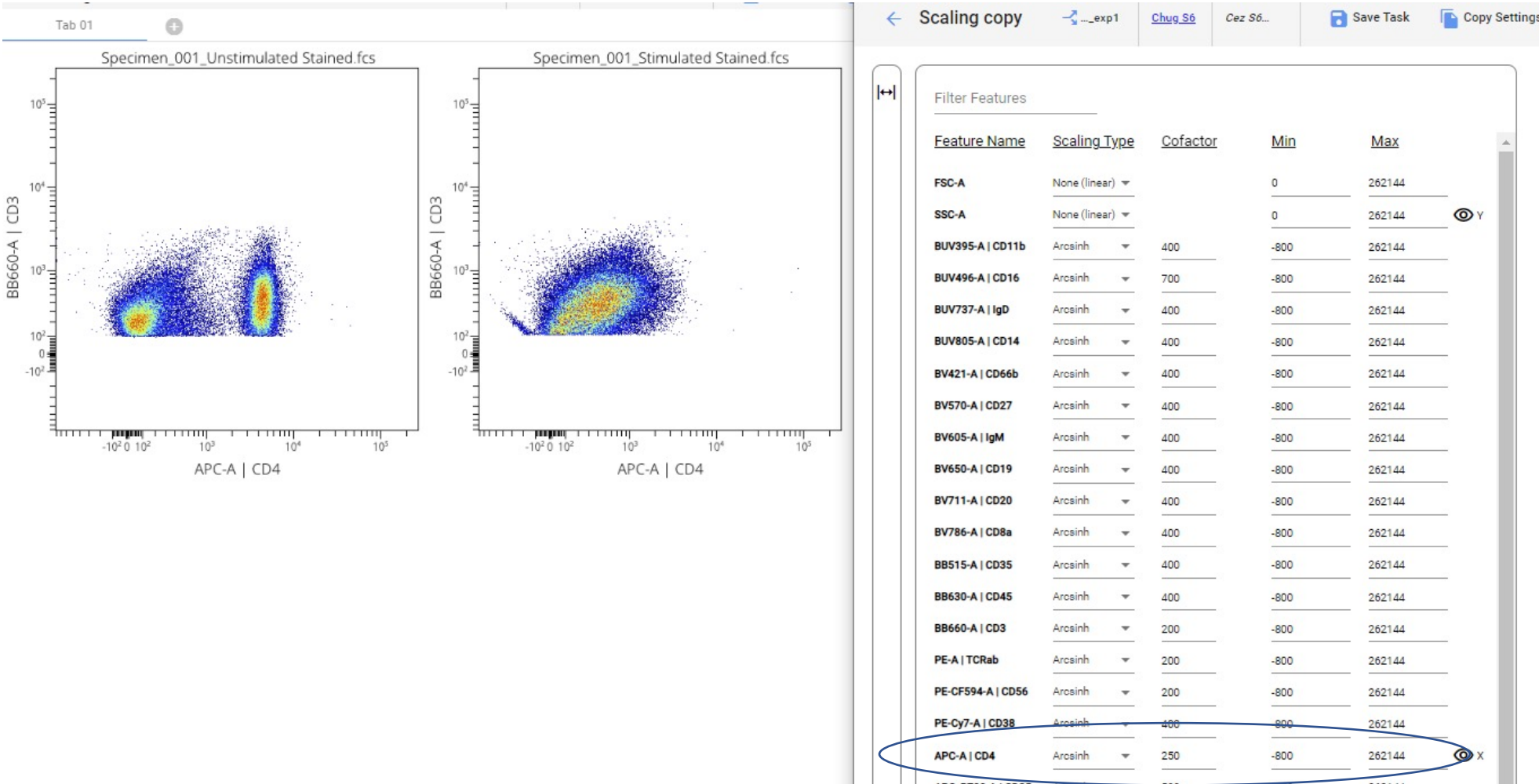
-13000

X Max

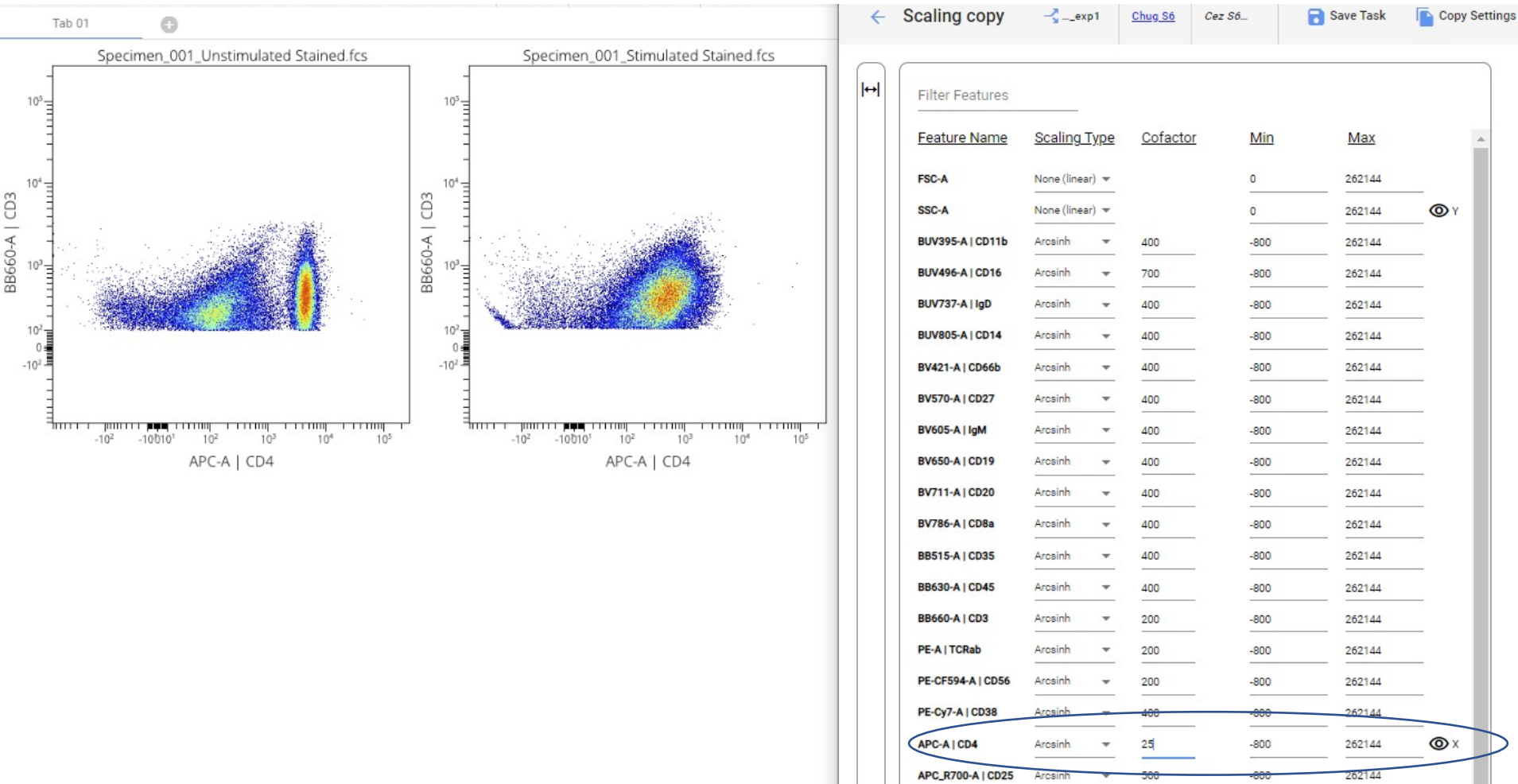
4194304



Scaling - aka transformation

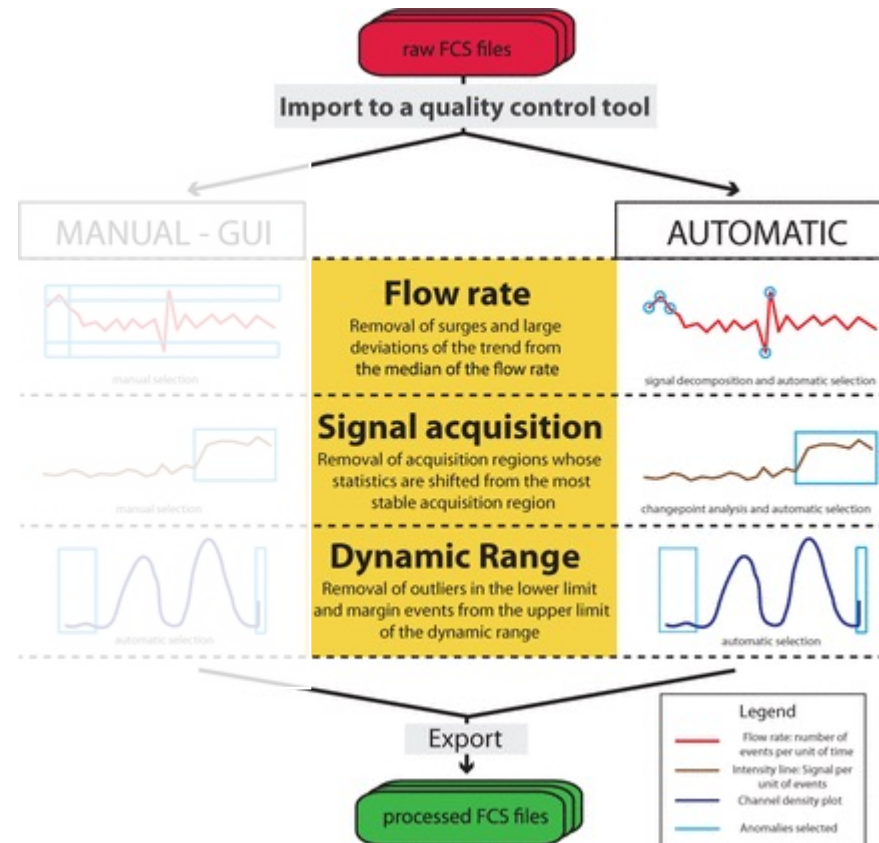


Scaling - aka transformation



Data cleanup

Automated methods that can identify and remove artefacts generated during data acquisition (clogs, drastic flow rate changes, out of range signal)



Data cleanup

3 cleanup algorithms were run (all default settings) on our data set (20 files from Aurora) for comparison purpose.

Data Cleaning

flowAI



flowCut



PeacoQC



% of rejected events

FlowAI (12min) flowCut (36min) PeacoQC (11min)

PBMC Stim	7.39	0	3.30
PBMC Unstim	14.6	2.48	13.26
PBMC Stim exp2	10.27	0.05	4.60
PBMC Unstim exp2	9	0	5.64
PBMC Stim_exp3_1	6.28	0	3.28
PBMC Stim_exp3_2	6.47	3.8	5.27
PBMC Stim_exp3_3	4.89	0.05	4.48
PBMC Unstim_exp3_1	8.59	0	7.56
PBMC Unstim_exp3_2	8.16	0	8.30
PBMC Unstim_exp3_3	8.38	0	6.64

<https://doi.org/10.1093/bioinformatics/btw191>

<https://doi.org/10.1002/cyto.a.24670>

<https://doi.org/10.1002/cyto.a.24501>

Gating = cleanup too



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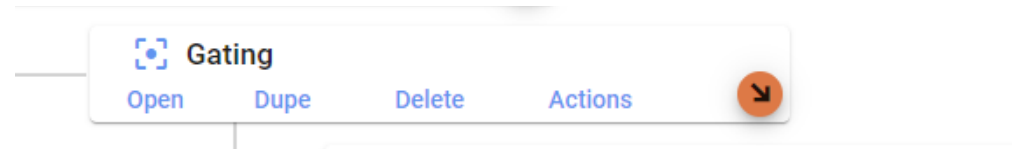


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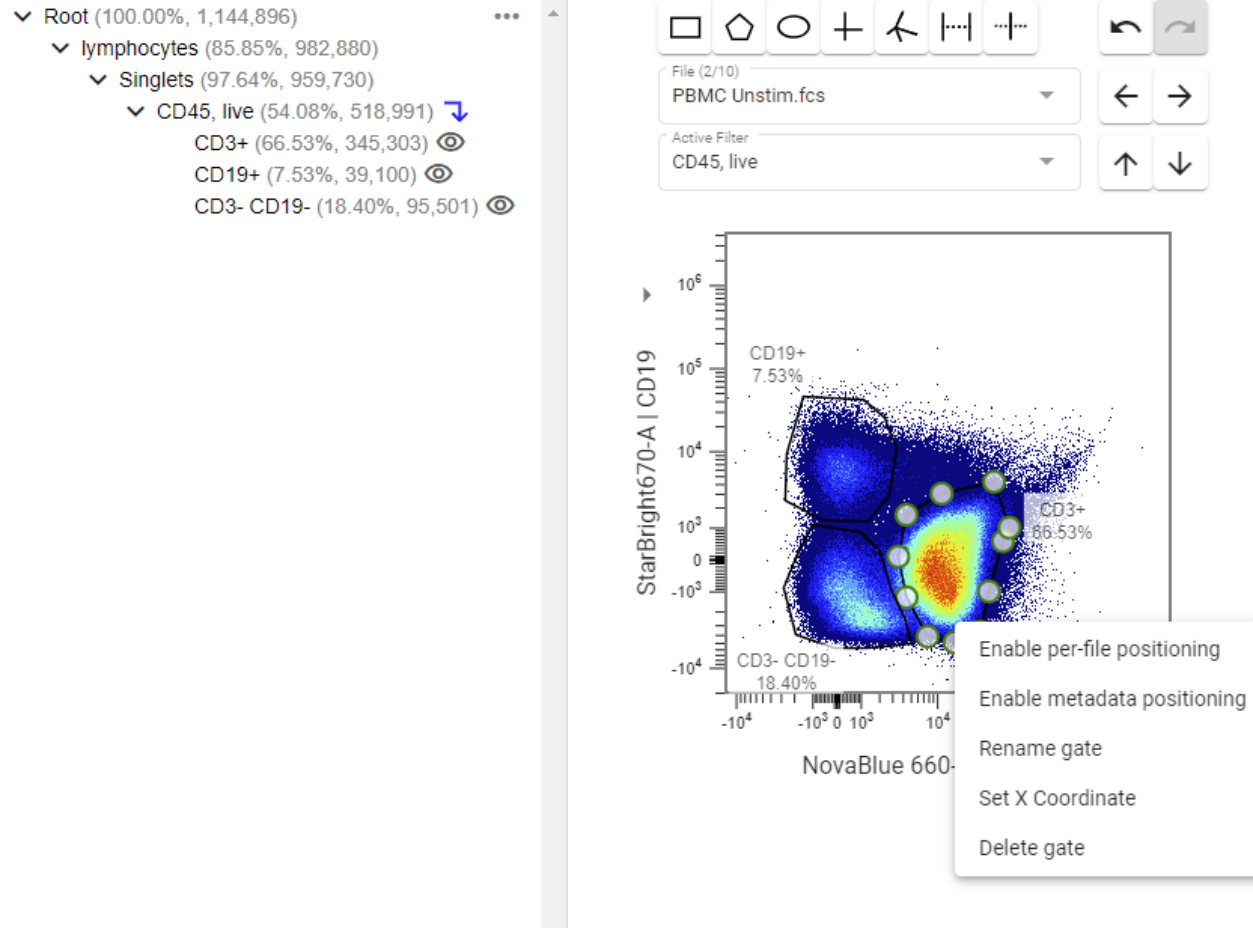


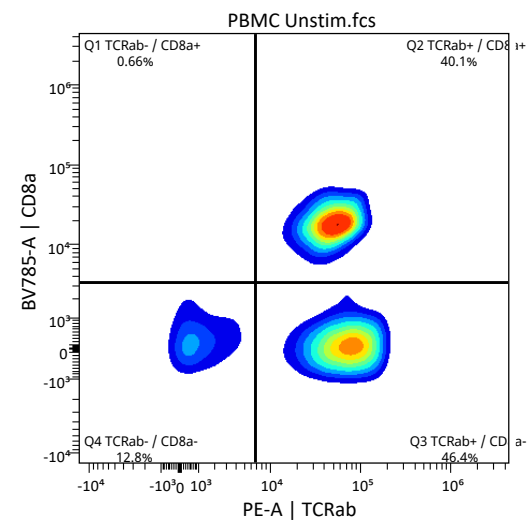
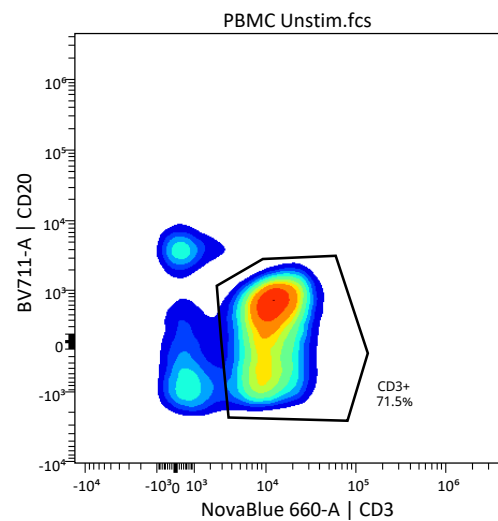
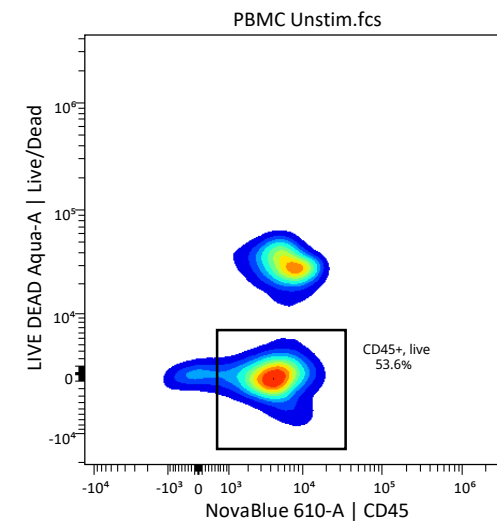
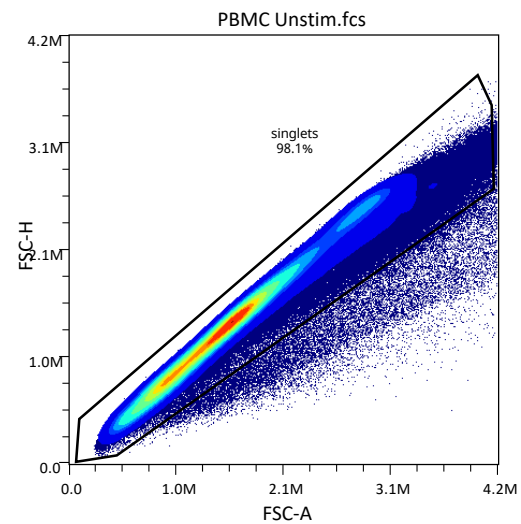
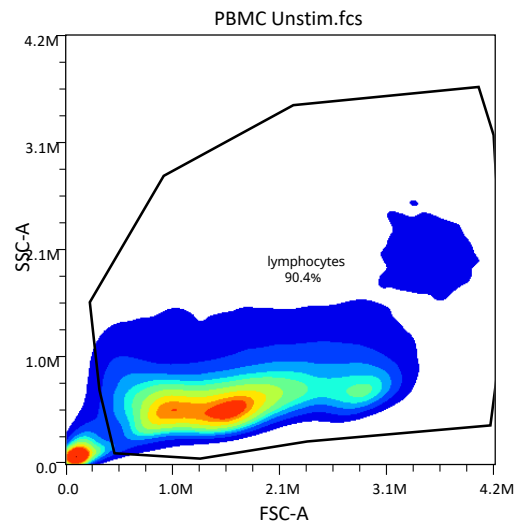
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supervised



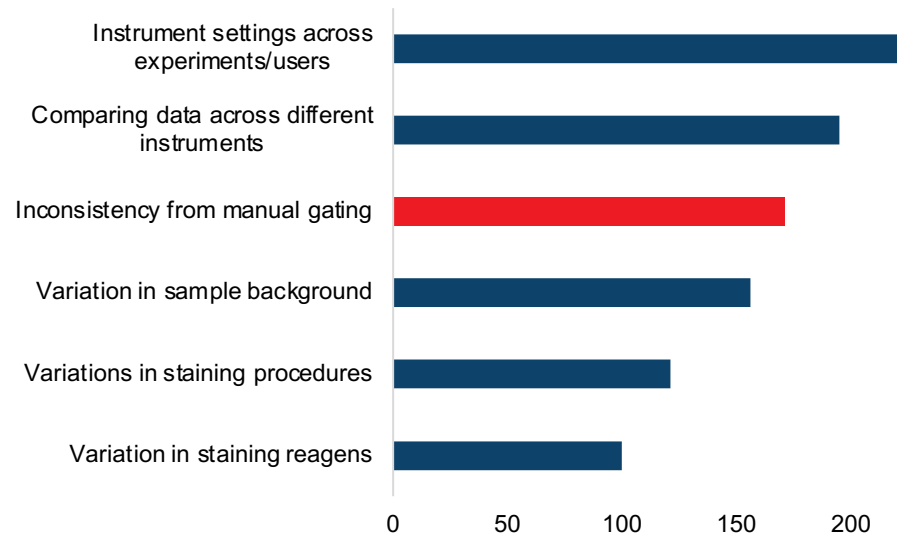
Supervised, gating





Manual gating is a concern!

What aspects of reproducibility are cytometrists most concerned about? [multi-select]



Source:

- 2019 Flow and Mass Cytometry User Survey, >300 participants



Cytobank v10 has recently added a new Automatic gating feature

- Automate the gating process to reduce intra- and inter-operator variability.
- Eliminate time spent on repetitive, manual tasks, to focus result interpretation and the generation of novel insights.



Cytobank Automatic gating is trained on your panel and gating strategy



Share your Cytobank Automatic gating model easily with other users



Complete gating in less than one minute per sample*

Slide courtesy of Cytobank,
Beckman Coulter

* For files with 300 000 events, 19 parameters and 42 populations.
Performance dependent on gating strategy and file size.



Subsampling/Downsampling

Select Filter
CD3+ ▼

Target Count *
50000

Counting Method
☒ Per File ☐ Total

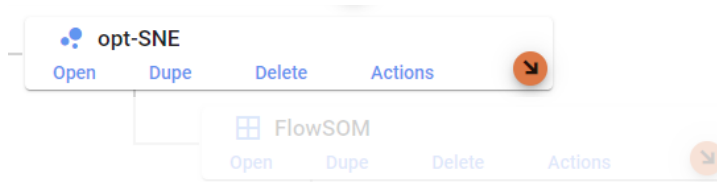
Sampling Method
☐ Equal ☒ Max Equal

☐ Group Files by Metadata

File Name (10)	Available	Available in Filter	Sampling Count
PBMC Stim.fcs	618,808	63,910	50,000
PBMC Unstim.fcs	1,144,896	245,692	50,000
PBMC Stim EXP 2.fcs	1,036,896	93,528	50,000
PBMC Unstim EXP 2.fcs	2,060,040	453,656	50,000
PBMC Stim_exp3_3.fcs	806,640	78,542	50,000
PBMC Stim_exp3_1.fcs	776,728	76,020	50,000
PBMC Stim_exp3_2.fcs	987,632	118,508	50,000
PBMC Unstim_exp3_3.fcs	1,193,664	242,162	50,000
PBMC Unstim_exp3_1.fcs	1,286,360	223,917	50,000
PBMC Unstim_exp3_2.fcs	1,316,456	236,166	50,000
Total	11,228,120	1,832,101	500,000




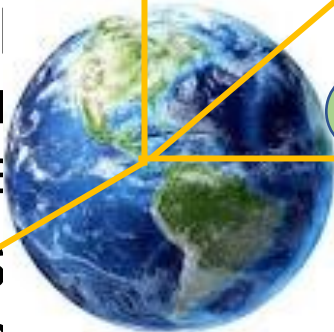

unsupervised



Dimensionality reduction



Common approaches to dimensionality reduction-

- viSNE/tSNE- non-linear dimensionality reduction algorithm developed based on Stochastic Neighbor Embedding-similar objects are modeled by nearby points and dissimilar objects are modeled by distant points with high probability
- tSNE-CUDA- GPU accelerated t-distributed stochastic neighbor embedding  computational time
- UMA |  Manifold Approach for Dimensionality Reduction of the underlying dataset using a topological framework
- opS  produce superior dimensionality reduction results than the original t-SNE algorithms



Fast Dimensionality Reduction: How To Choose?

	viSNE	Opt-SNE	tSNE-CUDA	UMAP
Set the Seed	Yes	Yes	-	-
Max # total events	1.3M	3M	10M	10M
Global structure	-	-	-	Yes
Speed	Slower	Medium	Fast	Fast
Low events/sample	More sparse map			More condense map/ islands
Visualization	Bigger, less compact islands with points more spread			More dense islands with overlapping dots

- ❖ UMAP can generate islands that are distance from each other and dense, therefore, it might be more clear than a tSNE map for samples with few events
- ❖ tSNE-CUDA islands are bigger, less compact, the points are more spread out and don't overlap each other as much as UMAP. It will be easier to discriminate subpopulations within the tSNE-CUDA islands than the dense UMAP islands
- ❖ Distance between islands in UMAP is meaningful while it's not for the tSNEs maps
- ❖ tSNE-CUDA might be a fast option to explore large datasets working well with default settings BUT if you want to be able to include the seed for your run in your paper, then you need to use opt-SNE



The good and the bad...

Advantages of Dimensionality Reduction

- Data compression and better visualization
- Reduces time of analysis
- Allows automation, identification of unique populations, biomarker discovery and predictive modelling to correlate with clinical outcome.

Disadvantages of Dimensionality Reduction

- Potential data loss
- Requires either a good code writer or potentially expensive software



Multiparameter data analysis

Tools/algorithms

Dimensionality Reduction

t-SNE



opt-SNE



t-SNE-CUDA



FIt-SNE



UMAP



PCA



EmbedSOM



TriMap



Isomap



<https://www.chugcytometry.com>



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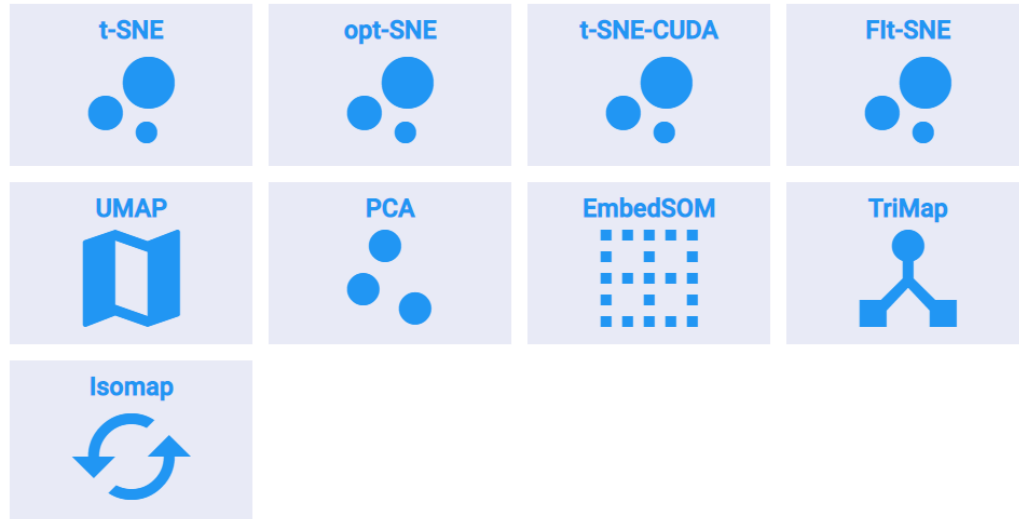


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Multiparameter data analysis

Tools/algorithms

Dimensionality Reduction



Opt-SNE: <https://doi.org/10.1101/451690>

UMAP: <https://arxiv.org/abs/1802.03426>

EmbedSOM: <https://doi.org/10.12688%2Ff1000research.21642.2>

TriMap: <https://arxiv.org/abs/1910.00204>

Isomap: <https://doi.org/10.1126/science.290.5500.2319>



<https://www.chugcytometry.com>

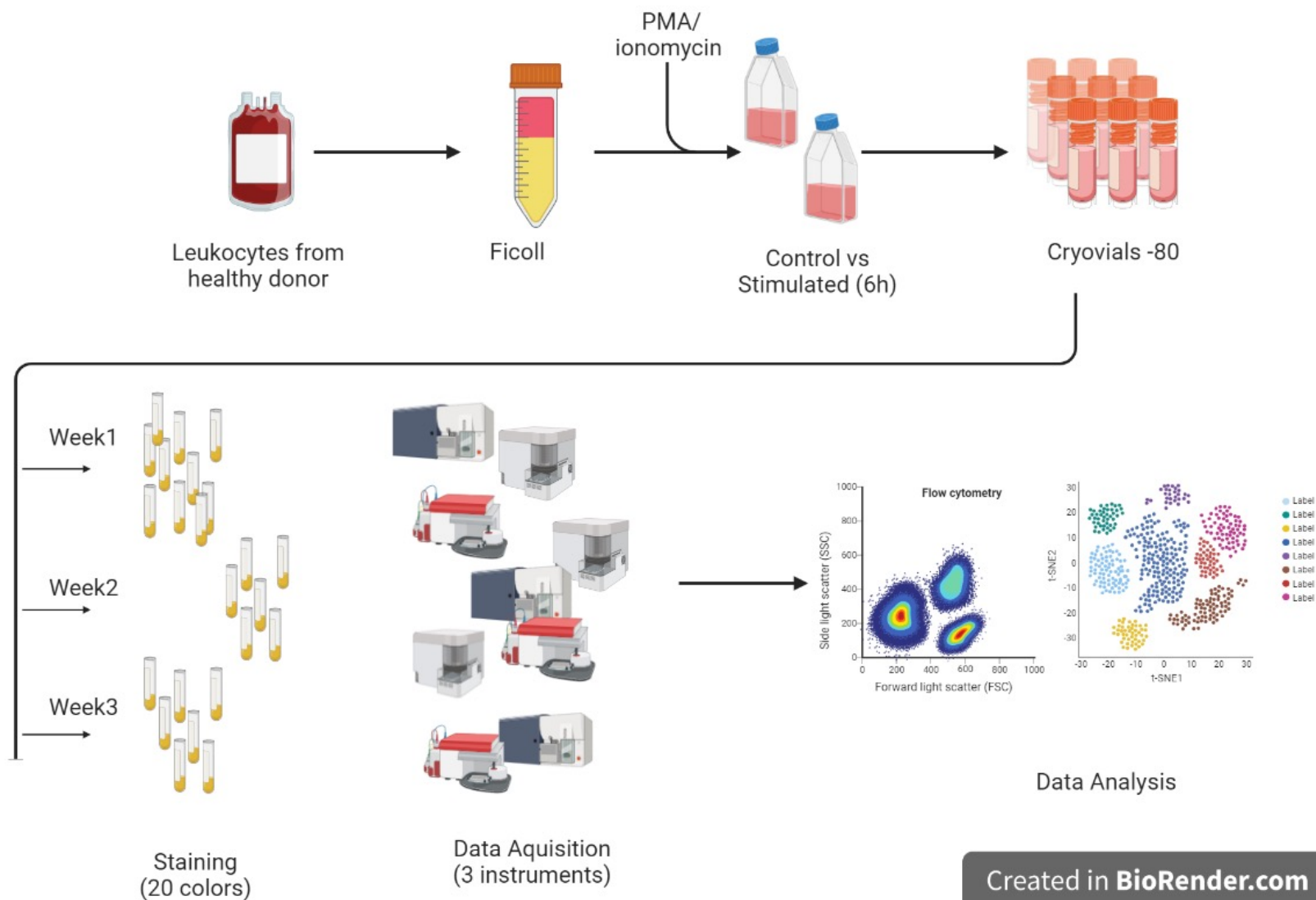


@chugcytometry



ChUG Cytometry

Experiment outline



Created in **BioRender.com** **bio**



<https://www.chugcytometry.com>

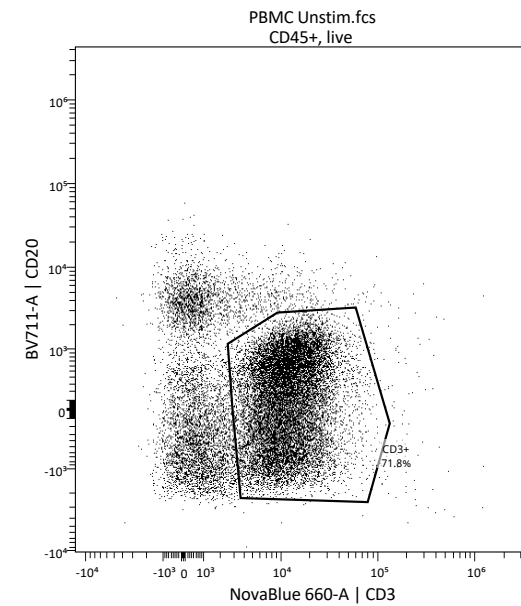
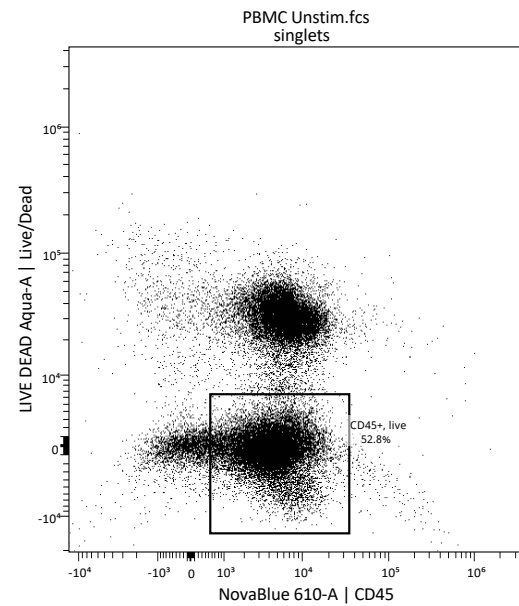
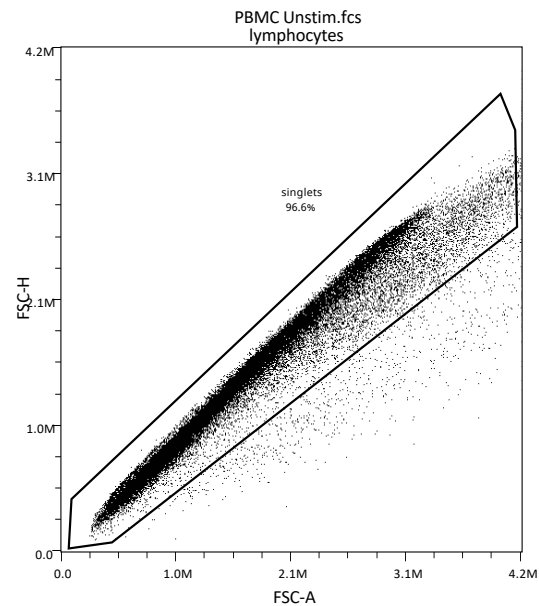
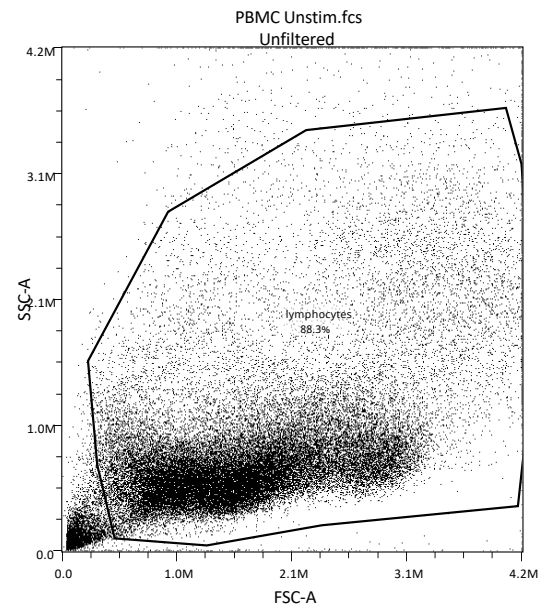


@chugcytometry



ChUG Cytometry

Supervised, gating

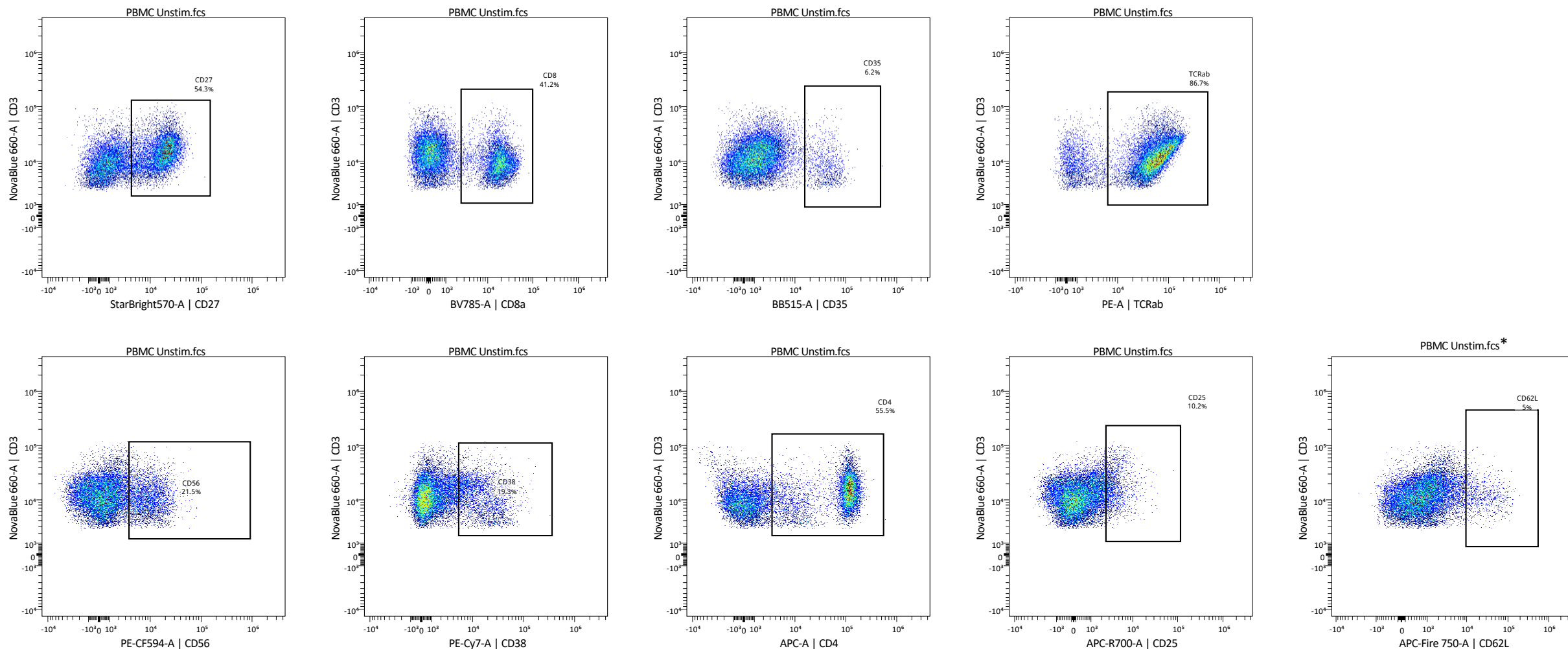


BUV395	CD11b	BB515	CD35
BUV496	CD16	NB610	CD45
Live/Dead Aqua	Viability	NB660	CD3
BUV737	IgD	PE	TCRa/b
BUV805	CD14	PE-CF594	CD56
BV421	CD66b	PE-Cy7	CD38
SBV570	CD27	APC	CD4
BV605	IgM	APC-R700	CD25
SBV670	CD19	APC-Fire750	CD62L
BV711	CD20		
BV785	CD8a		

SBV570	CD27
BV785	CD8a
BB515	CD35
PE	TCRa/b
PE-CF594	CD56
PE-Cy7	CD38
APC	CD4
APC-R700	CD25
APC-Fire750	CD62L



Supervised, gating



*CD62L expression pattern seems to be incorrect, new antibody the same pattern but titration on fresh PBMC ok, Possible that freeze/thaw cycle affect CD62L expression
[https://doi.org/10.1016/s0022-1759\(03\)00202-3](https://doi.org/10.1016/s0022-1759(03)00202-3)



<https://www.chugcytometry.com>



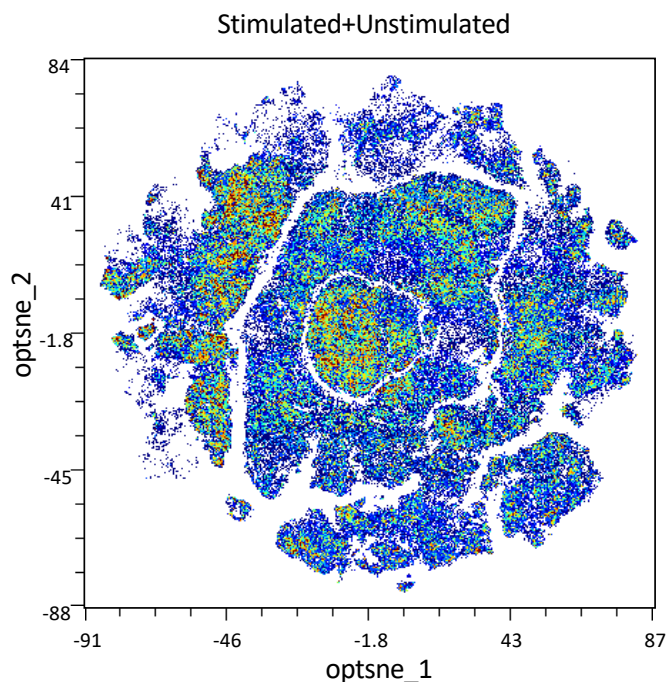
@chugcytometry



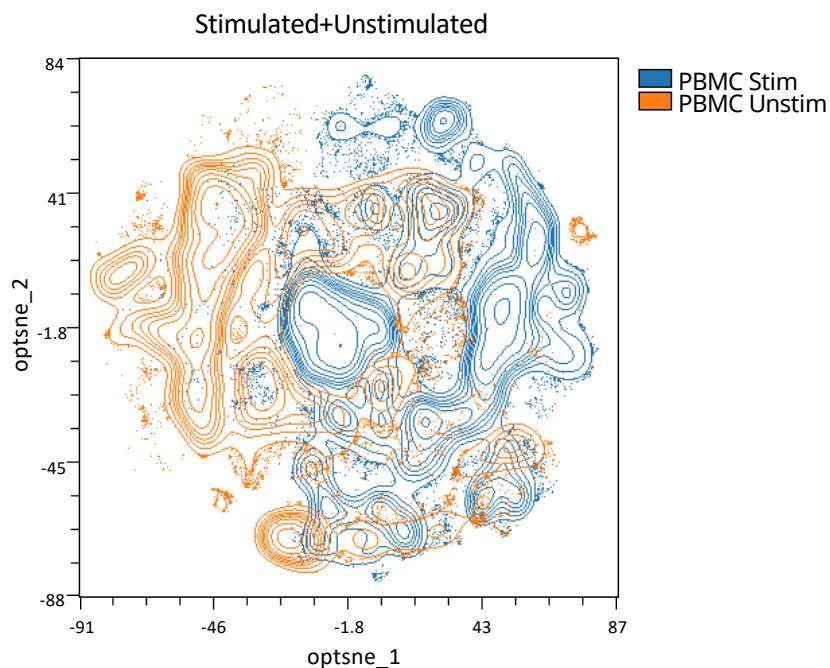
ChUG Cytometry

opt-SNE

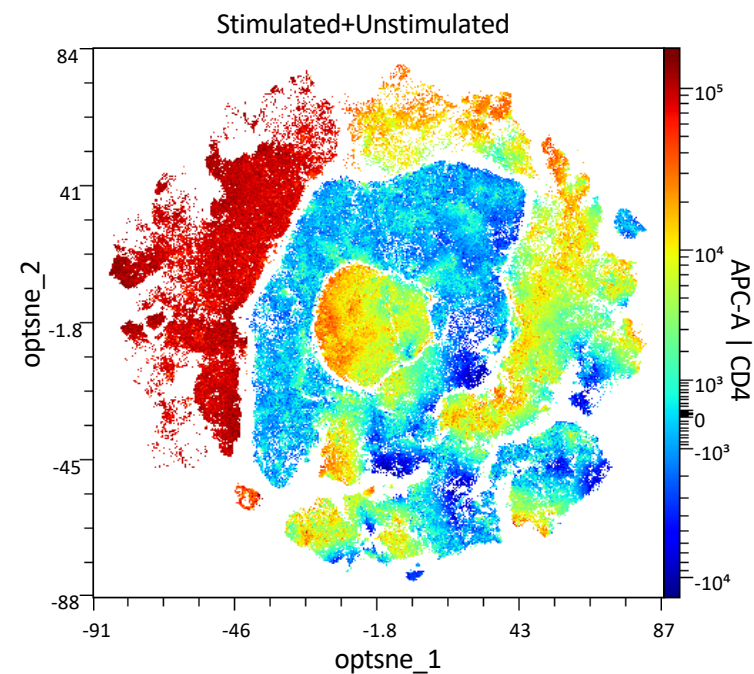
Modified version of t-SNE that enables high-quality embeddings in the optimal amount of compute time without having to tune algorithm parameters.



tSNE with Scatterplot (density)



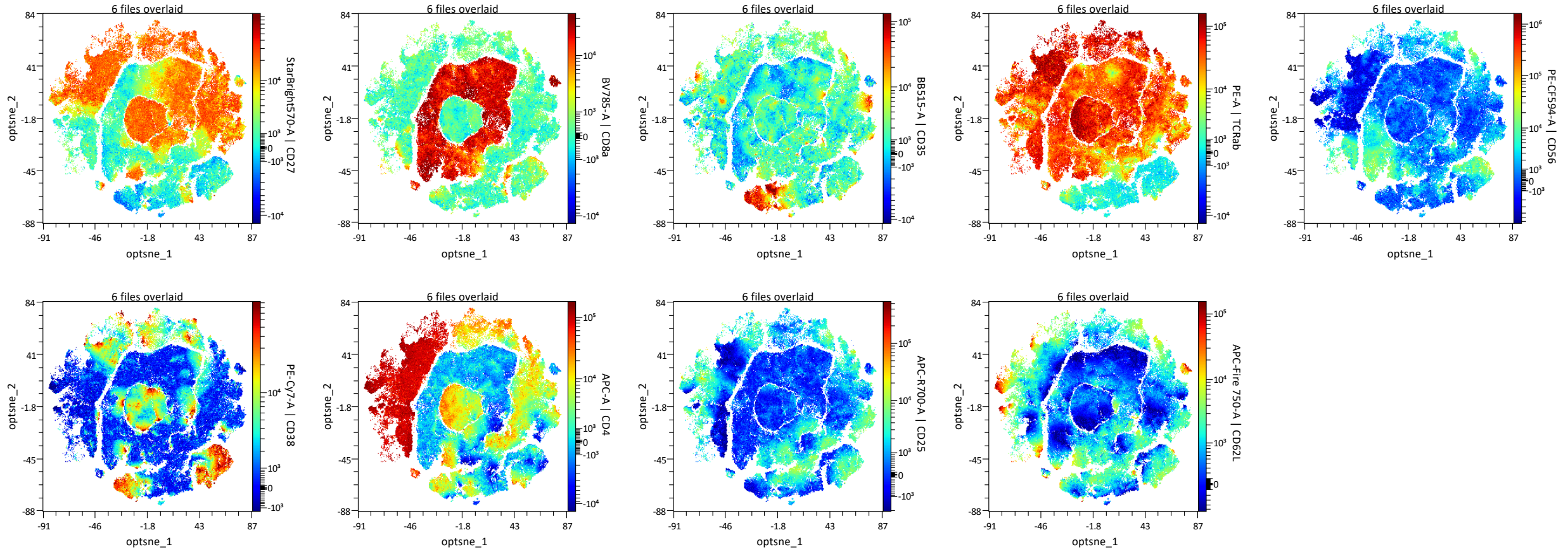
tSNE with Contour Plot



Scatterplot (colored-continuous)



Visualization of data in higher number of dimension

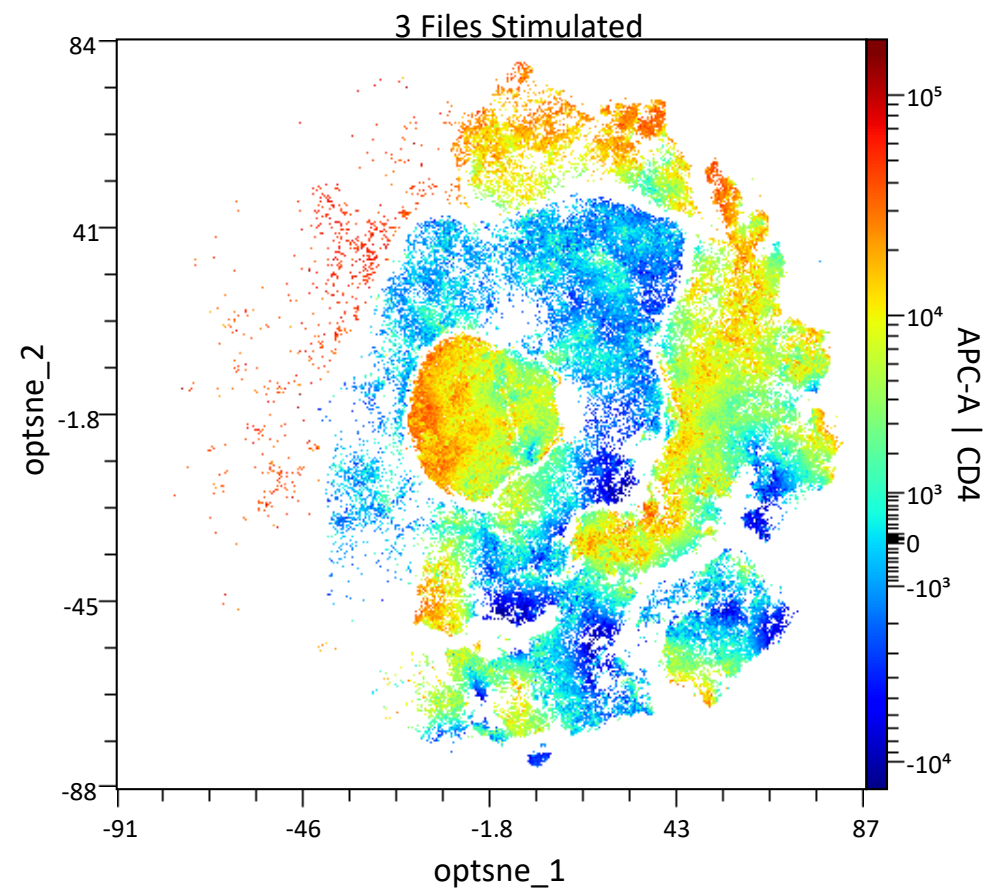
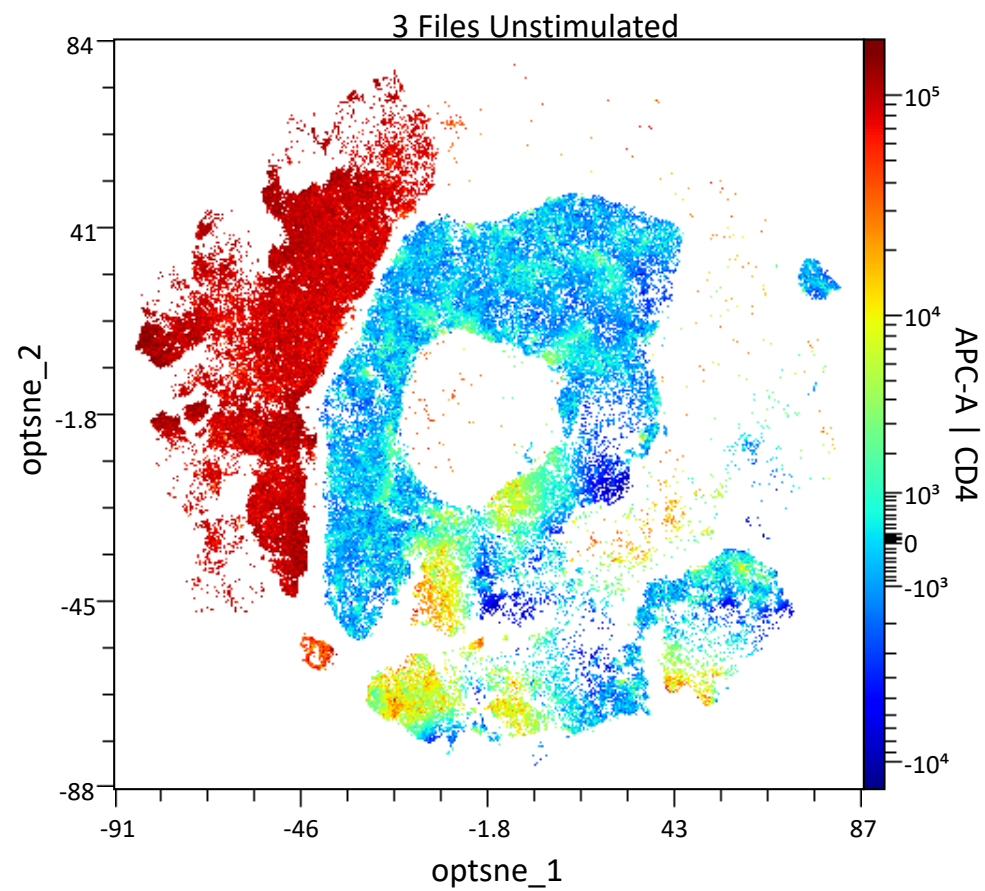


The diagram illustrates the experimental workflow for measuring cytokine production. It consists of three main steps:

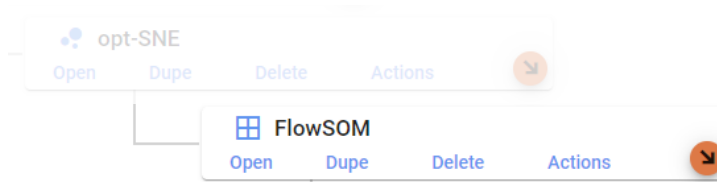
- Stimulation:** Cells are stimulated with PMA/ionomycin.
- Control vs Stimulated (6h):** Cells are grown in Control vs Stimulated (6h) conditions.
- Harvest and Analysis:** Cells are harvested and analyzed by flow cytometry.



Yes, we can!



unsupervised



Clustering



Clustering

- Dividing the population or data points into groups where data points in the same groups are more similar to other data points and dissimilar to the data points in other groups.



Clustering

- *SPADE*=Spanning-tree Progression Analysis of Density-normalized Events
 - clusters phenotypically-similar cells into a hierarchy that allows high-throughput, multidimensional analysis of heterogeneous samples.
- *FlowSOM*= analysis and quality of clustering with Self-Organizing Maps (SOMs)
 - Quicker analysis, clusters cells based on chosen markers

CITRUS=algorithm developed to automatically find stratifying signatures from within a data set to explain differences between multiple groups of samples (responders vs non-responders). Very useful for human studies with a high number of experimental samples.



Some references-

- Toward deterministic and semi-automated SPADE analysis-[Cytometry A. 2017 Mar; 91\(3\): 281–289.](#)
- viSNE enables visualization of high dimensional single-cell data and reveals phenotypic heterogeneity of leukemia-[Nat Biotechnol. 2013 Jun; 31\(6\): 545–552.](#)
- Analyzing high-dimensional cytometry data using FlowSOM-[Nature Protocols](#) **volume 16**, pages3775–3801 (2021)
- A Beginner's Guide to Analyzing and Visualizing Mass Cytometry Data 2018 Jan 1;200(1):3-22. doi: 10.4049/jimmunol.1701494



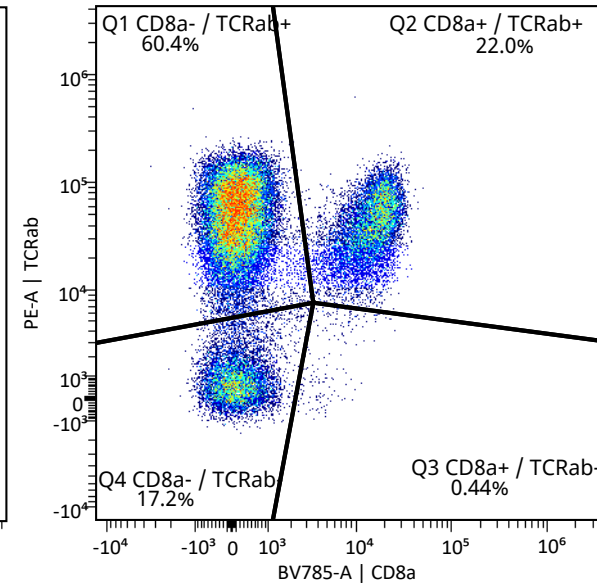
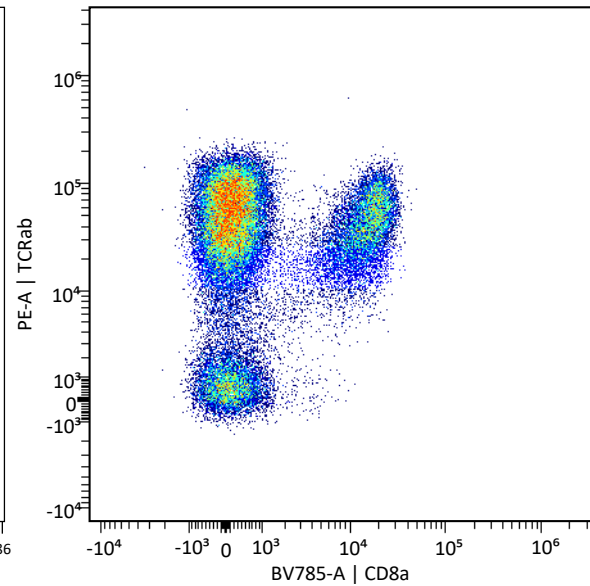
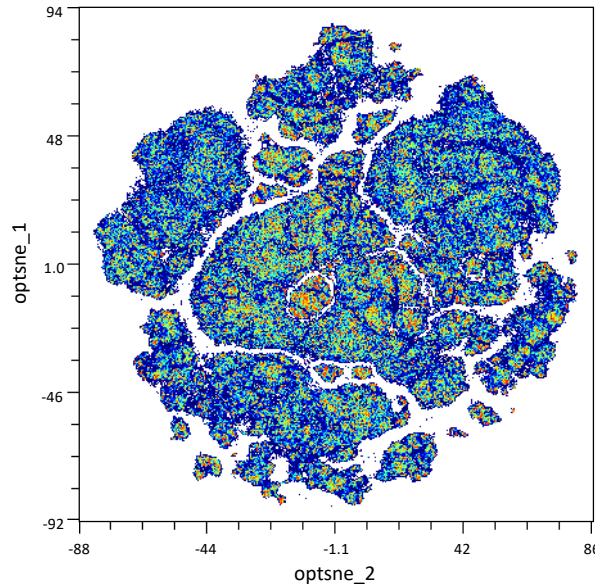
Advantages vs limitations for clustering

Individual cells are not displayed, only clusters are.

No automatic methods to determine the number of clusters and meta-clusters based on the individual panel. It is user-defined and often needs to be optimized by a couple of runs



Clustering

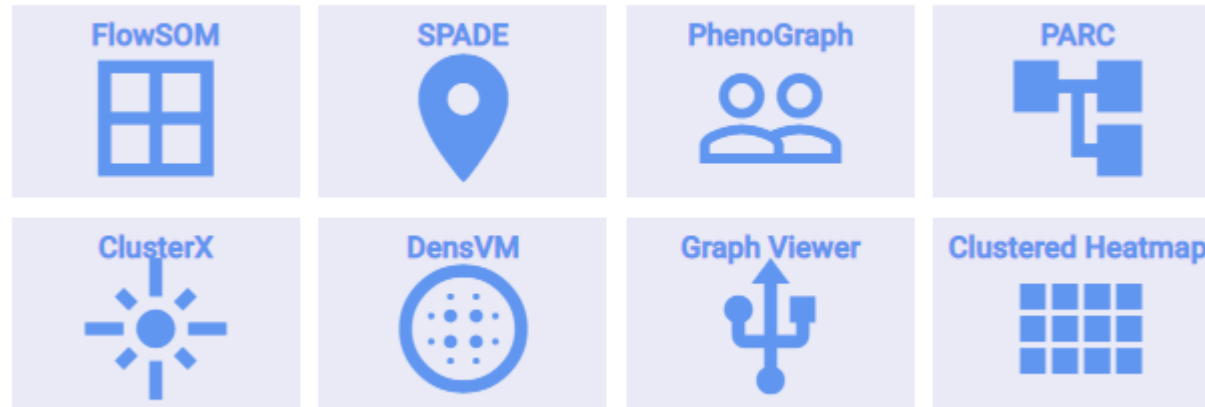


Clustering is like gating
but for multidimensional
data set



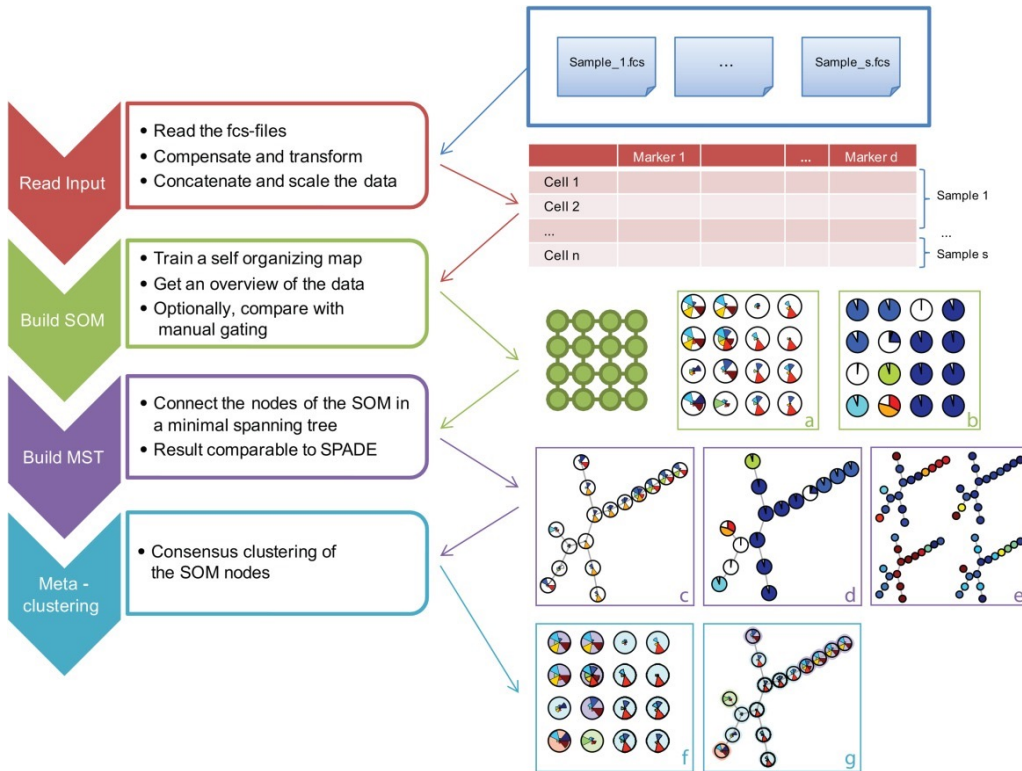
Clustering

Clustering



Clustering with FlowSOM

Uses Self-Organizing Map
Allows to set seed
Reproducible
Fast
Works with large number of events and samples
Outputs clusters (~overclustering) and metaclusters

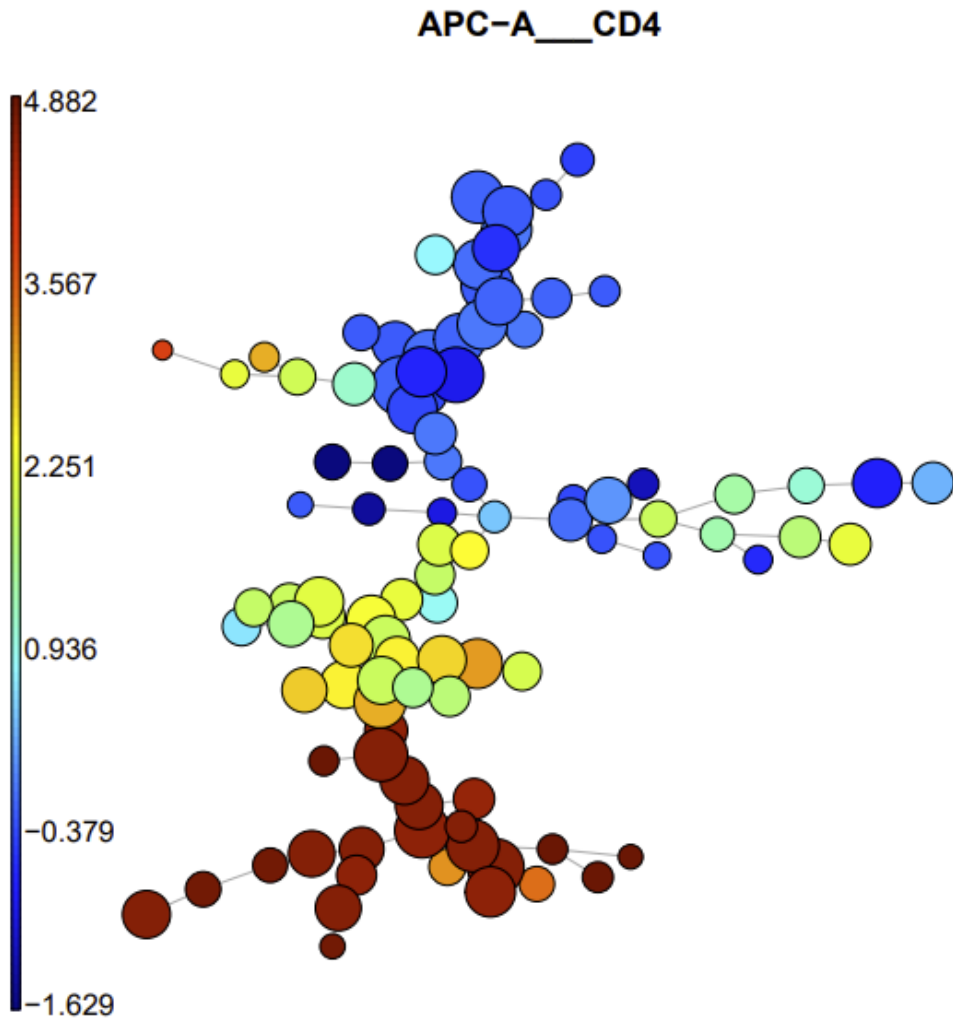


DOI: 10.1002/cyto.a.22625



Clusters and metaclusters

Minimal spanning tree

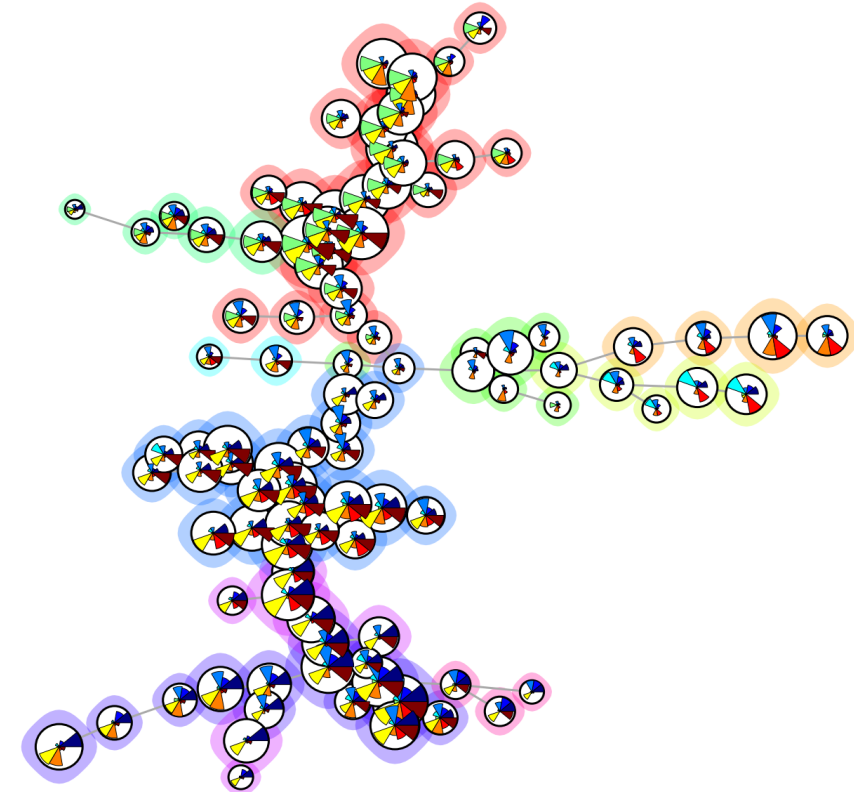


Metaclusters from consensus method (k=10) visualized on tree

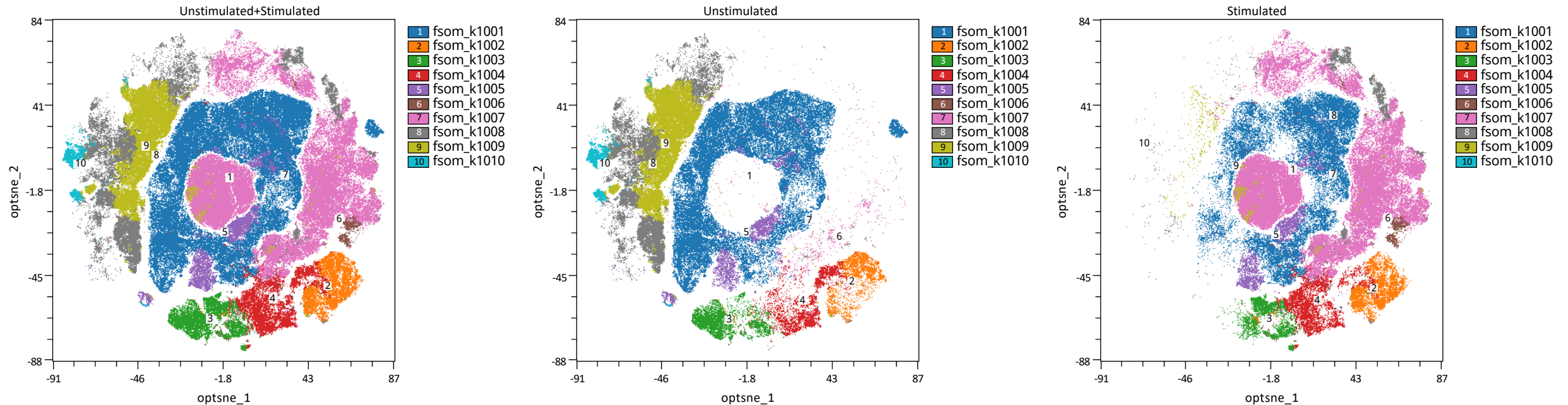
APC-R700-A__CD25
BB515-A__CD35
BV785-A__CD8a
PE-A__TCRab
PE-CF594-A__CD56
APC-Fire 750-A__CD62
APC-A__CD4
StarBright570-A__CD27
PE-Cy7-A__CD38

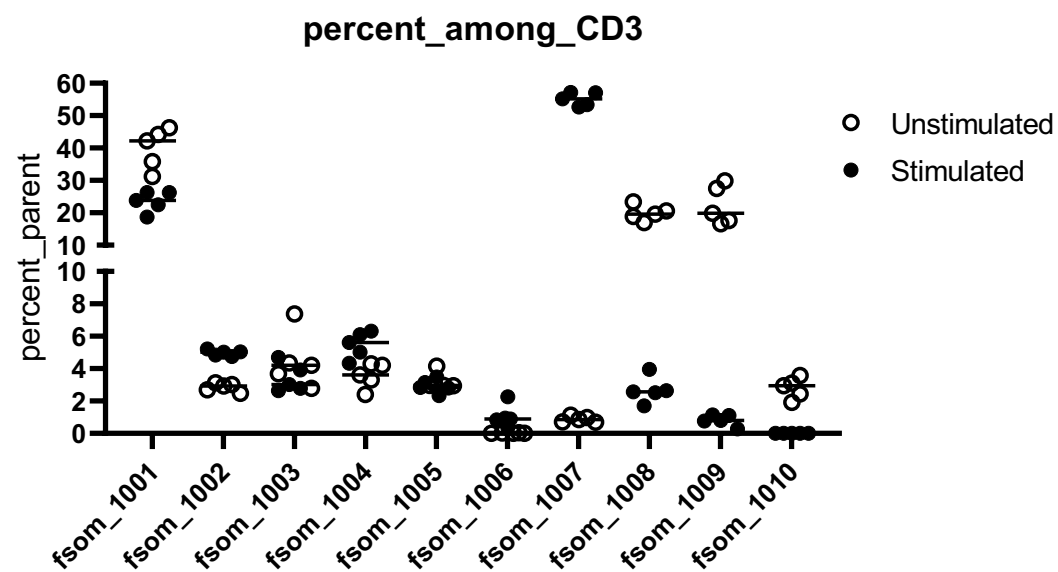
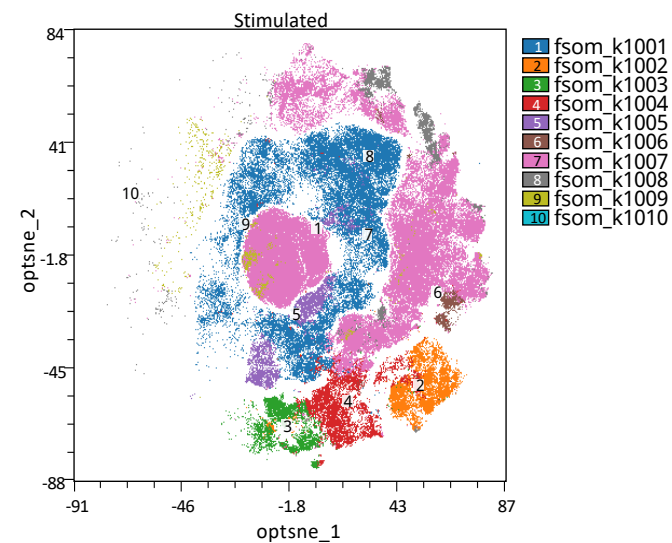
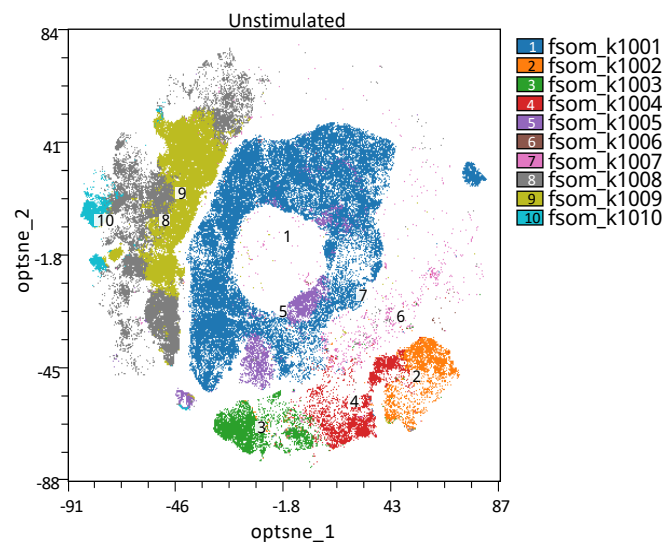
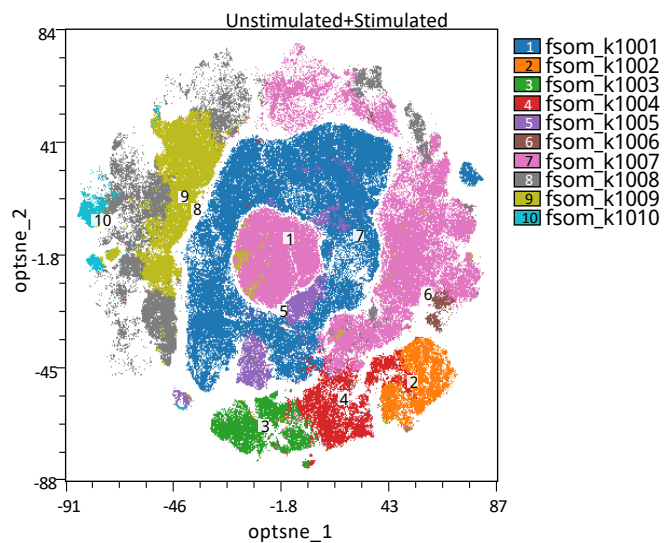
Metaclusters

- 1
- 2
- 3
- 4
- 5
- 6
- 7
- 8
- 9
- 10

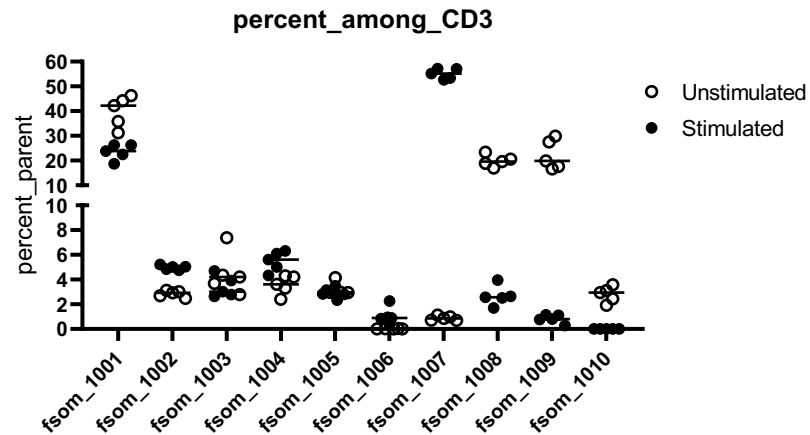
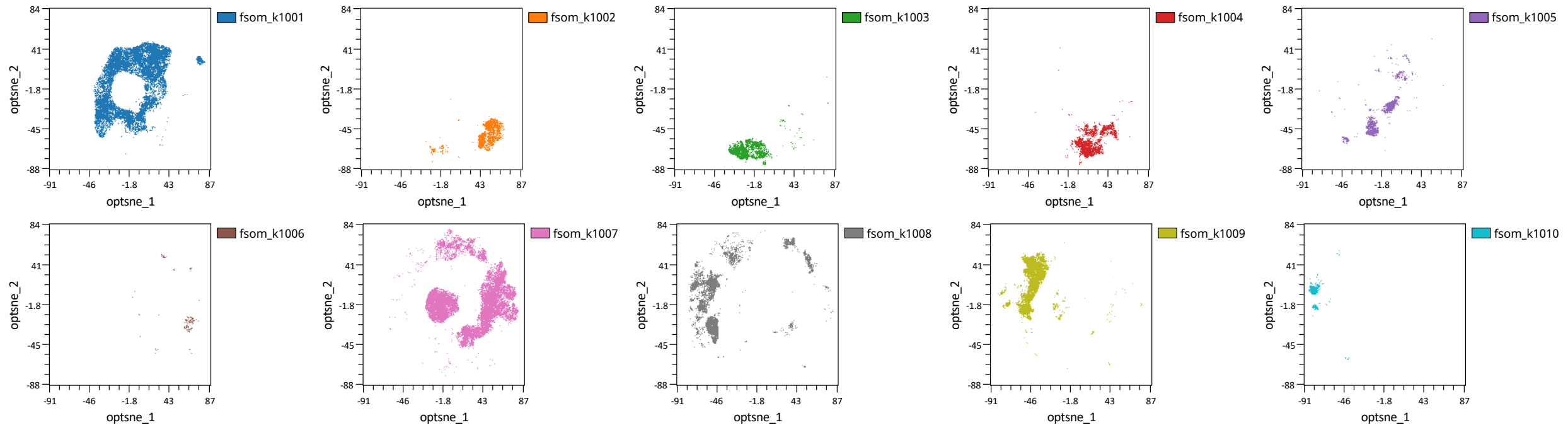


Clustering, allows us to derive multiple statistics for different populations

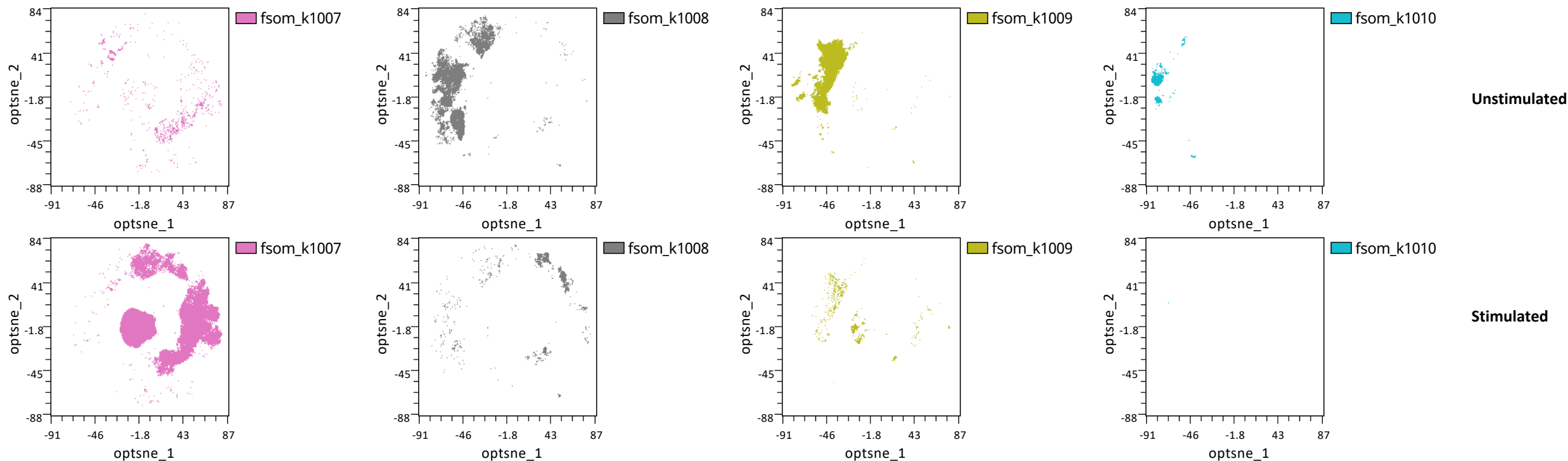




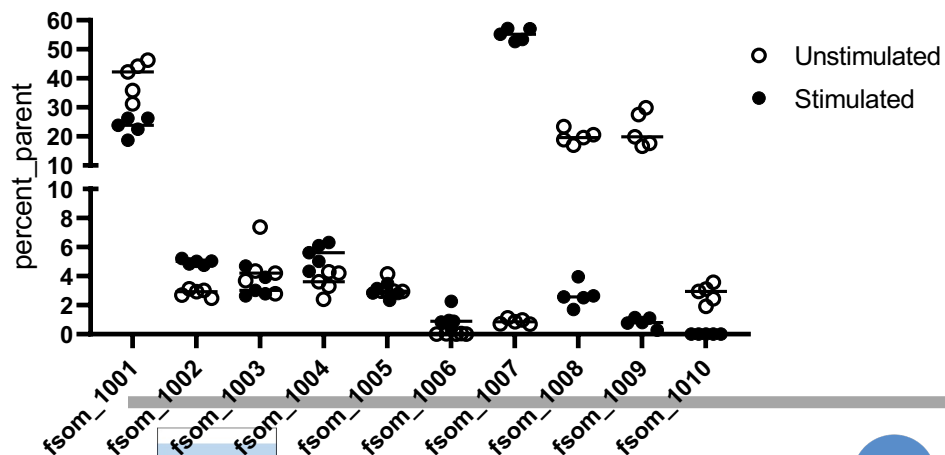
Clustering, individual clusters visualization (couple clicks in OMIQ)



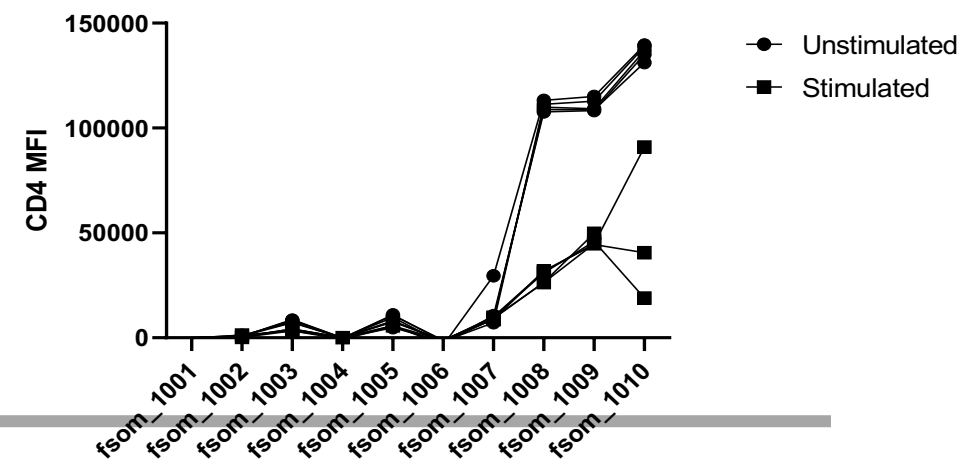
Clustering, focus on cluster 8 and 9



percent_among_CD3

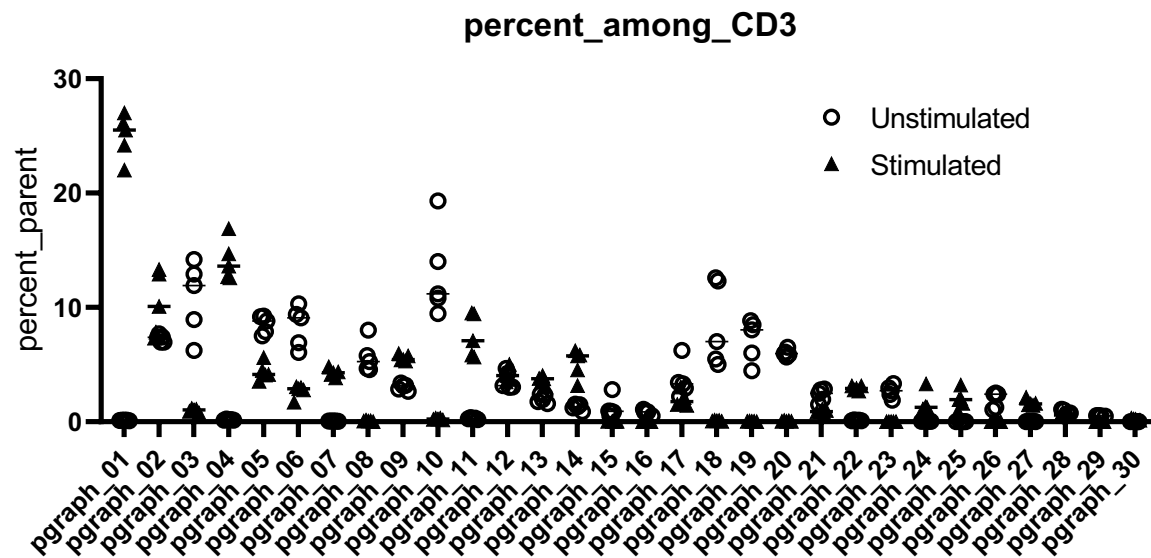
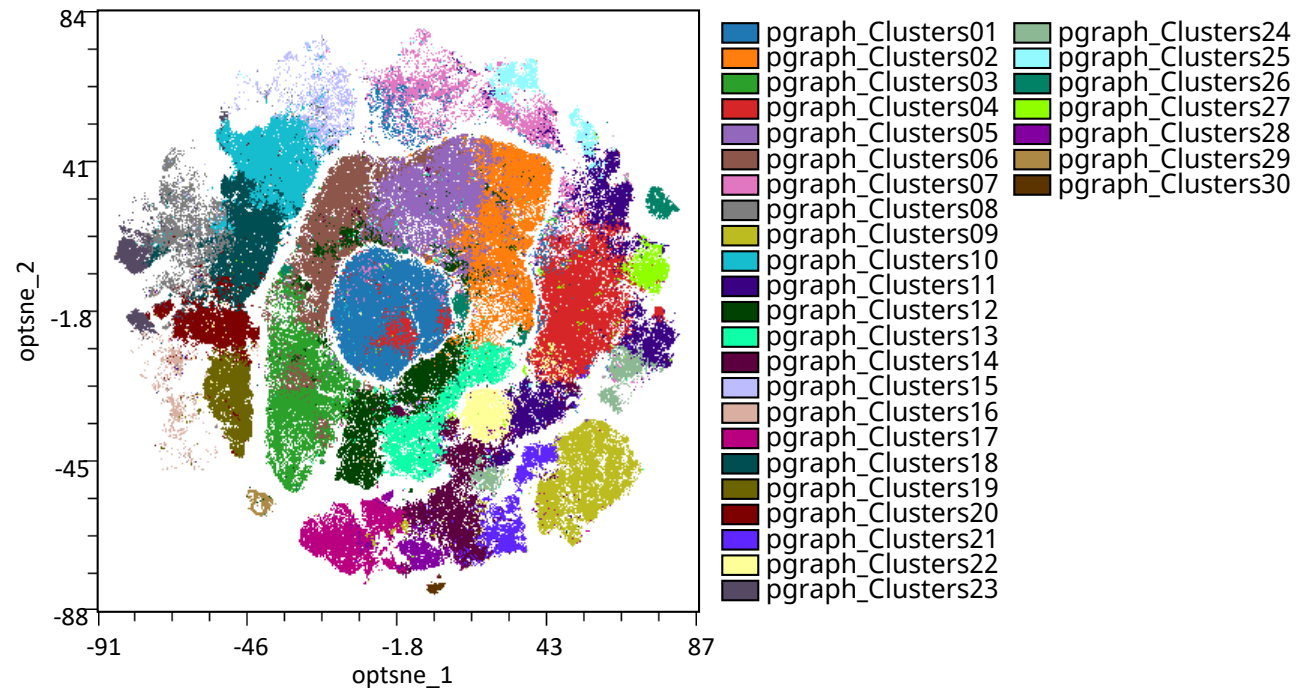


CD4_MFI_Unstimulated_vs_Stimulated

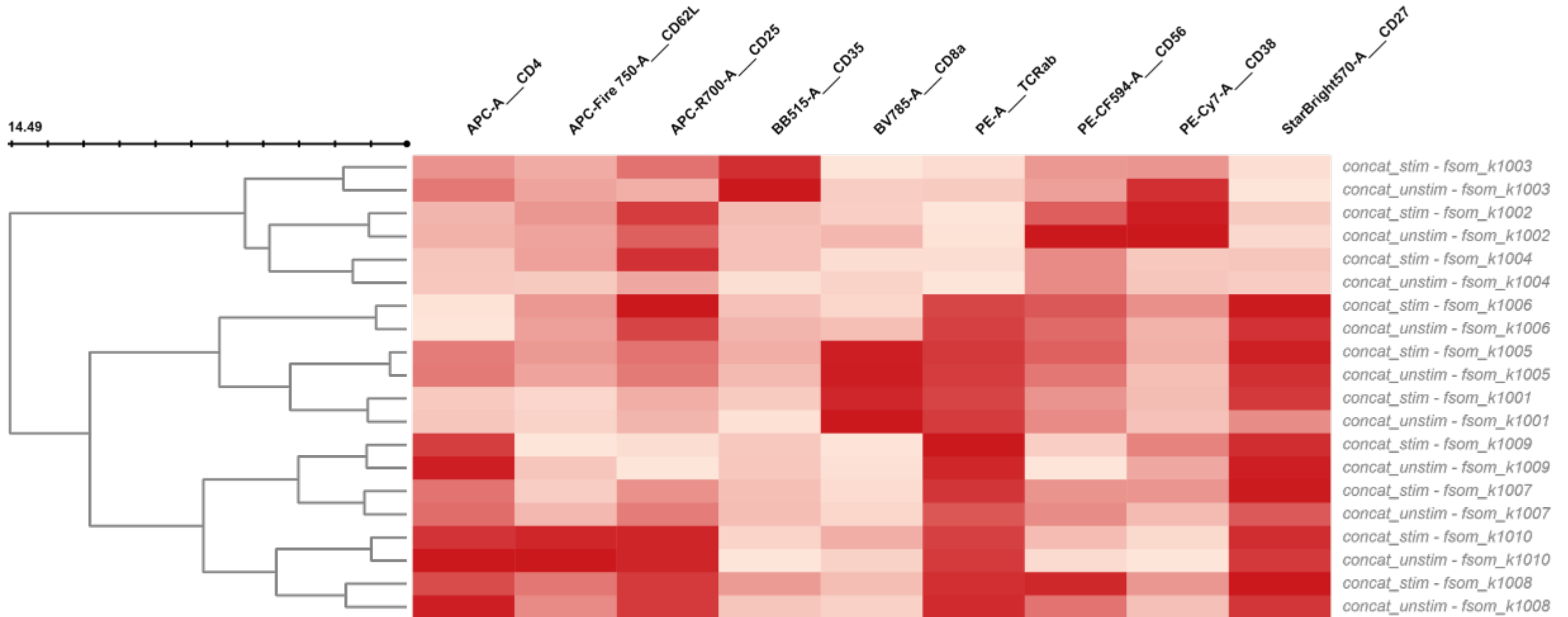


How many clusters to pick?

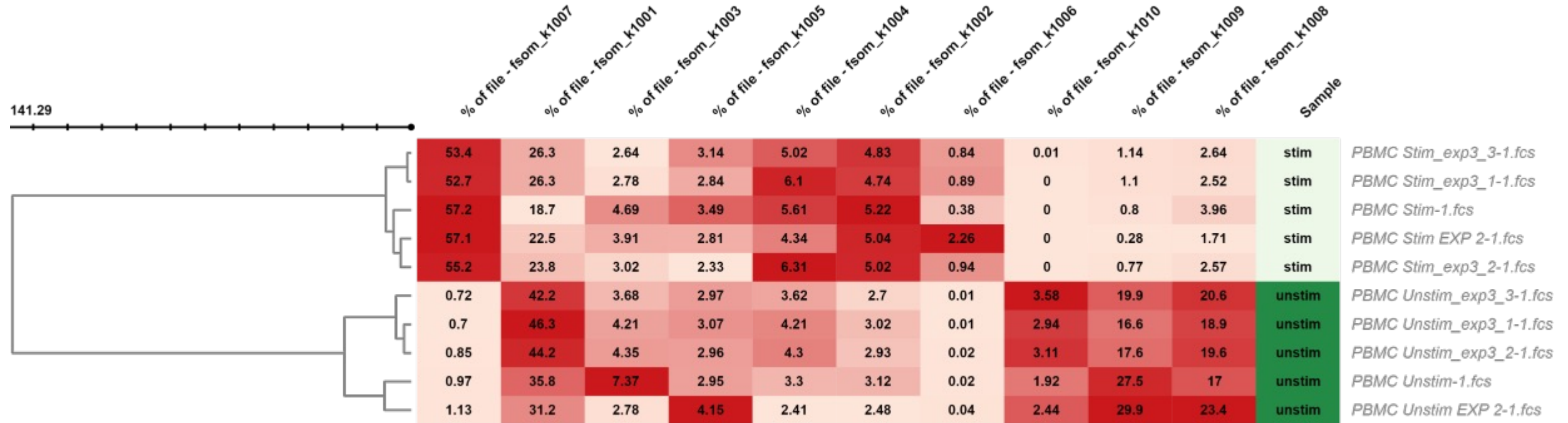
30 Clusters with Phenograph



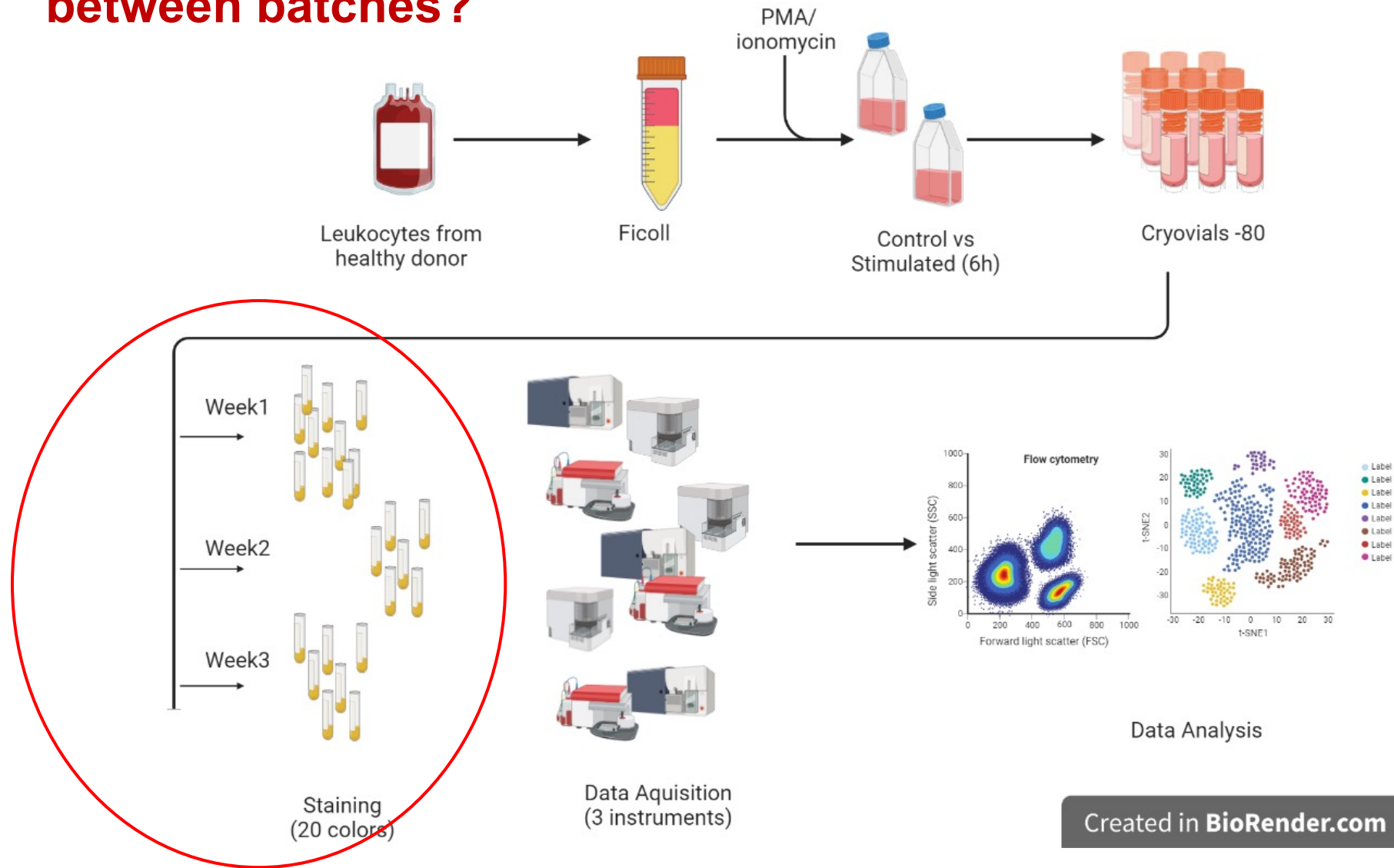
Clusters heatmap, results from OMIQ for advanced statistics (MFI)



Clusters heatmap results from OMIQ for advanced statistics (Abundance)



Can we identify differences between batches?



Created in **BioRender.com** **bio**



Week1
→



Week2
→



STAINED CONTROLS		
<input type="checkbox"/> From Library	Fluorescent Tag	Control
<input type="checkbox"/>	FITC	FITC (Beads) ▾
<input type="checkbox"/>	PE	PE (Beads) ▾
<input type="checkbox"/>	PerCP	PerCP (Beads) ▾

20 data files
10- unmixed with new set of
reference controls for each “week”

Week3
→



Unstimulated

Stimulated

10 Samples

Aurora - Unmixing 2x

10- unmixed using reference
controls recorded in library on week1



<https://www.chugcytometry.com>

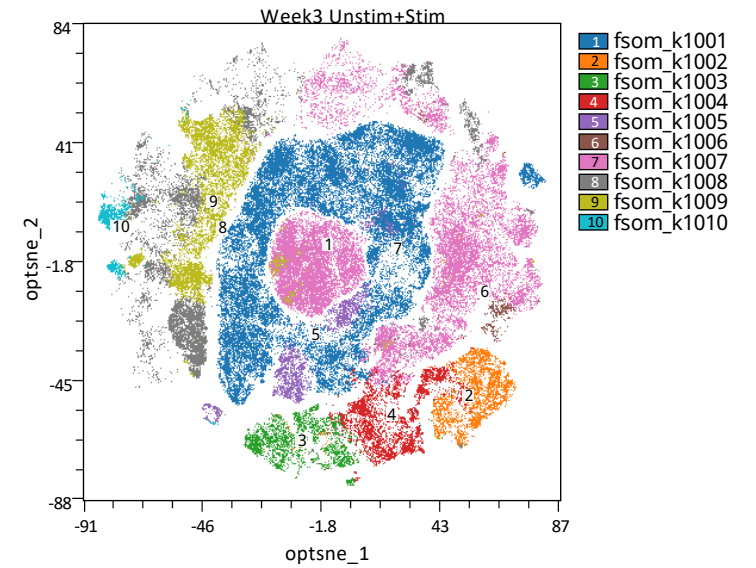
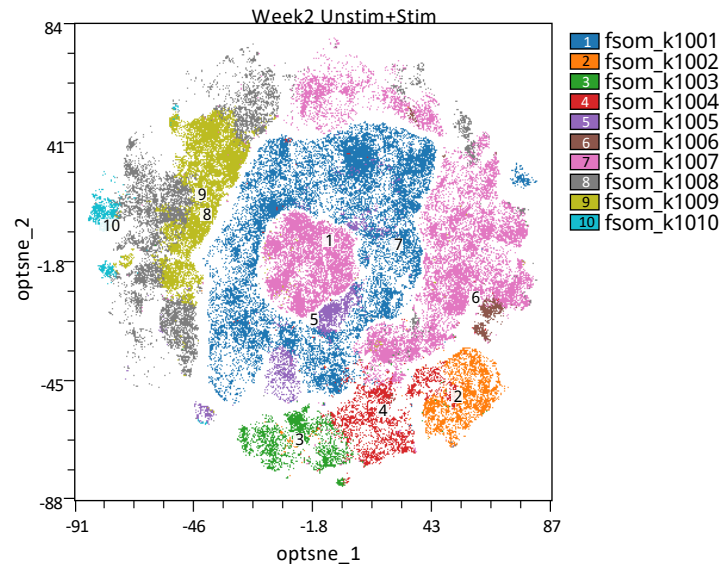
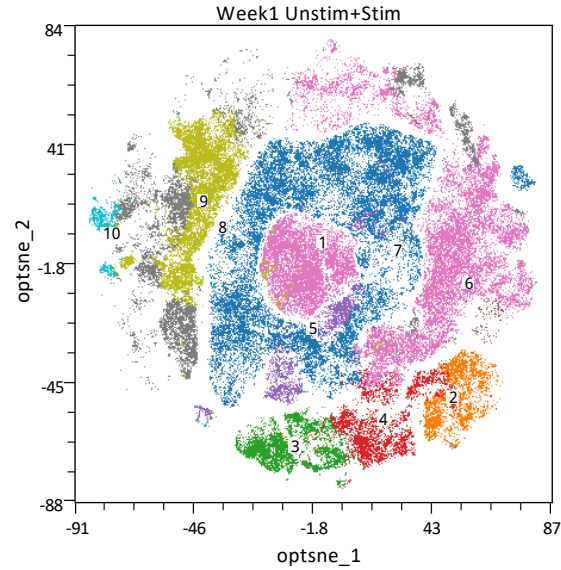


@chugcytometry

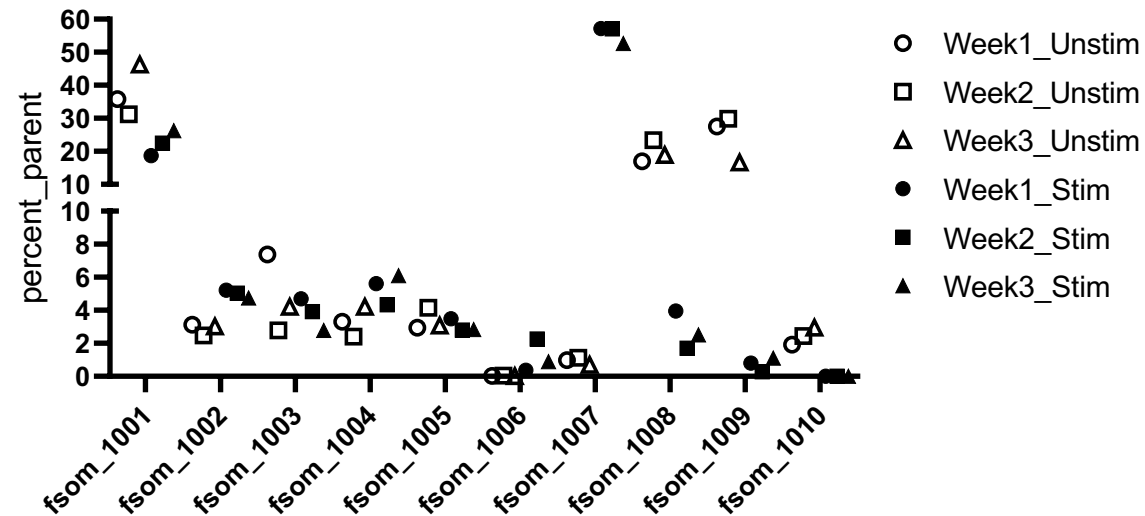


ChUG Cytometry

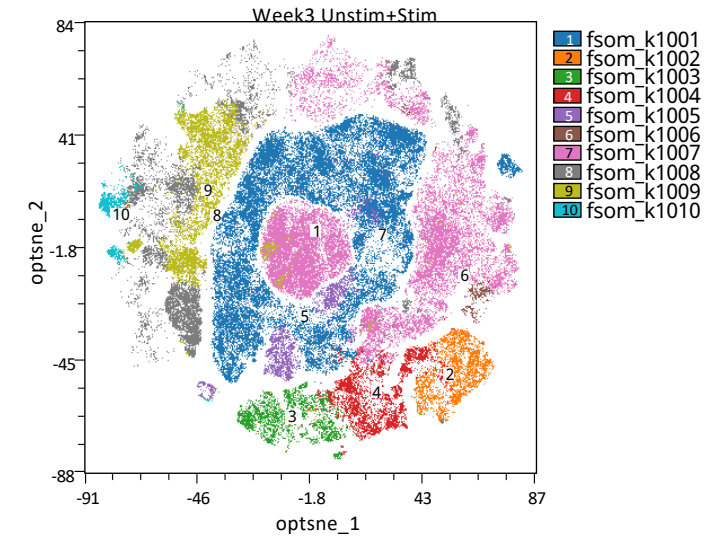
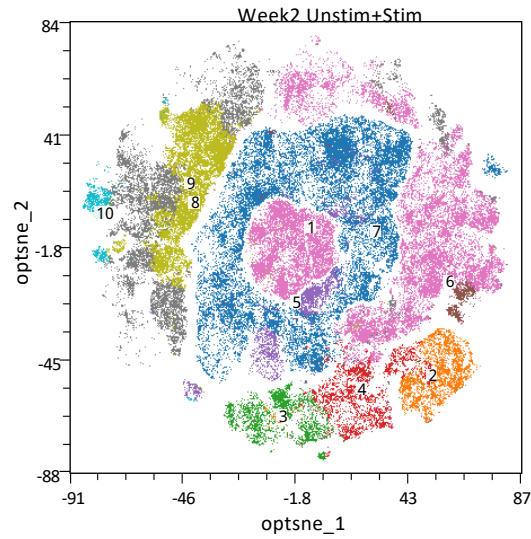
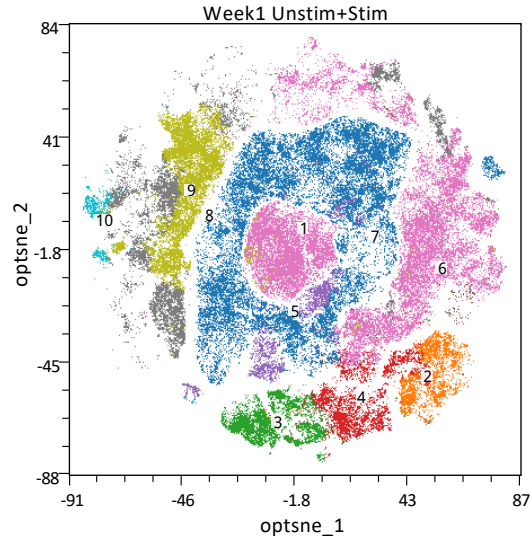
tSNE and Clustering overtime



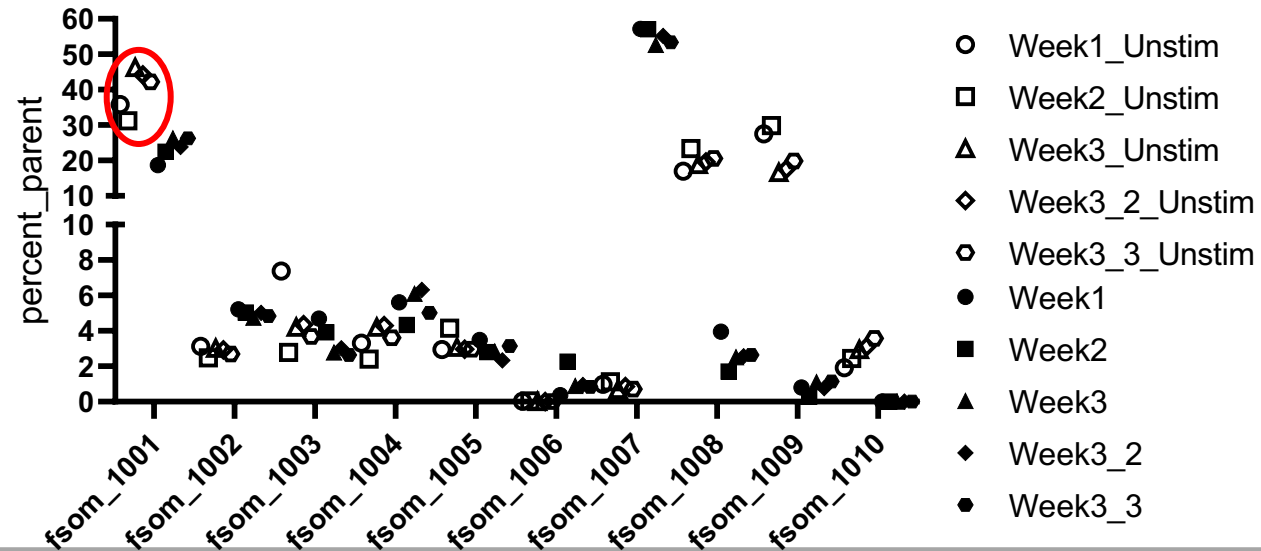
Week1 2 3 Unmixed experiment %CD3



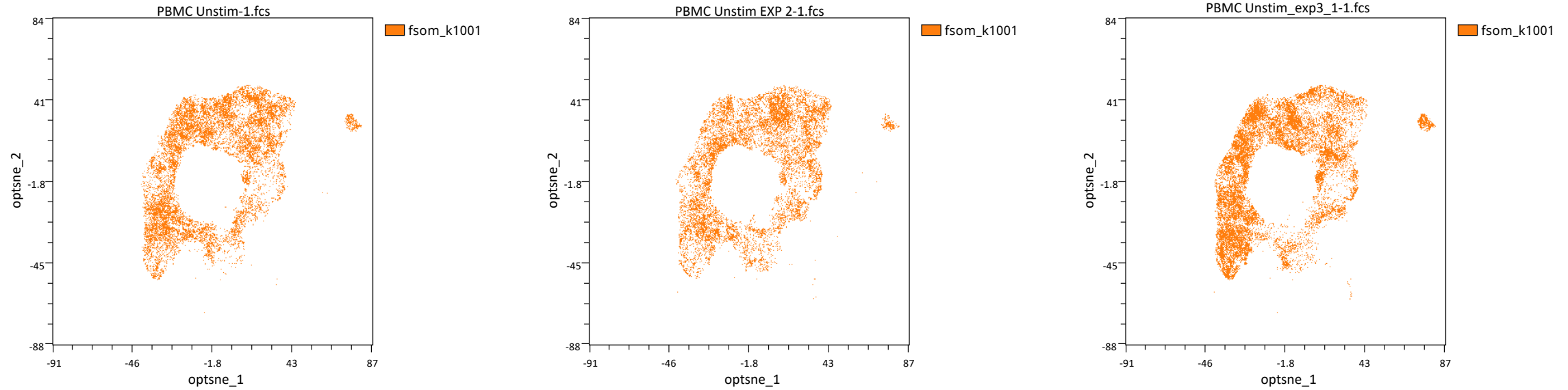
tSNE and Clustering overtime



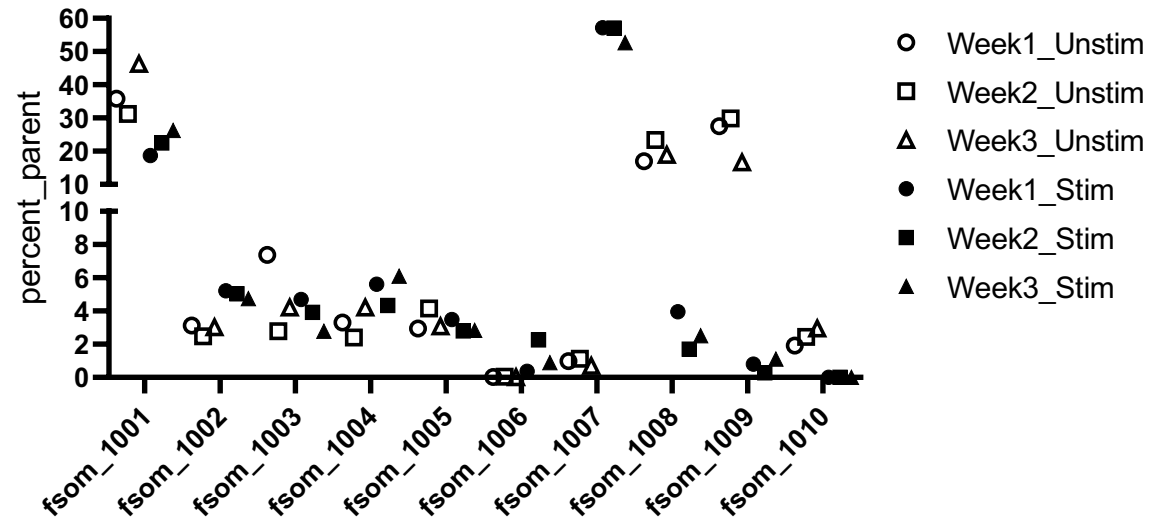
Week1 2 3 Unmixed experiment %CD3_n=5



tSNE and Clustering overtime

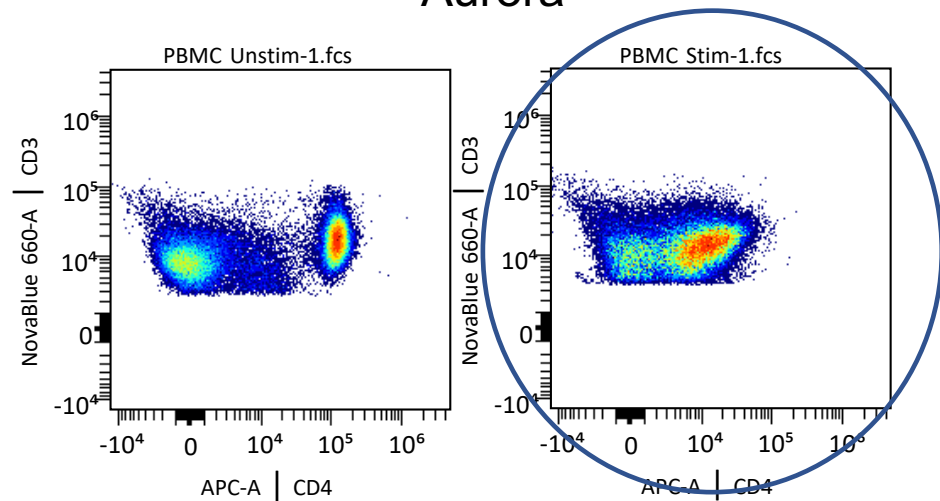


Week1 2 3 Unmixed experiment %CD3

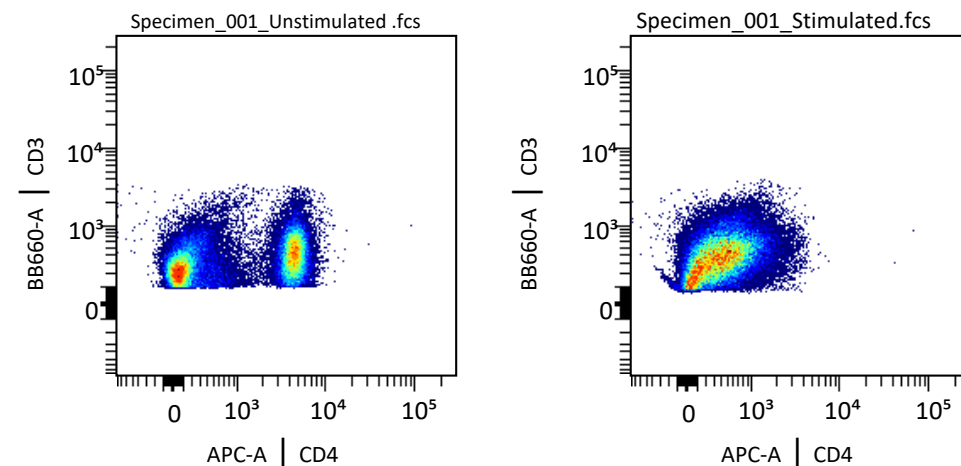


Instruments comparison, sensitivity

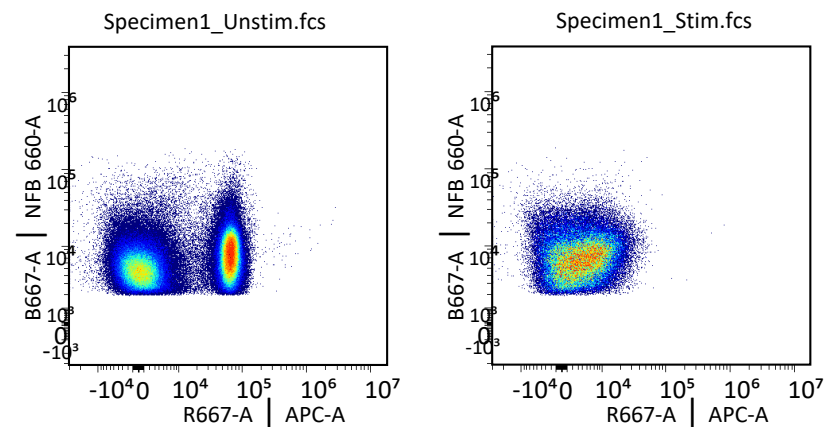
Aurora



Symphony S6



Penteon



BD Cell Sorter Symphony S6 Data (compensation matrix generated in OMIQ)

Week1

Channels	BUV395-A	BUV496-A	BUV737-A	BUV805-A	BV421-A	BV570-A	BV605-A	BV650-A	BV711-A	BV786-A	BB515-A	BB630-A	BB660-A	PE-A	PE-CF594-A	PE-Cy7-A	APC-A	APC-R700-A	APC-Cy7-A	BV510-A
BUV395-A	100	5.28	0.117	0	0.158	0.318	0.311	0.181	0.147	0	0	0.296	0.15	0	0	0	0	0	-0.124	0.655
BUV496-A	20.5	100	2.43	1	0.405	8.28	4.73	1.48	0.966	0.35	2.81	1.83	0.82	0	0.215	0	0.238	0.151	0.238	41
BUV737-A	2.69	0.507	100	46.6	0	0.167	0.117	0.13	2.39	1.79	0	0.163	0.161	0	0	1.69	0.246	15.6	33.9	0.384
BUV805-A	4.88	0.833	3.29	100	0	0.23	0.182	0.109	0.125	1.33	0	0.198	0	0	0	1.17	0	0.147	15.6	0.396
BV421-A	1.97	1.81	0.598	0.199	100	1.63	1.11	0.585	0.376	0	0.172	0.892	0.495	0	0.142	0.144	0	0	0.224	7.8
BV570-A	0.613	3.2	3.71	1.49	1.18	100	61.6	17.3	10.7	2.36	0.325	6.88	1.87	5.37	3.01	0.287	0	0	0	38.8
BV605-A	1.48	0.573	10.8	4.17	9.86	34.6	100	50.3	32.3	8.17	0	8.58	3.77	17.5	27	5.02	0.29	0.311	0.305	1.78
BV650-A	0.492	0.956	39.2	20.6	5.8	16.6	51.3	100	82.7	29	0	3.54	8.26	1.18	3.11	5.72	5	3.48	2.67	5.71
BV711-A	2.76	1.01	34.7	16.2	9.21	1.24	10.5	6.6	100	31.9	0.121	1	1.16	0.187	0.17	4.22	4.16	63.3	54.4	2.82
BV786-A	2.48	0.61	49.1	39.5	8.44	1.21	1.01	0.564	31.2	100	0	1.08	0.517	0.131	0.111	3.23	0.12	4.84	48.8	2.47
BB515-A	1.36	7.56	0.279	0.164	0	0.947	0.795	0.304	0.258	0	100	16.1	2.8	0	0	0.101	0	0	0.167	4.5
BB630-A	3.03	0.841	1.6	0.65	0.346	1.94	9.8	3.75	2.49	0.552	2.2	100	29.8	5.88	34.5	3.82	0.945	1.01	0.826	2.51
BB660-A	3.19	0.961	2.72	1.12	0	1.49	1.22	15.8	8.16	1.71	2.66	2.84	100	0	0.139	2.54	72	111	65	2.94
PE-A	0.35	0.141	0.84	0.279	0	11.4	5.26	1.38	0.852	0.151	0	43.4	8.9	100	45.3	2.61	0	0	0	0.414
PE-CF594-A	0.559	0.158	2.26	0.871	0	1.25	7.93	3.08	2.13	0.48	0	85	27.4	22.6	100	12.5	0.256	0.295	0.226	0.442
PE-Cy7-A	0.336	0.121	1.58	11.3	0	0.185	0.153	0	0.146	4.01	0	0.431	0.189	0.489	0.276	100	0	0.107	14.2	0.262
APC-A	1.7	0.453	12	3.92	0.239	0.681	0.979	11.4	5.7	1.29	0.155	0.864	2.75	0	1.15	9.11	100	71.3	46.6	1.29
APC-R700-A	0.498	0.141	11.7	4.15	0	0.204	0.16	0.624	5.66	1.46	0	0.18	0.313	0	0	9.79	8.71	100	67	0.37
APC-Cy7-A	0.2	0	1.34	13	0	0	0.192	0.17	3.22	0	0	0	0	0	0	17	1.29	1.5	100	0.157
BV510-A	-4.73	56.8	2.52	0.545	-1	33.3	21.1	5.44	2.61	0.391	-0.106	-0.469	-0.379	0	0	0	-0.165	-0.243	100	

Week2

Channels	BUV395-A	BUV496-A	BUV737-A	BUV805-A	BV421-A	BV570-A	BV605-A	BV650-A	BV711-A	BV786-A	BB515-A	BB630-A	BB660-A	PE-A	PE-CF594-A	PE-Cy7-A	APC-A	APC-R700-A	APC-Cy7-A	BV510-A
BUV395-A	100	5.59	0.302	0.148	0.369	0.515	0.444	0.26	0.155	0	0.108	0.411	0.242	0	0	0.101	0	0	0	0.832
BUV496-A	17.1	100	2.72	1.18	0.758	7.94	5.03	1.4	0.985	0.208	3.11	1.76	0.779	0.111	0.218	0.215	0	0.163	0.277	38.9
BUV737-A	2.25	0.45	100	44.2	0	0.147	0.152	0	1.84	1.35	0	0.162	0.149	0	0	1.66	0.216	12.4	26.5	0.355
BUV805-A	4.86	0.92	3.64	100	0.133	0.173	0.165	0.107	0	1.05	0	0.196	0	0	0	1.14	0	0.151	11.5	0.314
BV421-A	2.05	1.78	0.394	0.275	100	1.58	1.2	0.577	0.439	0.145	0.121	0.626	0.474	0.128	0.186	0.226	0.102	0.108	0.226	7.93
BV570-A	0.533	3.03	3.74	1.53	1.08	100	62	16.8	9.97	2.23	0.366	7.47	2.01	5.71	3.2	0.352	0	0	0.316	37.9
BV605-A	1.47	0.625	10.4	4.14	8.97	33.8	100	46.7	28.9	7.06	0.101	9.09	3.94	19.2	29.4	5.19	0.195	0.207	0	1.65
BV650-A	0.385	0.89	40.6	20.2	4.98	16.5	52.5	100	77.8	26.1	0	4.05	9.49	1.31	3.4	5.79	4.39	2.94	2.23	5.35
BV711-A	2.82	1.34	37.7	17.3	10.7	1.36	1.32	6.83	100	31.4	0.173	1.05	1.24	0.102	0.279	4.4	4.09	57.7	47.5	3
BV786-A	2.81	1.56	54.9	42.7	9.43	1.31	1.38	0.692	32.6	100	0.151	0.913	0.439	0.134	0.234	4.15	0.162	5.13	46.5	2.94
BB515-A	0.714	5.8	0.133	0	0	0.599	0.412	0.172	0.144	0	100	15.5	2.62	0	0	0	0	0	0	3.46
BB630-A	2.39	1	1.27	0.442	0.657	1.71	7.96	2.87	1.82	0.363	2.46	100	29.9	5.88	33.5	3.42	0.875	0.786	0.389	2.73
BB660-A	2.32	0.609	2.28	0.81	0.204	1.2	1.19	11.5	5.7	1.28	3.04	2.96	100	0	0.276	2.37	124	76.9	44	2.66
PE-A	0.26	0	0.599	0.21	0	8.35	3.82	0.937	0.566	0	0	44	8.78	100	44.3	2.37	0	0	0	0.356
PE-CF594-A	0.355	0.109	1.7	0.623	0	0.954	6.14	2.26	1.47	0.272	0	86.5	27.6	22.8	100	11.5	0.198	0.215	0.138	0.308
PE-Cy7-A	0.246	0	1.29	8.33	0	0.148	0.106	0	0	2.73	0	0.447	0.182	0.522	0.284	100	0	0	9.48	0.224
APC-A	1.47	0.61	11.9	4.02	0.454	0.756	0.909	10.6	5.04	1.2	0	0.61	3.12	0	1.39	10.3	100	68.6	43.2	1.5
APC-R700-A	0.436	0.152	12.1	4.16	0	0.207	0.207	0.641	5.37	1.29	0	0.187	0.406	0	0	12.2	9.02	100	62.9	0.455
APC-Cy7-A	0.391	0.217	1.33	13.1	0.229	0.128	0.142	0.182	0.178	3.13	0	0.112	0	0	0	14.4	1.1	1.33	100	0.276
BV510-A	-0.701	41.7	3.35	1.71	1.32	33.3	21.8	6.58	4.36	1.28	0.16	0.862	0.733	0.529	0.743	0.965	1.13	1.49	2.09	100

Week3

Channels	BUV395-A	BUV496-A	BUV737-A	BUV805-A	BV421-A	BV570-A	BV605-A	BV650-A	BV711-A	BV786-A	BB515-A	BB630-A	BB660-A	PE-A	PE-CF594-A	PE-Cy7-A	APC-A	APC-R700-A	APC-Cy7-A	BV510-A
BUV395-A	100	5.63	0.288	0.148	0.462	0.52	0.429	0.254	0.183	0	0	0.362	0.19	0	0	0	0	0	0.108	0.991
BUV496-A	17.8	100	2.73	1.22	0.835	8.12	5.08	1.55	1.13	0.225	2.99	1.75	0.776	0.134	0.319	0.257	0.159	0.199	0.279	39.5
BUV737-A	2.31	0.54	100	43.4	0.12	0.183	0.188	0.112	1.85	1.32	0	0.133	0.134	0	0	1.63	0.213	12.9	27.6	0.452
BUV805-A	4.64	0.945	3.54	100	0.165	0.241	0.206	0.131	0.102	1.09	0	0.184	0	0	0	1.21	0	0.189	12.9	0.529
BV421-A	2.32	2.29	0.404	0.279	100	1.74	1.41	0.521	0.427	0.134	0	0.726	0.38	0.116	0.254	0.158	0.145	0.196	0.122	8.39
BV570-A	0.53	2.98	3.58	1.4	1.09	100	61.5	16.3	9.64	2.1	0.342	7.09	1.91	5.73	3.16	0.284	0	0	0	37.3
BV605-A	1.55	0.761	10.4	4.05	9.11	33.5	100	46.4	29.2	6.99	0.107	9.04	3.98	19.4	29.1	5.11	0.306	0.283	0.291	1.85
BV650-A	0.435	0.945	40.2	19.4	5.17	16.4	52.5	100	78.4	26.1	0	3.8	9.08	1.32	3.37	5.8	4.68	3.18	2.45	5.51
BV711-A	3.32	1.61	36.9	16.4	9.77	1.19	1.31	6.63	100	30.7	0	0.937	1.06	0.167	0.221	4.48	4.18	62.4	52.6	3.26
BV786-A	2.59	1.39	54.8	42	9.57	1.26	1.18	0.763	32.2	100	0	0.783	0.518	0.107	0.244	3.96	0.177	5.08	48.6	3.04
BB515-A	0.685	6.29	0.187	0	0	0.718	0.482	0.195	0.106	0	100	15.3	2.62	0	0	0	0	0	0	3.71
BB630-A	2.43	1.12	1.41	0.676	0.545	2.05	8.55	3.04	1.95	0.451	2.51	100	30.1	6.15	34.2	3.72	0.877	0.907	0.388	3.16
BB660-A	2.76	1.21	2.64	0.811	0.612	1.41	1.15	12.6	6.26	1.31	2.92	2.74	100	0.184	0.385	2.45	148	87.6	51.4	3.08
PE-A	0.247	0	0.677	0.225	0	9.16	4.18	1.04	0.622	0.105	0	43.3	8.69	100	44	2.34	0	0	0	0.339
PE-CF594-A	0.367	0.174	1.93	0.715	0	1.03	6.65	2.47	1.64	0.351	0	85.7	27.2	23	100	11.7	0.256	0.259	0.159	0.371
PE-Cy7-A	0.275	0.11	1.39	8.94	0	0.156	0.14	0	0.111	2.94	0	0.397	0.175	0.558	0.302	100	0	0.103	10.9	0.258
APC-A	1.2	0.483	11.6	3.94	0.174	0.556	0.771	10.2	4.92	1.14	0	0.498	3.01	0.121	1.38	9.98	100	70.4	43.5	1.14
APC-R700-A	0.329	0.125	11.6	3.78	0	0.131	0.135	0.571	4.98	1.24	0	0.111	0.309	0	0	10.7	8.97	100	64.2	0.282
APC-Cy7-A	0.37	0.262	1.4	12.8	0.225	0.169	0.15	0.208	0.21	2.97	0	0.104	0.107	0	0	14.7	1.14	1.36	100	0.374
BV510-A	-1.13	46.4	2.81	1.12	0.382	32.7	21.5	6.31	3.96	1.26	-0.103	0	0.216	0.278	0.465	0.561	0.903	1.14	1.3	100



Panteon Data (compensation matrix generated in OMIQ)

Week1

Channels	B525-A	B615-A	B667-A	R667-A	R695-A	R725-A	UV445-A	UV525-A	UV725-A	UV780-A	V445-A	V525-A	V586-A	V615-A	V667-A	V725-A	V780-A	Y586-A	Y615-A	Y780-A
B525-A	100	2.69	0.542	0	0	0	0.235	2	0	0	0.13	1.43	0.186	0	0	0	0	0	0	0
B615-A	19.6	100	33.1	1.13	0.746	1.06	3.03	1.63	0.442	0	1.26	4.98	2.61	4.94	2.37	0.654	0	7.13	57.5	0.658
B667-A	21.2	2.95	100	123	41.9	64.7	3.17	1.66	0.538	0.336	1.23	5.59	1.69	0.541	9.39	2.17	0.482	-0.183	0.417	0.653
R667-A	1.1	0.861	2.88	100	25.6	62.3	1.82	0.68	2.54	0.753	0.856	2.45	0.745	0.374	7.74	2	0.371	0.188	2.42	2.89
R695-A	1.38	0.671	1.08	15.2	100	353	2.03	1.01	12.1	3.67	1.37	2.62	0.803	0.313	1.29	8.83	1.57	0	0.448	12.9
R725-A	6.34	4.14	4.07	51.4	18.1	100	7.43	3.27	3.33	66.9	1.59	10.7	3.38	1.06	4.18	3.9	40.5	1.6	7.98	236
UV445-A	0.478	0.282	0.153	0	0	0	100	6.55	0	0	0.705	1.09	0.289	0.108	0	0	0	0	0	0
UV525-A	9.82	0.903	0.477	0	0	0	23.1	100	0.523	0	3.46	35.2	3.98	0.947	0.378	0.202	0	0	0.256	0
UV725-A	1.37	0.655	0.547	0.384	5.64	84.3	20.3	2.48	100	50	1.1	2.62	0.856	0.243	0.167	6.12	2.03	0.177	0.422	1.89
UV780-A	2.29	1.37	0.941	-0.239	0.331	1.43	63.7	6.9	2.98	100	2.56	5.18	1.64	0.628	0.378	0	2.85	0.389	0.647	2.8
V445-A	1.1	0.61	0.361	0	0	0.129	11.9	1.5	0	0	100	8.74	1.05	0.27	0.191	0	0	0.122	0.242	0
V525-A	-0.234	0	0	0.304	0.186	0.365	-1.35	30.2	0.249	0.121	9.74	100	20.2	4.69	1.66	0.374	0	0	0.223	0
V586-A	6.13	7.04	2.14	0	0	0	0.894	13	1.47	0.483	6.23	73.2	100	23.3	8.9	2.3	0.483	9.89	4.71	0
V615-A	2.4	15.4	7.92	0.6	0.237	0.601	6.29	1.82	7.72	2.55	44.9	7.93	103	100	67.6	19.6	4.17	49.7	99.4	2.9
V667-A	0.897	7.04	17.6	7.89	2.42	5.07	5.63	3.2	31.1	13	73.5	29.2	42.8	45.4	100	42.4	12.7	3.4	10.8	3.4
V725-A	6.73	3.54	3.37	2.33	26.3	94.9	13	2.94	34.4	12.3	58	18.2	5	1.23	7.37	100	24.4	1.16	2.34	2.38
V780-A	8.56	4.35	4.44	1.31	1.26	33.4	16.6	5.66	31.1	48.4	81.7	25.5	7.08	2.37	2.57	103	100	2.53	1.73	3.65
Y586-A	2.71	35.6	8.82	0	0	0	0.288	0.203	0.118	0	0	0.893	23	3.91	1.28	0.307	0	100	35	0.375
Y615-A	0.725	75.5	27	0.333	0.158	0.223	0.499	0.239	0.355	0	0.135	0.637	2.54	6.28	2.89	0.843	0.189	21.2	100	2.33
Y780-A	1.3	1.39	0.629	0.15	0	1.23	0.99	0.56	0.192	5.01	0.404	1.65	0.836	0.17	0.15	0.45	4.97	3.91	1.63	100

Week2

Channels	B525-A	B615-A	B667-A	R667-A	R695-A	R725-A	UV445-A	UV525-A	UV725-A	UV780-A	V445-A	V525-A	V586-A	V615-A	V667-A	V725-A	V780-A	Y586-A	Y615-A	Y780-A
B525-A	100	2.73	0.542	0	0	0	0.174	2.11	0	0	0	1.37	0.154	0	0	0	0	0	0	0
B615-A	18.8	100	33.5	0.953	0.707	0.376	2.41	1.03	0.541	0.348	1.22	4.37	2.35	5.03	2.27	0.639	0.172	7.46	59.6	0.89
B667-A	20.9	2.62	100	124	42.5	65.1	2.05	0.875	0.752	0.123	0.773	4.67	1.24	0.419	9.31	2.22	0.564	0	0.524	0.407
R667-A	0.811	0.271	2.81	100	25.8	62.4	1.25	0.504	2.47	0.581	0.769	1.41	0.626	0.316	7.63	1.98	0.378	0.148	2.19	2.9
R695-A	0.966	0.62	0.922	15.8	100	357	1.75	0.629	12.5	3.63	0.884	1.83	0.483	0.236	1.16	8.5	1.67	0.247	0.243	13.1
R725-A	6.92	3.48	4.78	42.8	16.7	100	7.5	3.91	4.79	67.4	5.6	13.2	5.02	1.42	4.03	4.55	41.2	2.92	7.86	234
UV445-A	0.36	0.241	0.138	0.105	0	0	100	6.57	0	0	1.18	1.09	0.27	0.188	0.111	0	0	0.104	0.189	0
UV525-A	9.66	0.851	0.423	0	0	0	23.7	100	0.424	0	3.61	35.2	3.97	0.939	0.323	0	0.111	0.144	0.344	0
UV725-A	1.11	0.771	0.312	0.355	5.91	83.5	19.8	2.29	100	49.1	0.236	1.64	0.621	0.235	0.266	5.95	2.02	0	0.176	2
UV780-A	1.44	1.07	0.461	0.282	0.223	1.71	70.2	8.15	3.39	100	2.64	3.87	1.2	0.319	0.195	0.26	2.72	0.185	0.878	3.13
V445-A	0.604	0.485	0.342	0	0	0.112	10.9	1.35	0	0	100	7.8	0.853	0.303	0.147	0.147	0	0.205	0	0
V525-A	0.75	0.505	0.36	0.359	0.279	0.474	-5.16	27.5	0.403	0.12	10	100	20.3	4.81	1.83	0.473	0.14	0.285	0.452	0.127
V586-A	5.94	6.93	2.17	0	0	0	1.14	13.1	1.56	0.486	6.21	74.4	100	23.3	8.97	2.29	0.477	9.72	4.64	0.182
V615-A	1.44	14.3	7.66	0.553	0.542	0.371	6.45	1.66	7.85	2.57	43.7	7.23	102	100	67.6	19.3	4.06	50	96.9	3.03
V667-A	0.779	6.98	17.4	8.16	2.5	5.13	5.69	3.24	31.2	13.2	73.9	29	42.9	45	100	42.5	12.7	3.6	11.3	3.43
V725-A	4.32	2.03	3.84	2.77	25.5	90.9	11.7	4.26	33.7	13.2	64.6	16.1	4.28	1.32	6.72	100	23.6	1.61	1.35	2.61
V780-A	4.23	4.45	1.6	0.293	-0.776	33.5	14.8	5.15	31	45.6	74.9	14	4.22	1.91	0.484	99.7	100	1.66	1.83	2.76
Y586-A	2.68	34.2	8.64	0	0	0	0.288	0.236	0.114	0	0.136	0.885	22	3.79	1.22	0.265	0	100	35.3	0.452
Y615-A	0.494	74.9	26.7	0.286	0.159	0.312	0.267	0.203	0.231	0	0.182	0.418	2.42	6.1	2.8	0.784	0.163	21.2	100	2.26
Y780-A	0.98	1.22	0.557	0	0.214	1.22	1.05	0.488	0.269	5.1	0.61	1.07	0.55	0	0.138	0.373	5.1	3.95	1.76	100

Week3

Channels	B525-A	B615-A	B667-A	R667-A	R695-A	R725-A	UV445-A	UV525-A	UV725-A	UV780-A	V445-A	V525-A	V586-A	V615-A	V667-A	V725-A	V780-A	Y586-A	Y615-A	Y780-A
B525-A	100	2.69	0.522	0	0	0	0.145	1.92	0	0	0	1.26	0.131	0	0	0	0	0	0	0
B615-A	17.7	100	33.6	1.01	0.686	0.536	2.18	0.834	0.113	0.194	1.06	3.99	2.48	4.54	2.09	0.705	0.114	7.07	58.6	0.926
B667-A	20.2	2.35	100	121	42.1	64.2	2.64	1.05	0.647	0.274	0	3.71	1.22	0.463	9.06	2.07	0.491	0.467	0.434	0.728
R667-A	0.342	0.332	2.69	100	26.3	63	0.97	0.28	2.47	0.813	0	1.08	0.337	0.247	7.77	2.04	0.397	0.243	2.69	2.99
R695-A	0.495	0.187	0.896	15.7	100	353	0.903	0.329	12.3	3.55	0.597	1.42	0.484	0.105	1.15	8.56	1.56	0.158	0.387	13
R725-A	3.07	2.53	3.74	42.4	16.9	100	9.11	4.58	4.02	65.9	0.886	13.9	3.24	1.52	4.23	4.61	40.5	2.12	8.2	230
UV445-A	0.247	0.163	0.146	0	0	0	100	6.55	0	0	0.827	0.89	0.248	0	0	0	0	0.145	0.211	0
UV525-A	9.4	0.725	0.399	0	0	0	23.4	100	0.467	0.146	2.88	35.1	3.98	0.934	0.337	0.104	0	0.128	0.209	0
UV725-A	0.411	0.401	0.26	0.298	5.7	83.6	20.2	2.57	100	49.6	0.05	1.75	0.516	0.245	0.19	6.07	1.93	0.181	0.341	1.9
UV780-A	0.834	0.855	0.548	0	0.12	1.55	66.5	7.22	3.02	100	1.92	3.67	1.14	0.45	0	0	2.7	0.513	0.677	2.66
V445-A	0.415	0.186	0.203	0	0	0	11.3	1.25	0	0	100	7.49	0.848	0.206	0.114	0.105	0	0	0.268	0
V525-A	0.142	0.13	0.143	0.232	0.155	0.273	-3.91	28.7	0.226	0	9.34	100	19.9	4.69	1.75	0.402	0.108	0	0.265	0
V586-A	5.78	6.94	2.03	0	0	0	0.837	13	1.53	0.463	6.19	74.5	100	23.6	8.99	2.29	0.484	10.1	4.58	0
V615-A	1.32	13.9	7.42	0.44	0.501	0.285	5.48	1.22	7.55	2.3	45.6	6.33	103	100	68.4	19.1	3.97	50.4	99.3	2.88
V667-A	0.499	6.69	17.4	7.76	2.39	5.12	5.48	2.99	30.6	13.1	72.9	28.9	42.3	44.6	100	41.9	12.6	3.44	11	3.3
V725-A	2.43	2.99	1.33	2.63	26.8	92.3	11.7	4.19	34.4	12.1	60.2	15.3	4.1	1.11	7.7	100	23.6	0.918	1.2	2.84
V780-A	3.9	2.85	1.52	-0.386	0.963	33.1	13.1	5.33	31.7	46.7	78.1	17.8	5.59	1.47	0.972	102	100	0	1.71	2.81
Y586-A	2.65	35.6	8.85	0	0	0	0.219	0.186	0.126	0	0	0.817	23.2	3.98	1.26	0.286	0	100	35.3	0.38
Y615-A	0.442	75.5	27	0.316	0.195	0.254	0.301	0.105	0.363	0	0	0.37	2.43	6.23	2.88	0.784	0.186	21.2	100	2.21
Y780-A	0.84	1.02	0.539	0	0	1.13	0.735	0.433	0.299	4.99	0.477	1.13	0.53	0.127	0.132	0.445	4.97	4.06	1.76	100

Symphony S6 – is comparing cell sorters to analyzers fair?



<https://www.chugcytometry.com>



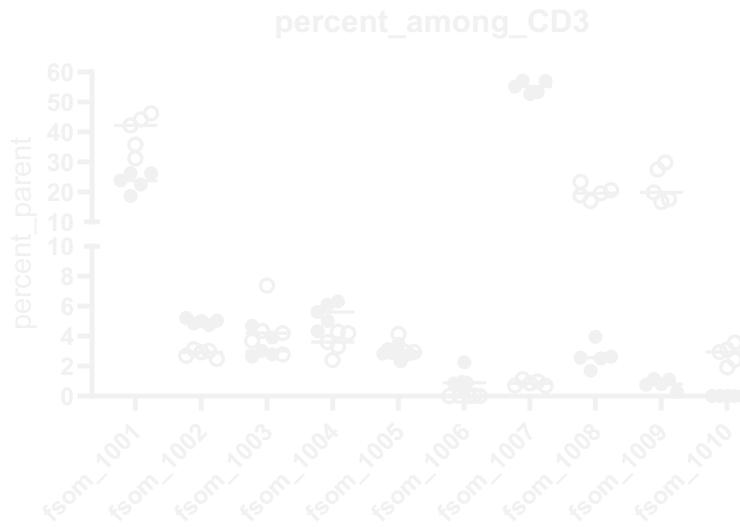
@chugcytometry



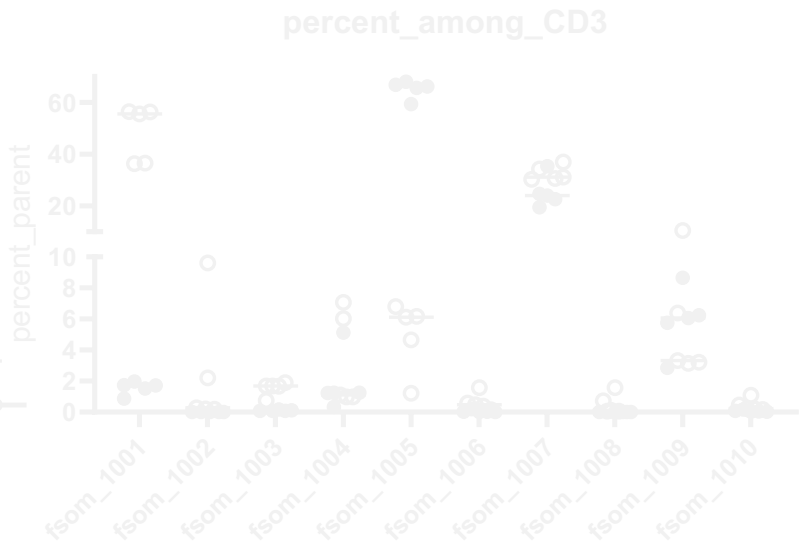
ChUG Cytometry

Instruments comparison

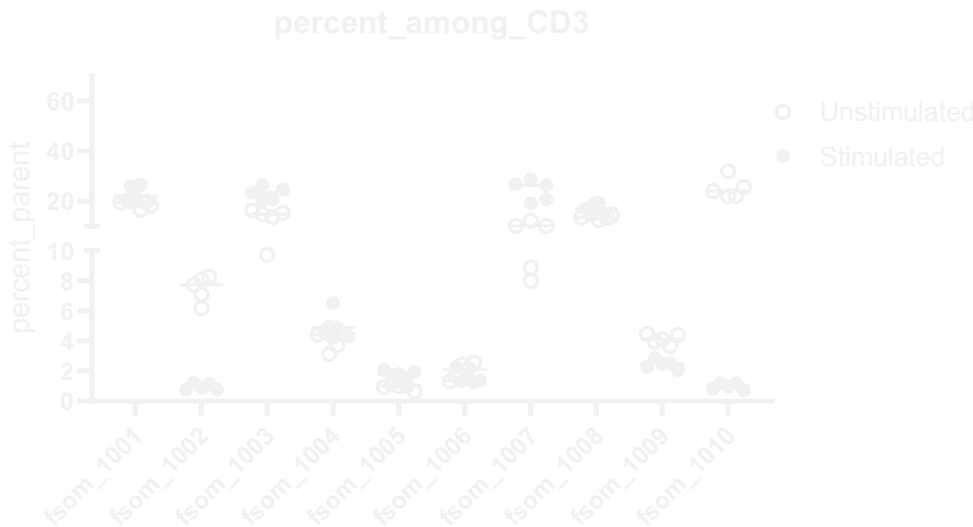
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Symphony S6



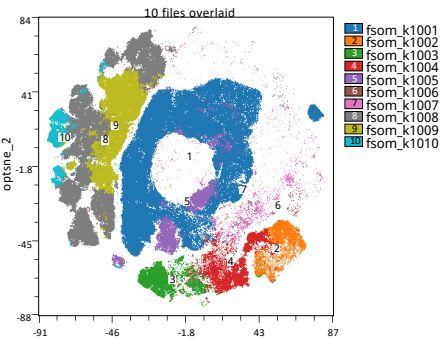
Penteon



Symphony S6 – is comparing cell sorters to analyzers fair?

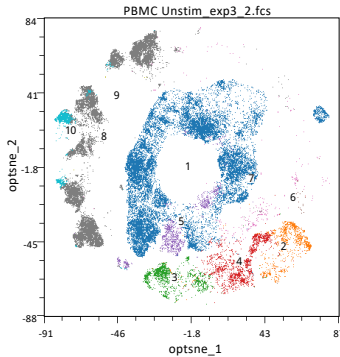
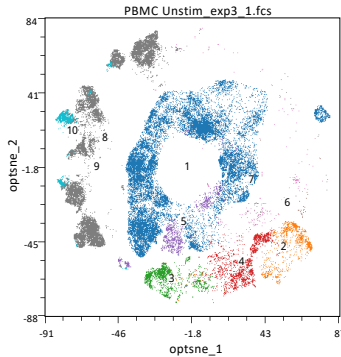
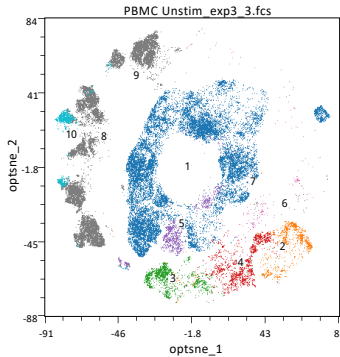
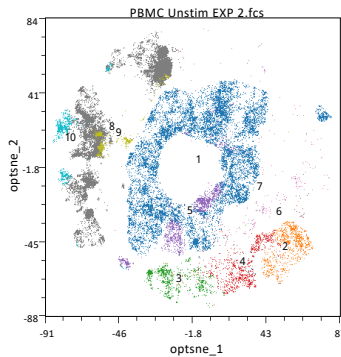
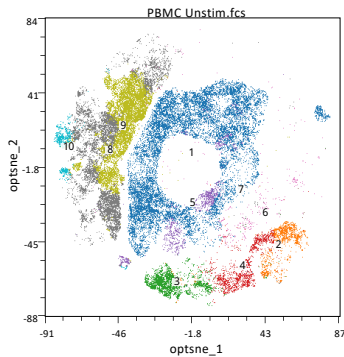


Library vs experiment reference controls unmixing analysis, Unstimulated samples:

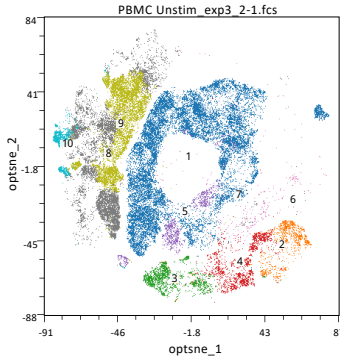
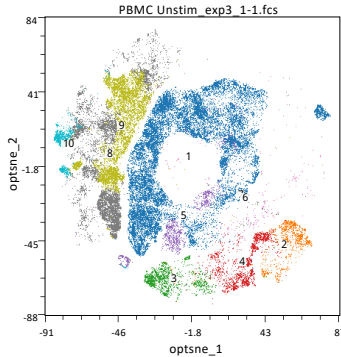
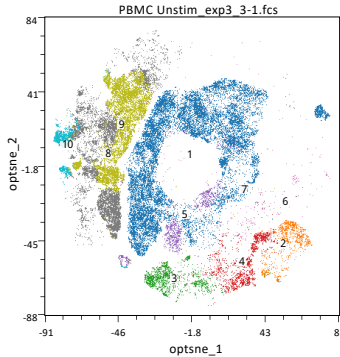
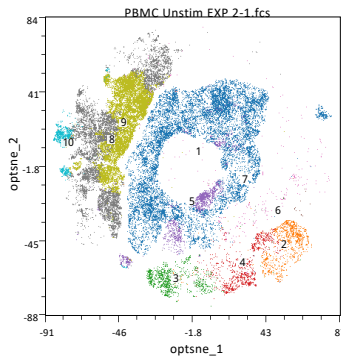
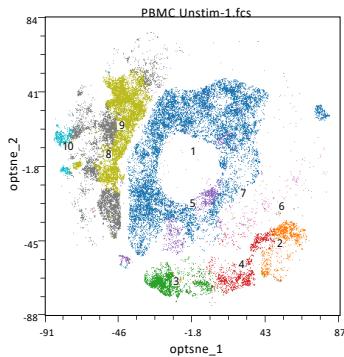


STAINED CONTROLS

From Library	Fluorescent Tag	Control
<input type="checkbox"/>	FITC	FITC (Beads) ▼
<input type="checkbox"/>	PE	PE (Beads) ▼
<input type="checkbox"/>	PerCP	PerCP (Beads) ▼



Library

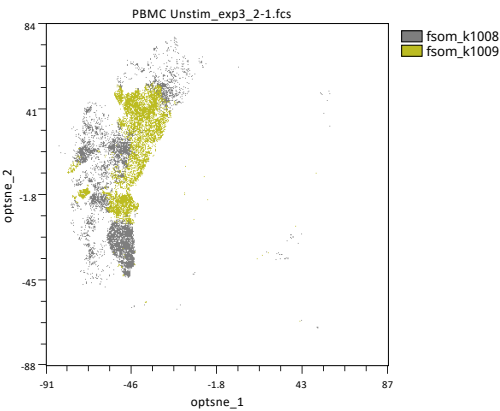


Experiment

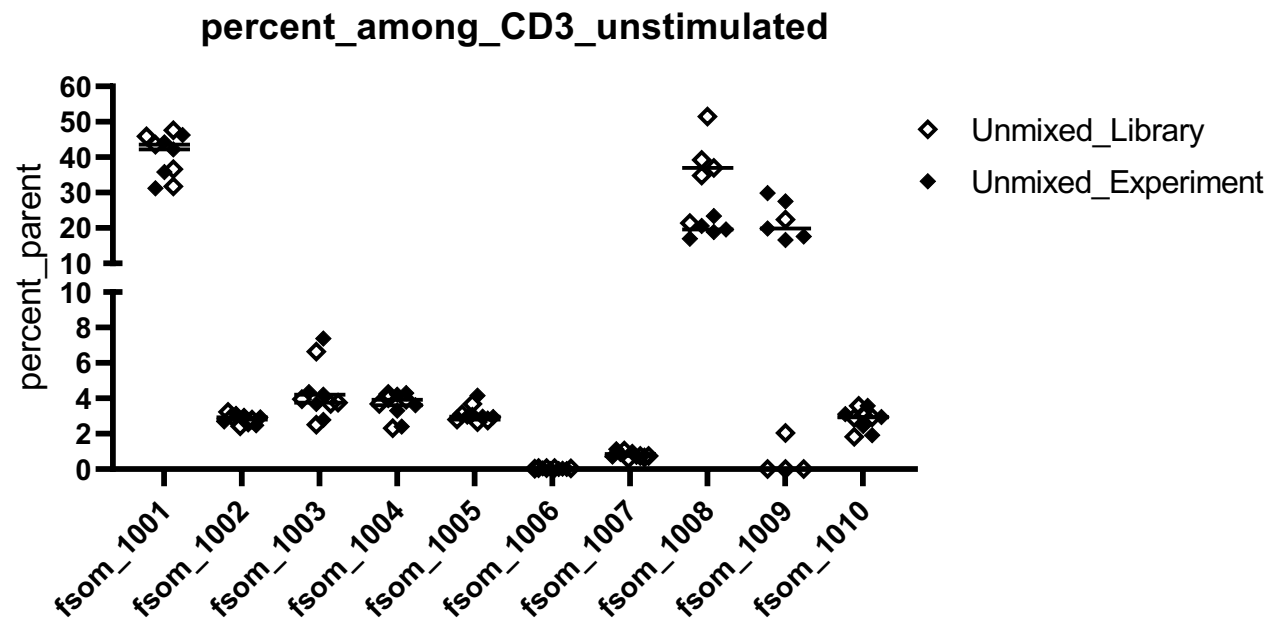
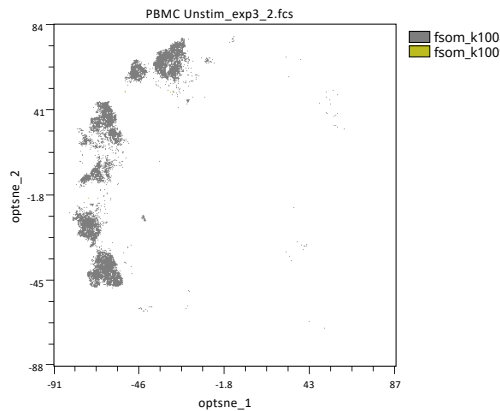


Library vs experiment reference controls unmixing analysis Unstimulated samples:

Experiment



Library



Normalization

Data normalization refers to the process of transforming variables to fit a limited range of values and create consistency between samples.



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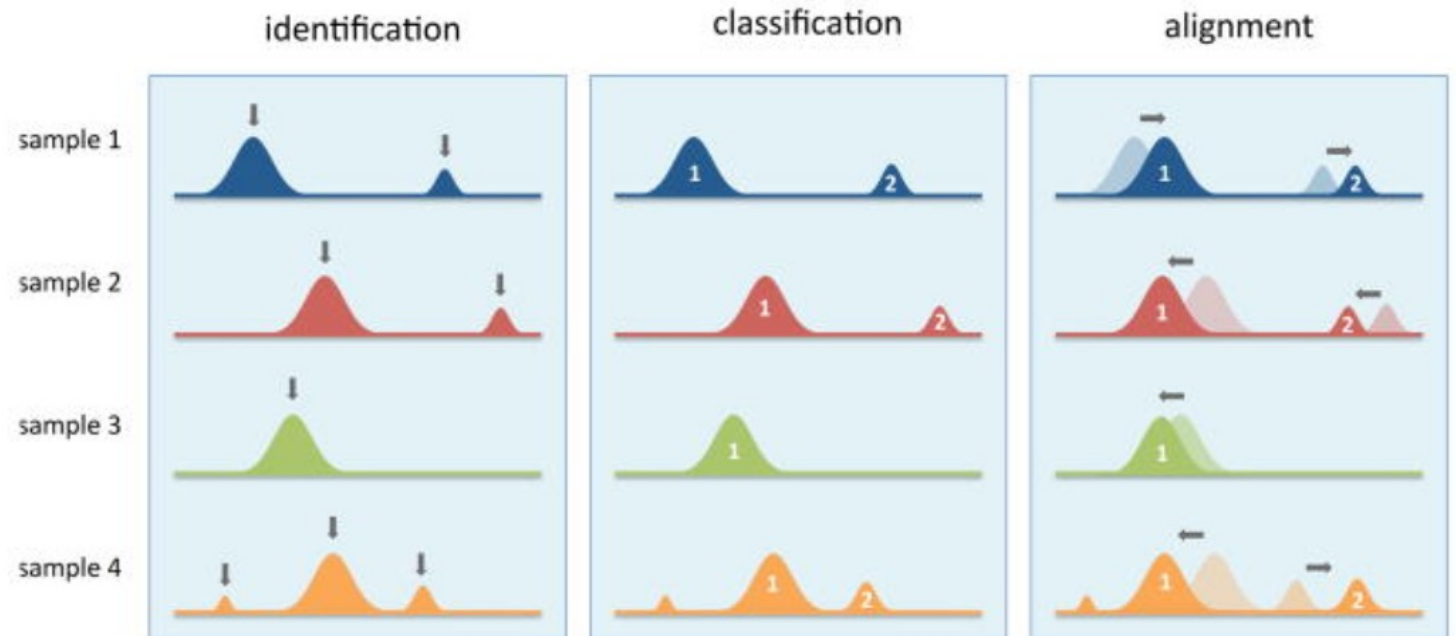
Normalization in OMIQ

Normalization



<https://doi.org/10.1002%2Fcyto.a.20823>

<https://doi.org/10.1002%2Fcyto.a.23904>



<https://www.chugcytometry.com>

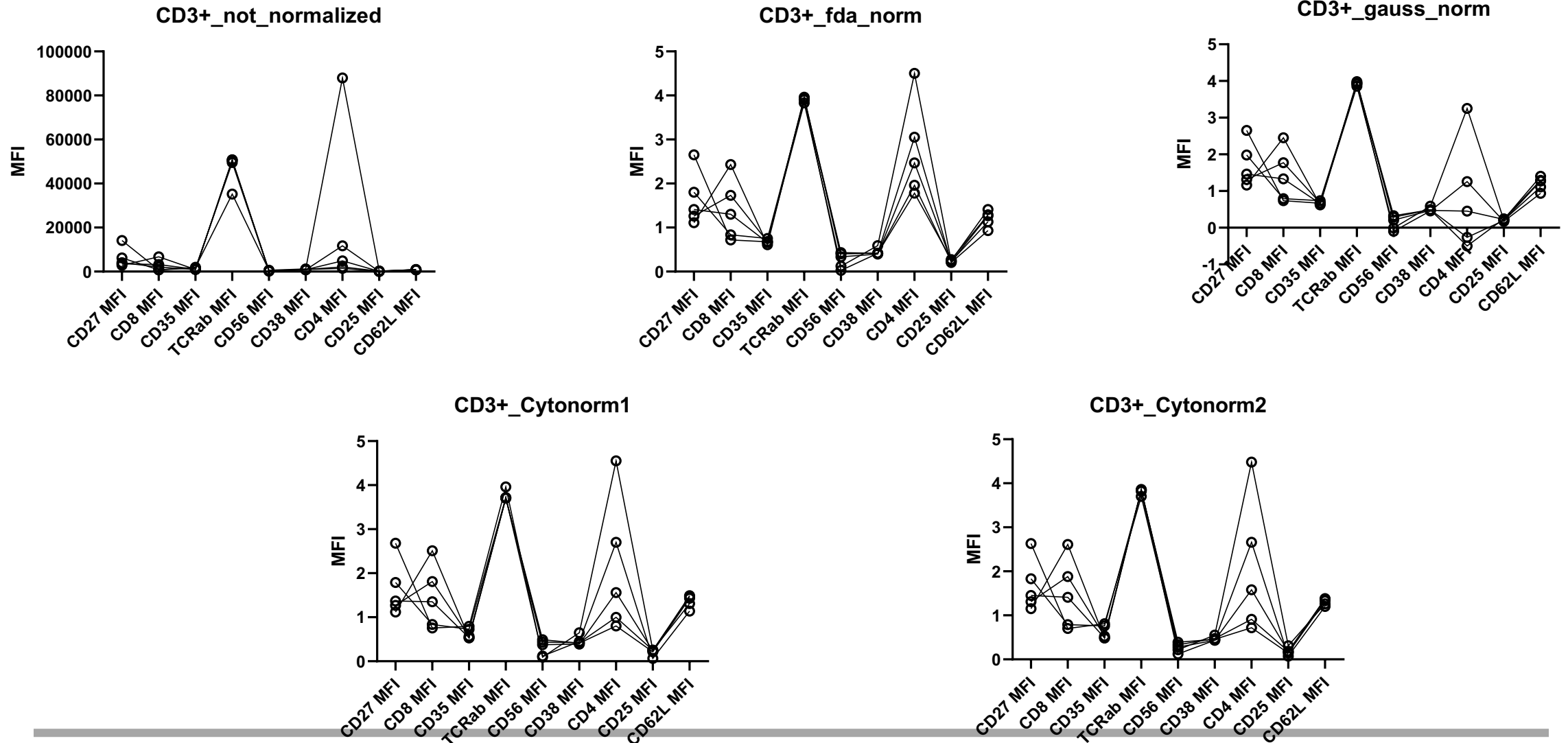


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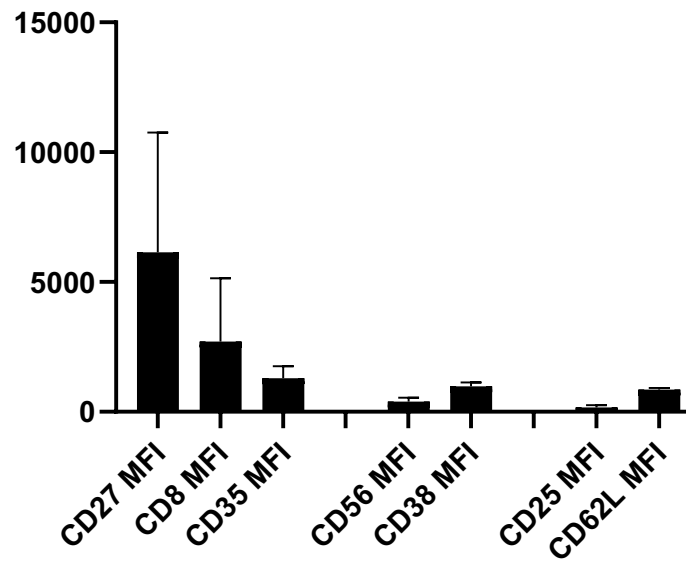
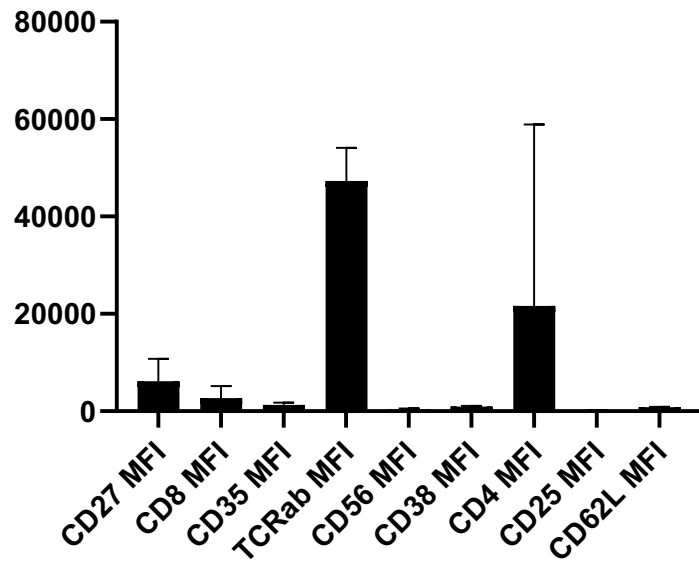


ChUG Cytometry

normalization

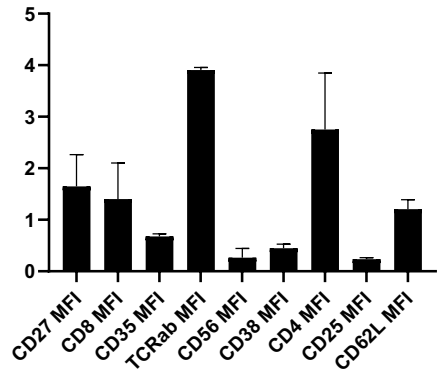


MFI_Unstimulated_not_normalize



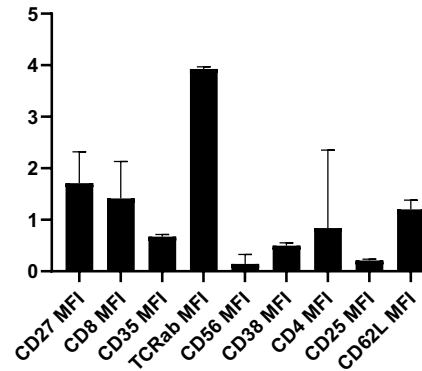
	Mean	%CV
CD27 MFI	6147.8	74.9
CD8 MFI	2705	90.3
CD35 MFI	1295	35.8
TCRab MFI	47323.6	14.3
CD56 MFI	390.6	40.1
CD38 MFI	980.4	15.1
CD4 MFI	21625.6	172.4
CD25 MFI	169.56	52.0
CD62L MFI	853.4	7.8

MFI_Unstimulated_fda_norm



	Mean	%CV
CD27 MFI	1.646	37.4
CD8 MFI	1.4028	49.9
CD35 MFI	0.6758	7.5
TCRab MFI	3.908	1.2
CD56 MFI	0.26422	67.7
CD38 MFI	0.447	17.8
CD4 MFI	2.752	39.7
CD25 MFI	0.2368	10.9
CD62L MFI	1.2066	15.1

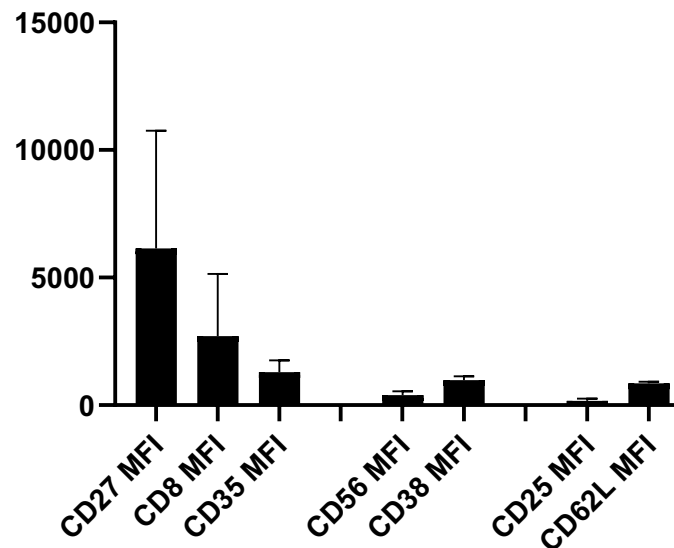
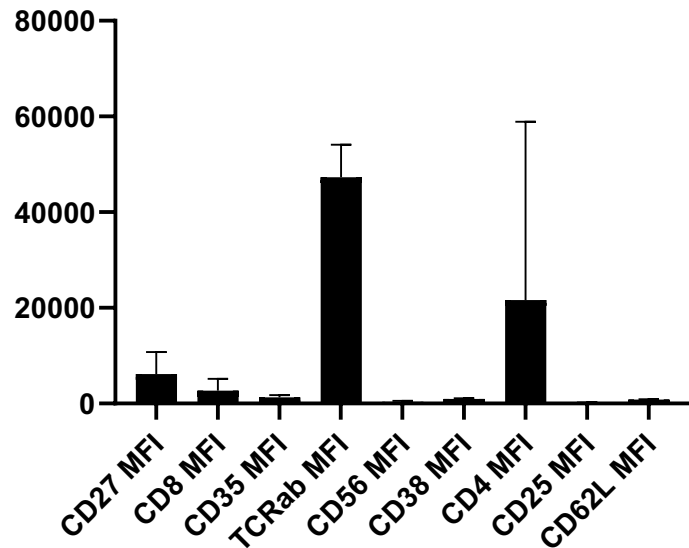
MFI_Unstimulated_gauss_norm



	Mean	%CV
CD27 MFI	1.712	35.5
CD8 MFI	1.416	50.5
CD35 MFI	0.6728	6.4
TCRab MFI	3.926	1.1
CD56 MFI	0.14238	130.8
CD38 MFI	0.4998	10.4
CD4 MFI	0.8396	180.1
CD25 MFI	0.2086	12.7
CD62L MFI	1.2014	15.0

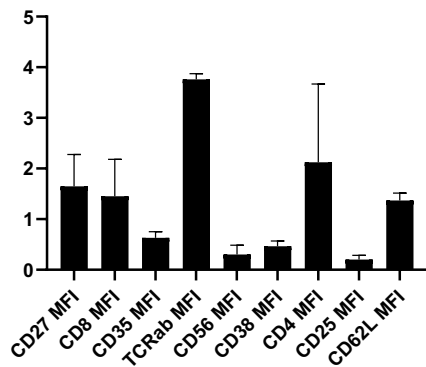


MFI_Unstimulated_not_normalize



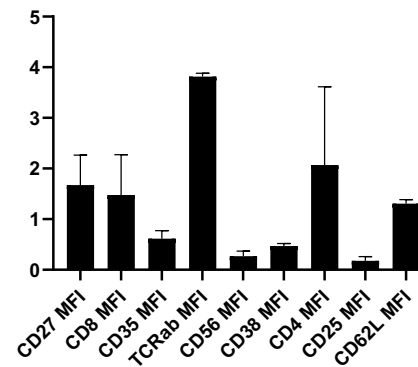
	Mean	%CV
CD27 MFI	6147.8	74.9
CD8 MFI	2705	90.3
CD35 MFI	1295	35.8
TCRab MFI	47323.6	14.3
CD56 MFI	390.6	40.1
CD38 MFI	980.4	15.1
CD4 MFI	21625.6	172.4
CD25 MFI	169.56	52.0
CD62L MFI	853.4	7.8

MFI_Unstimulated_Cytonorm1



	Mean	%CV
CD27 MFI	1.646	38.2
CD8 MFI	1.453	50.0
CD35 MFI	0.630	19.3
TCRab MFI	3.76	2.9
CD56 MFI	0.303	60.7
CD38 MFI	0.462	22.7
CD4 MFI	2.122	72.8
CD25 MFI	0.203	39.1
CD62L MFI	1.368	10.6

MFI_Unstimulated_Cytonorm2

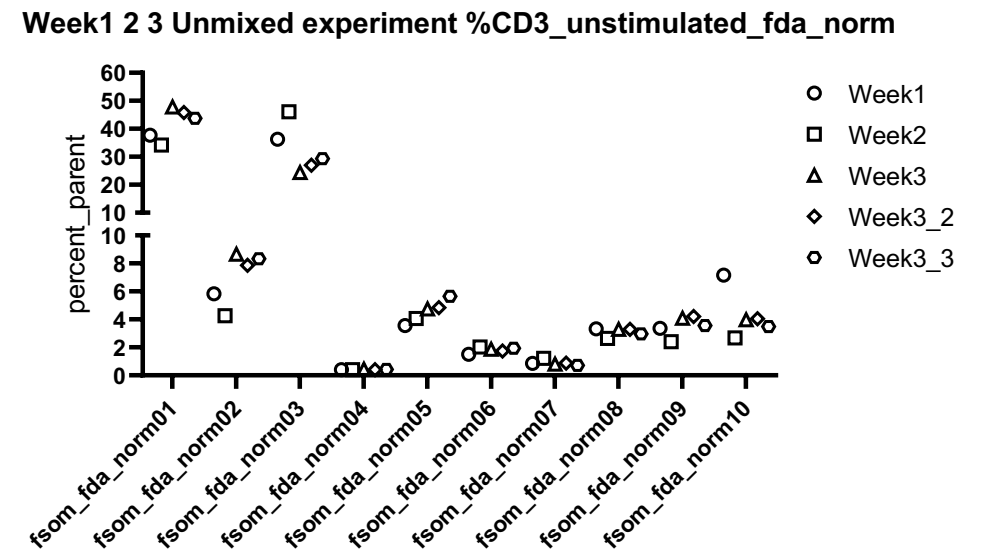
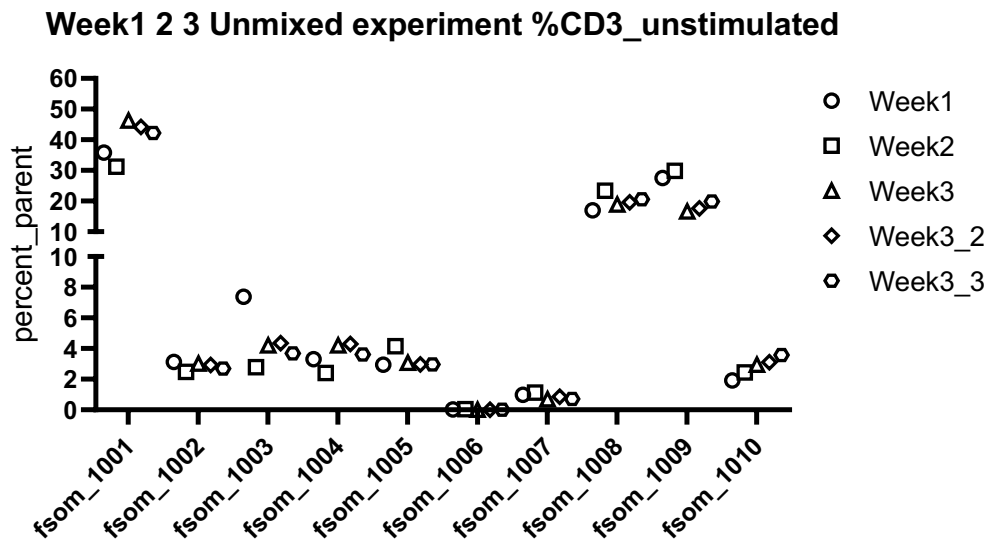


	Mean	%CV
CD27 MFI	1.674	35.2
CD8 MFI	1.4784	53.7
CD35 MFI	0.615	25.7
TCRab MFI	3.814	1.7
CD56 MFI	0.2668	38.3
CD38 MFI	0.4704	10.2
CD4 MFI	2.0702	74.6
CD25 MFI	0.1771	48.1
CD62L MFI	1.308	5.8



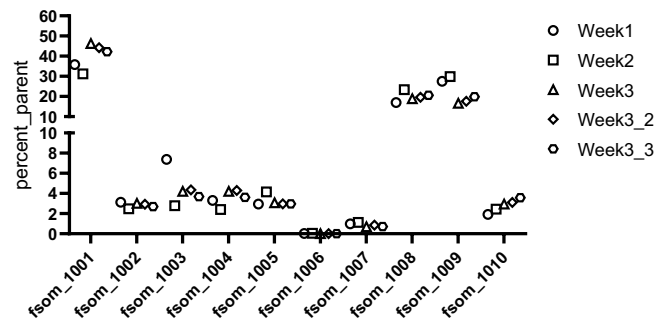
	not_normalized		fda_norm		gauss_norm		cytonorm1		cytonorm2	
	Mean	%CV	Mean	%CV	Mean	%CV	Mean	%CV	Mean	%CV
CD27 MFI	6147.8	74.9	1.646	37.4	1.712	35.5	1.646	38.2	1.674	35.2
CD8 MFI	2705	90.3	1.4028	49.9	1.416	50.5	1.453	50	1.4784	53.7
CD35 MFI	1295	35.8	0.6758	7.5	0.6728	6.4	0.63	19.3	0.615	25.7
TCRab MFI	47324	14.3	3.908	1.2	3.926	1.1	3.76	2.9	3.814	1.7
CD56 MFI	390.6	40.1	0.2642	67.7	0.1424	130.8	0.303	60.7	0.2668	38.3
CD38 MFI	980.4	15.1	0.447	17.8	0.4998	10.4	0.462	22.7	0.4704	10.2
CD4 MFI	21626	172.4	2.752	39.7	0.8396	180.1	2.122	72.8	2.0702	74.6
CD25 MFI	169.56	52	0.2368	10.9	0.2086	12.7	0.203	39.1	0.1771	48.1
CD62L MFI	853.4	7.8	1.2066	15.1	1.2014	15	1.368	10.6	1.308	5.8



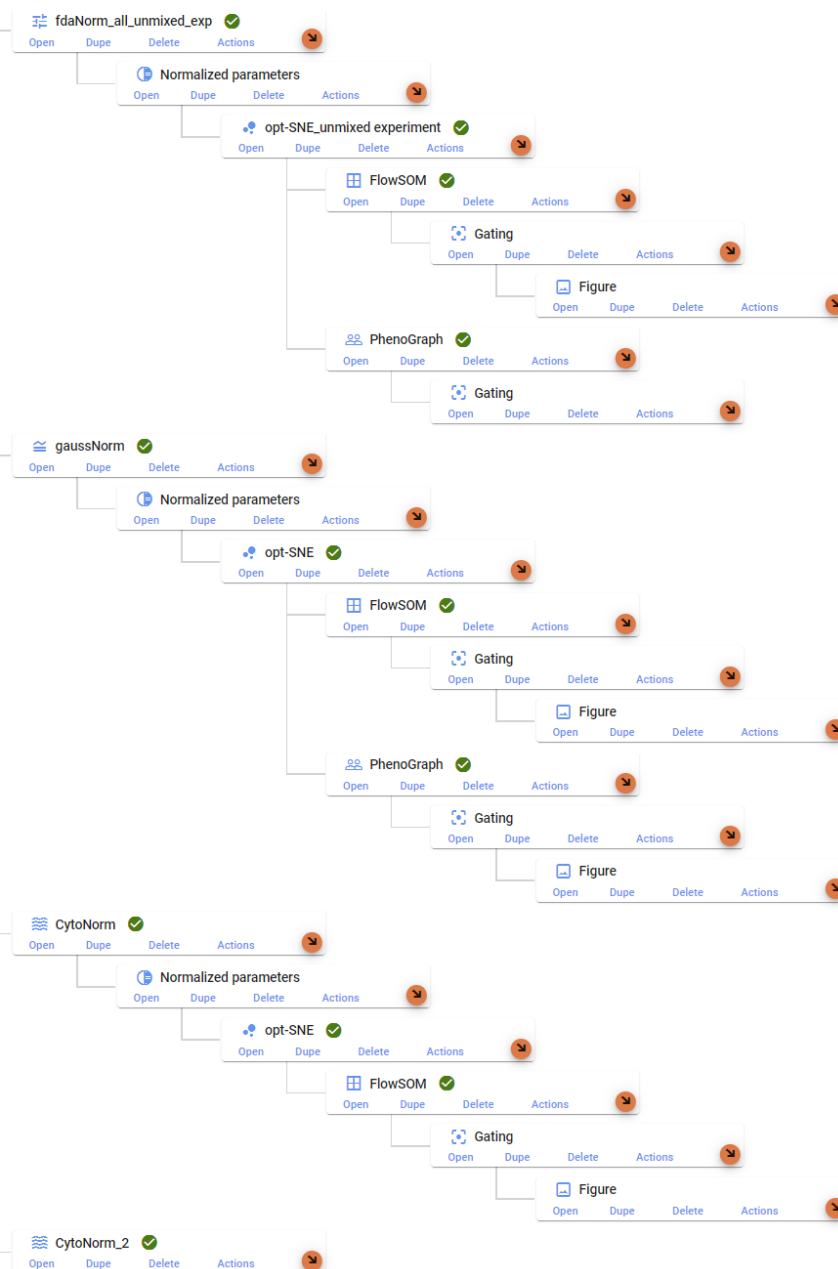
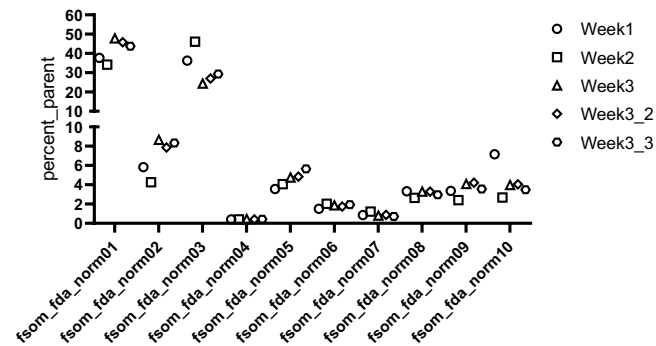


Normalization

Week1 2 3 Unmixed experiment %CD3_unstimulated



Week1 2 3 Unmixed experiment %CD3_unstimulated_fda_norm



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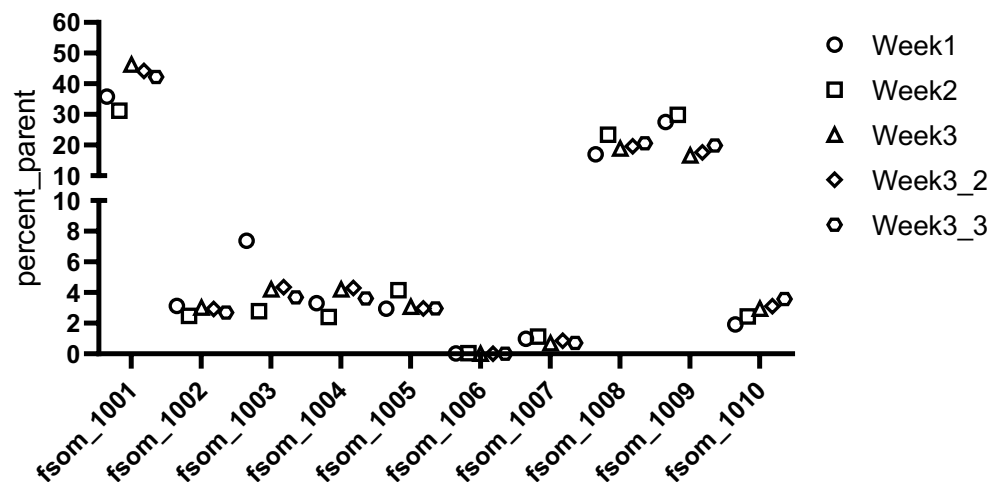


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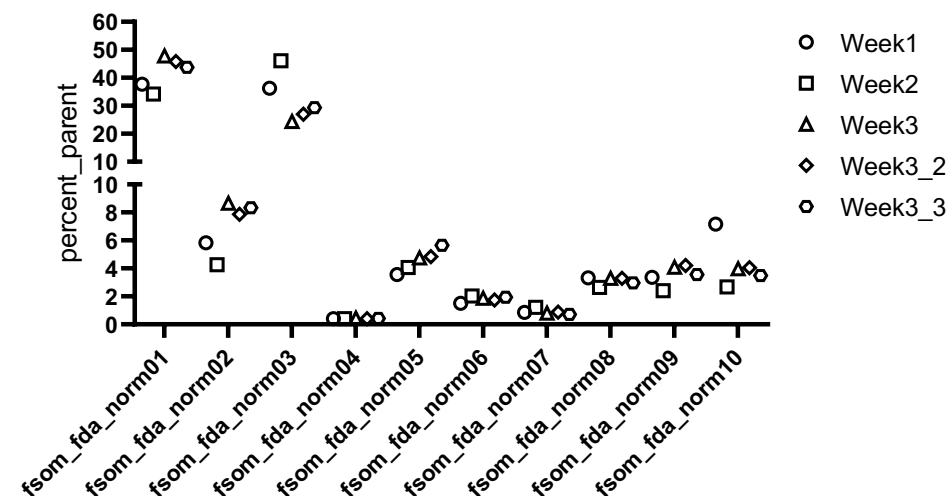


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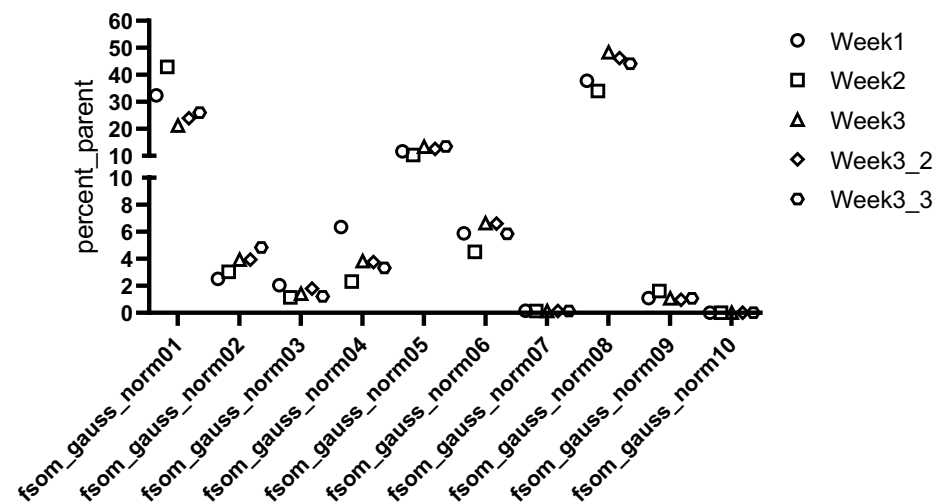
Week1 2 3 Unmixed experiment %CD3_unstimulated



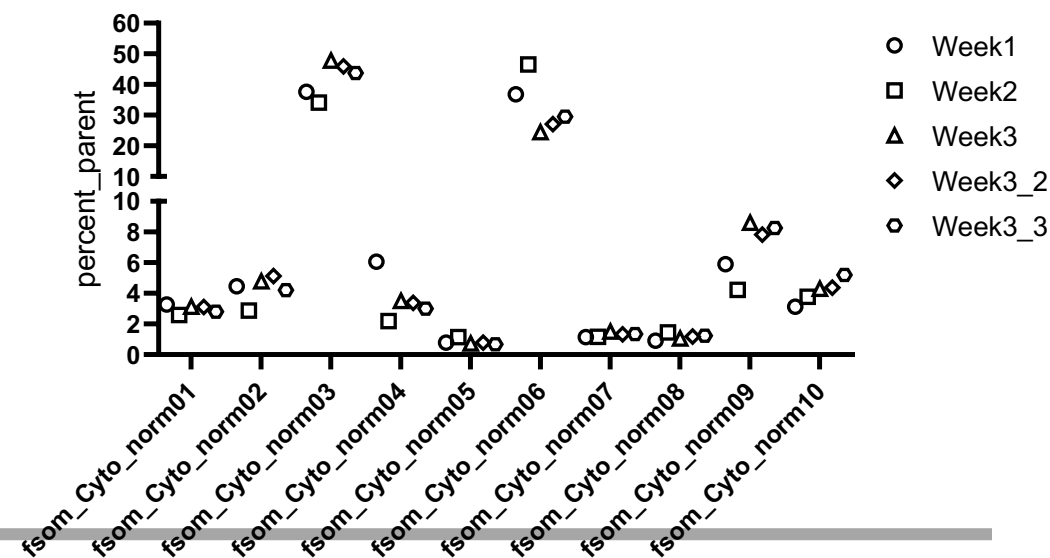
Week1 2 3 Unmixed experiment %CD3_unstimulated_fda_norm



Week1 2 3 Unmixed experiment %CD3_unstimulated_guauss_norm



Week1 2 3 Unmixed experiment %CD3_unstimulated_CytoNorm2







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